

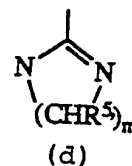
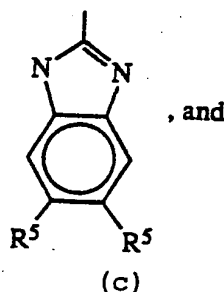
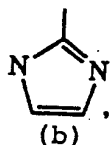
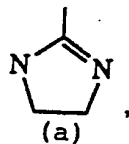
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(54) Title: PLATELET AGGREGATION INHIBITORS



(57) Abstract

An assay for screening snake venom for the presence or absence of platelet aggregation inhibitors (PAIs) based on specific receptor binding is described. Using this assay, the identification and characterization of PAIs in a wide range of snake venom samples was accomplished. The isolated and purified PAI from several of these active snake venoms is described and characterized. In addition, PAIs lacking the Arg-Gly-Asp (RGD) adhesion sequence but containing K*-(G/Sar)-D wherein K* is a modified lysyl residue of the formula $R^1_2 N(CH_2)_4 CHNHCO-$ wherein each R^1 is independently H, alkyl(1-6C) or at most one R^1 is $R^2-C=NR^3$ wherein R^2 is H, alkyl(1-6C), phenyl or benzyl, or is NR^4 in which each R^4 is independently H or alkyl(1-6C) and R^3 is H, alkyl(1-6C), phenyl or benzyl, or $R^2-C=NR^3$ is a radical selected from the group consisting of (a), (b), (c) and (d) where m is an integer of 2-3, and each R^5 is independently H or alkyl(1-6C); and wherein one or two (CH_2) may be replaced by O or S provided said O or S is not adjacent to another heteroatom are prepared and shown to specifically inhibit the binding of fibrinogen or von Willebrand Factor to GP IIb-IIIa.

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PLATELET AGGREGATION INHIBITORS10 Technical Field

This invention relates to a group of peptides which are, or are related to, platelet aggregation inhibitors isolated and purified from various snake venoms. These peptides are useful as therapeutic agents for the treatment of, and prevention of, platelet-associated ischemic disorders. More specifically, the invention concerns peptides which block specific receptors for adhesive proteins involved in platelet adherence and aggregation. Furthermore, this invention describes methods for detecting and purifying said polypeptides to substantial homogeneity from snake venoms, as well as processes for using the primary amino acid sequences of these polypeptides to prepare active peptides both synthetically and through use of recombinant DNA methods.

25

Background Art

Heart disease is the primary cause of death in most western societies. Death from heart disease is often induced by platelet-dependent ischemic syndromes which are initiated by atherosclerosis and arteriosclerosis and include, but are not limited to, acute myocardial infarction, chronic unstable angina, transient ischemic attacks and strokes, peripheral vascular disease, arterial thrombosis, preeclampsia, embolism, restenosis and/or thrombosis following angioplasty, carotid endarterectomy, anastomosis of vascular grafts, and chronic cardiovascular

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devices (e.g., in-dwelling catheters or shunts "extracorporeal circulating devices"). These syndromes represent a variety of stenotic and occlusive vascular disorders thought to be initiated by platelet activation either on vessel walls or within the lumen by blood-borne mediators but are manifested by platelet aggregates which form thrombi that restrict blood flow.

Numerous studies have contributed to an understanding of the mechanism of platelet aggregation and thrombus formation. Platelets respond to a variety of blood vessel injuries, such as narrowing of the lumen, plaque formation, and the presence of foreign bodies (e.g., catheters) and the like. The response of platelets to these injuries is a sequence of events including platelet adherence and activation, and the release of platelet granular components, including potent cellular mitogenic factors. The activated platelet aggregates induce the formation of fibrin, which further stabilizes the thrombus.

Much is now known about mechanisms regulating these responses. Although unstimulated platelets contain receptors for several adhesive proteins including laminin (VLA 2, VLA 6) and collagen (VLA 2, GPIV, others), the initial attachment of platelets to subendothelium is believed to be mediated by the binding of platelet membrane glycoprotein (GP) Ib to the immobilized von Willebrand factor. Subsequent platelet activation can be initiated by one or more of the known physiological agonists including: ADP, epinephrine, thrombin, collagen, and thromboxane A₂.

Platelet aggregation is mediated by GP IIb-IIIa complex on the platelet membrane surface. GP IIb-IIIa exists on the surface of unstimulated platelets in an inactive form. When platelets are activated by adhesion and the physiological agonists, the GP IIb-IIIa also becomes activated such that it becomes a receptor for fibrinogen

(Fg), von Willebrand Factor (vWF), and fibronectin (Fn) (see Phillips et al., Blood (1988) 71:831-843); however, it is the binding of fibrinogen and/ or von Willebrand factor that is believed to be principally responsible for platelet aggregation and thrombus formation in vivo. Therefore, substances which specifically inhibit the binding of fibrinogen or von Willebrand factor to GP IIb-IIIa inhibit platelet aggregation and could be candidates for inhibiting thrombus formation in vivo.

Platelet GP IIb-IIIa is now known to be a member of a superfamily of structurally related adhesive protein receptors known collectively as the "integrins." Like GP IIb-IIIa, all integrins known to date are two subunit molecules with a larger alpha-subunit (e.g., GP IIb) and a smaller beta-subunit (e.g., GP IIIa). There is a high degree of homology between the known sequences of the integrin subunits indicating that the integrins evolved from a common precursor. Integrins function in a variety of cellular adhesions and have been found in leucocytes, endothelial cells, smooth muscle cells and other cells in the vasculature. Because integrins are widely distributed, while GP IIb-IIIa is restricted to platelets, a preferred antiaggregating agent would selectively inhibit GP IIb-IIIa as opposed to other integrins.

Several classes of peptides have been disclosed which block the binding of adhesive proteins to activated platelets and inhibit platelet aggregation (see Hawiger et al., U.S. patent 4,661,471; and Rouslahti et al., U.S. patents 4,614,517; 4,578,079; 4,792,525; and UK application GB 2,207,922A). In one class of peptides, the sequence RGD is critical, and the tetrapeptide sequences RGDS, RGDT, RGDC, have been used specifically. The amino acid sequence RGD_X is found in a variety of adhesive proteins including Fg, Vn, vWF and Fn. This sequence has been demonstrated to play an important role in the interaction of adhesive proteins with adhesive protein recep-

tors because peptides containing this sequence block the binding of adhesive proteins. See, e.g., Pierschbacher, M.D., et al., J Biol Chem (1987) 262:17294-17298; Ruggeri et al., Proc Natl Acad Sci (USA) (1986) 83:5708-5712; and
5 Rouslahti et al., Cell (1986) 44:517-518. Tetrapeptides containing this sequence are disclosed in EP application 319,506 published 7 June 1989. Short peptides containing homoarginine instead of arginine in the RGD sequences are disclosed in PCT application WO89/07609 published 24
10 August 1989.

The structural variations permitted in RGD-containing peptides have been explored by Pierschbacher, M.D. et al. J Biol Chem (supra). In these studies, it was found that manipulating the RGD-containing sequence not
15 only affected the activity related to inhibition of binding of fibronectin or vitronectrin to substrate, but could also effect differentiation between binding of the two ligands. The peptide sequence GRGDSPC which was taken from the cell attachment domain of fibronectin was used as
20 a model peptide. Certain substitutions, such as replacement of L-Arg with D-Arg seem to have no effect on the binding of either ligand, but substituting D-Ala for Gly or D-Asp for L-Asp destroyed the inhibition activity. While substituting D-Ser for L-Ser reduced inhibition of
25 vitronectin interaction with vitronectin receptor, there was little effect on fibronectin interaction with fibronectin receptor; substitution of Asn for Ser resulted in a peptide that had enhanced inhibition of fibronectin binding, and a decreased effect on vitronectin binding. Alternate substitutions for Ser had other effects.
30 Threonine substituted for Ser gave a peptide with increased inhibition of binding to the vitronectin receptor; substitution of L-Pro led to an inactive peptide. A cyclic peptide was also prepared of the sequence Gly-Pen-Gly-Arg-Gly-Asp-Ser-Pro-Cys-Ala, wherein "Pen" is
35 penicillamine and a disulfide bridge was formed between

the Pen and Cys. In the view of the authors, penicillamine had the function of increasing conformational restraints on the ring whereas the N-terminal Gly and carboxy-terminal Ala were added to distance the free amino and carboxyl groups from the ring. This cyclic peptide was able to inhibit vitronectin binding more strongly than the same peptide before cyclization, but was ineffective in inhibiting fibronectin binding.

10 Recently, an antithrombotic peptide with a modification of the RGD sequence having the "R" residue alkylated was reported by Samanen, J., et al., J Cell Biochem (1990) Suppl 14A:A229. A review of structure/activity relationships in RGD-containing peptides has been published by Ali, F.E. et al. in Proc 11th Am Peptide Symp, Marshall et al., ed. ESCOM Leiden 1990.

15 European Patent Application publication no. 341,915 published 15 November 1989 discloses two groups of peptides, one linear and the other cyclic, which are said to bind the platelet GP IIb-IIIa receptor and thus to inhibit its ability to bind vWF, fibronectin and fibrinogen-fibrin. No data are provided which relate to the specificity of binding of these peptides. The group of cyclic peptides includes modifications of the RGD sequence wherein the R is substituted by D or L homoarginine, dimethyl or diethyl arginine, lysine, or an alpha-alkylated derivative of these residues. Minimal cyclic structures comprise simply the "R" GD sequence bracketed between the two residues which form the disulfide bridge.

20 A separate class of inhibitory peptides utilizes peptide sequences modeled on the carboxyl terminal sequence derived from the gamma chain of fibrinogen, the dodecapeptide HHLGGAQKAGDV (Kloczewiak et al., Biochemistry (1989) 28:2915-2919; Timmons et al., (Ibid), 2919-2923 US Patent 4,661,471 (supra); EP application

298,820,). Although this sequence inhibits Fg and vWF binding to GP IIb-IIIa and subsequent platelet aggregation, the usefulness of this peptide is limited because it has a low affinity of interaction with platelet receptors (IC₅₀=10-100 uM).

Recently, several groups have isolated and characterized a new class of low molecular weight polypeptide factors from snake venoms which have extremely high affinity for the GP IIb-IIIa complex. Huang, T.-F., et al., J Biol Chem (1987) 262:16157-16163; Huang, T.-F., et al., Biochemistry (1989) 28:661-666 report the primary structure of trigramin, a 72 amino acid peptide containing RGD and 6 disulfide bridges isolated from Trimeresurus gramineus. Gan, Z.-R., et al., J Biol Chem (1988) 263:19827-19832, report the properties and structure of echistatin, a 49 amino acid peptide also containing RGD and 4 putative disulfide bridges which is isolated from Echis carinatus. Williams, J.A., et al., FASEB Journal (1989) 3:A310, Abstr. No. 487m, report the sequence and properties of the related peptides elegantin, albolabrin, and flavoviridin. In addition, characterization of bitistatin was reported by Shebuski, R.J., et al., J Biol Chem (1989) 264:21550-21556; and the PAI from Agkistrodon piscivorus piscivorus was reported by Chao, B.H., et al., Proc Natl Acad Sci USA (1989) 86:8050-8054. The relationship between various GP IIb-IIIa antagonists from snake venoms was discussed by Dennis, M.S., et al., Proc Natl Acad Sci USA (1989) 87:2471-2475.

Included in this group of inhibitory peptides from snake venoms are alboabrin isolated from Trimeresurus albolabris, elegantin isolated from T. elegans, flavoviridin isolated from T. flavoviridis, batroxostatin isolated from Bothrops atrox, bitistatin isolated from Bitis arietans reported by Niewiarowski, S., et al., Thromb Haemostas (1989) 62:319 (Abstr. SY-XIV-5). In addition, applaggin has been purified from Agkistrodon p.

piscivorus and reported by Chao, B., et al., Thromb Haemostas (1989) 62:50 (Abstr. 120) and halysin, purified from Agkistrodon halys which was reported by Huang, T.F., et al., Thromb haemostas (1989) 62:48 (Abstr. 112). All
5 of these peptides show a high degree of sequence homology. In addition, all of the peptides reported to date from snake venoms which inhibit the binding of adhesive proteins to integrin receptors contain the RGD sequence.

Although these reported snake venom factors are
10 potent platelet aggregation inhibitors in vitro, these peptides also bind with high affinity to other members of the adhesive protein receptors such as the vitronectin and fibronectin receptors (Knudsen, K.A., et al., Exp Cell Res (1988) 179:42-49; Rucinski, B., et. al., Thromb Haemostas
15 (1989) 62:50 (Abstr. 120). This lack of specificity of snake venom factors for GP IIB-IIIa is an undesirable feature of their therapeutic use as inhibitors of thrombus formation, because they have the potential of affecting the adhesive properties of other cells in the vasculature,
20 particularly those adhesions mediated by integrins.

Another approach developed for the generation of platelet thrombus inhibitors has been the use of murine anti-GP IIB-IIIa monoclonal antibodies which block the binding of the adhesive proteins to stimulated platelets.
25 These monoclonal antibodies have been used to prevent coronary artery reocclusion after reperfusion with tissue plasminogen activator in dogs (Yasuda, T., et al., J Clin Invest (1988) 81:1284-1291) and to prevent cyclic reduction of flow in injured canine coronary arteries with a
30 high grade stenosis. Potential side effects of the use of such monoclonal antibodies in humans may result from their long-lasting effects and from their potential immunogenicity.

Clearly, additional therapeutic treatment
35 regimens are needed for preventing or at least mitigating undesirable thrombus formation. In particular,

therapeutic agents capable of blocking or inhibiting thrombus formation at specific locations without compromising hemostasis and without affecting other cellular adhesions, would provide major therapeutic benefits.

5 Ideally, these agents should be potent, specific for GP IIB-IIIA, and nonimmunogenic to most patients; they also should be easy to administer, stable and economical to produce. Further, these agents should act transiently and be capable of functioning at the earliest stages of

10 thrombus formation, without interfering with long-term hemostasis. The present invention fills these and other related needs.

Disclosure of the Invention

15 The invention provides a simple screening procedure to identify low molecular weight (<10kd) factors in snake venom or other biological sources that specifically inhibit thrombus formation mediated by platelet aggregation. This procedure takes advantage of

20 the understanding that platelet aggregation is primarily effected through binding of fibrinogen and/or vWF to GP IIB-IIIA at the surface of platelets when the platelets are treated with appropriate stimuli, such as ADP. By using these criteria, i.e., inhibition of binding of

25 fibrinogen and/or vWF to isolated receptor and analogous criteria related to inhibition of binding of fibronectin (Fn) to fibronectin receptor (Fn/FnR binding) and vitronectin to vitronectin receptor (Vn/VnR binding), as well as the binding of other factors, such as Fn and Vn to

30 GP IIB-IIIA, a specificity profile for the platelet aggregation inhibitor (PAI) can be rapidly and conveniently obtained. This approach has been used to screen and characterize an extensive panel of snake venoms for the presence or absence of PAI, to characterize the specific-

35 ity of PAI identified from this panel for their specificity in inhibiting binding to GP IIB-IIIA as opposed to

inhibiting other integrins, and to identify active peptides which are derivatives of these PAIs.

Accordingly, in one aspect, the invention is directed to a rapid screening method for the presence or
5 absence of PAI in a biological fluid, which method comprises contacting the fluid with isolated GP IIB-IIIa in a test reaction in the presence of fibrinogen and comparing the amount of fibrinogen bound to GP IIB-IIIa in
10 this test reaction with the amount of fibrinogen bound to GP IIB-IIIa in a control reaction. The method may further include test and control reactions which involve contact-
ing Fn with Fn receptor, Vn with Vn receptor, Fn with GP IIB-IIIa, or vWF with GP IIB-IIIa to characterize the
specificity of the PAI.

15 In another aspect, the invention is directed to novel PAI in isolated form which is identified in, and can be isolated from, active snake venom according to the methods of the invention. In particular, the invention
relates to PAI, in isolated form, which can be isolated
20 from Echis colorata, Eristicophis macmahonii; A. hypnale, A. acutus, A. piscivorus leucostoma, A. piscivorus conanti; Bothrops asper; Bothrops cotiara, B. jararaca, B. jararacussu, B. lansbergi, B. medusa, B. nasuta, B. neuwiedi, B. pradoi, B. schlegli; Crotalus atrox, C.
25 basilicus, C. cerastes cerastes, C. durissus durissus, C. durissus totonatacus, C. horridus horridus, C. molossus molossus, C. ruber ruber, C. viridis cereberus, Crotalus v. helleri, Crotalus v. lutosus, Crotalus v. oreganus, Crotalus v. viridis; Lachesis mutas; Sistrurus catenatus
30 tergeminus, and Sistrurus milarus barbouri.

Preferred are PAIs in isolated form prepared from, or having the amino acid sequences of, those
obtained from Eristicophis macmahonii (eristicophin);
Bothrops cotiara (cotiarin); B. jararacussu; Crotalus
35 atrox (crotatroxin); Crotalus basilicus (basilicin); C. cerastes cerastes (cerastin); C. durissus totanatacus

(durissin); C. durissus durissus (durissin); C. h. horridus (horridin); Crotalus m. molossus (molossin); C. ruber ruber (ruberin); Crotalus viridis lutosus (lutosin); C. v. viridis (viridin); Crotalus v. oreganus (oreganin);
5 Crotalus v. helleri; Lachesis mutas (lachesin); Sistrurus catenatus tergeminus (tergeminin); and S. milarus barbouri (barbourin).

Especially preferred are eristicophin, cotiarin, crotatroxin, cerastin, durissin, horridin, ruberin,
10 lachesin, basilicin, lutosin, molossin, oreganin, viridin, tergeminin and barbourin.

The invention also includes peptides of the amino acid sequences as described above which are truncated and/or modified forms of the naturally occurring
15 peptides and/or have one or more peptide linkages replaced by alternate linkages such as $-CH_2NH-$ or $-CH_2CH_2-$.

In a preferred aspect, the invention relates to PAI in isolated form which can be prepared from active snake venom identified by the method of the invention, and
20 shown to specifically inhibit the binding fibrinogen (Fg) and/or von Willebrand Factor (vWF) to GP IIb-IIIa, and their truncated and/or modified forms.

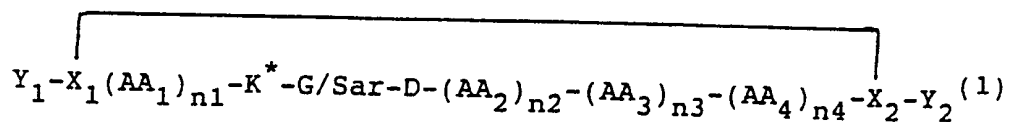
In still another preferred aspect, the invention relates to PAI of snake venom in isolated form wherein the
25 sequence responsible for binding to the adhesive protein receptor includes the sequence KGD.

In another major aspect, the invention is directed to a group of peptides or peptide-related compounds in general which are platelet aggregation
30 inhibitors that are capable of inhibiting binding of Fg or vWF to GP IIb-IIIa at a substantially higher potency than that at which they inhibit binding of vitronectin to vitronectin receptor or fibronectin to fibronectin receptor. These peptides are characterized by having the binding
35 sequence K^*GDX in place of the $RGDX$ binding sequence which is found in the prior art PAI proteins. K^* is a

- substituted or unsubstituted lysyl residue of the formula $R^1_2 N(CH_2)_4 CHNHCO-$ wherein each R^1 is independently H or a substituent which is sufficiently electron donating so as not to destroy the basicity of the adjacent nitrogen, and
- 5 wherein one or two of the methylene residues may optionally be substituted by O or S, as described below. The barbourin PAI isolated from S. milarus barbouri is one illustration of this series of peptides. However, shorter forms of this peptide can also be used, as well as
- 10 analogous sequences which also contain 1-10 amino acid residue modifications elsewhere in the peptide chain, and/or replacement of peptide linkages with alternate linkages. Other illustrative embodiments include isolated PAI peptides having a native RGD X sequence wherein this is
- 15 replaced by K X GD X . As in the case of barbourin, these isolated PAI may be otherwise in native form, or may be truncated and/or may contain 1-10 amino acid residue substitutions or deletions, and/or may have non-peptide linkages substituted for peptide linkages.
- 20 Another group of compounds which falls within the scope of the invention is that wherein the foregoing compounds are as described, except that the glycyl residue in the RGD or K X GD sequence is replaced by a sarcosyl residue. This class of compounds retains the potency and
- 25 specificity of the related RGD or K X GD-containing peptides.

Another illustrative group of embodiments are peptides or modified peptides having specific PAI activity

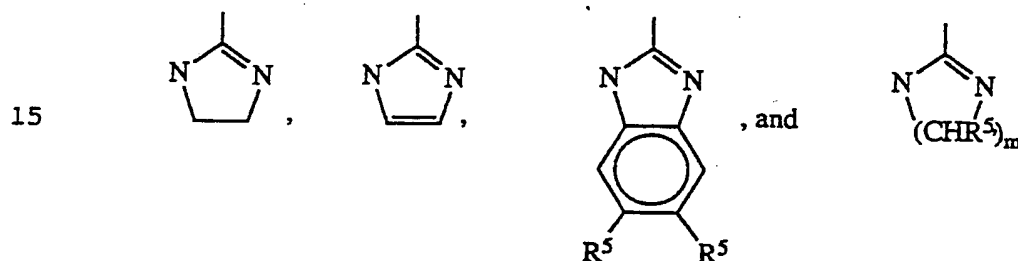
30 of the formula



wherein K^* is a substituted or unsubstituted lysyl residue of the formula $R^1_2N(CH_2)_4CHNHCO-$ as described above,

wherein each R^1 is independently H, alkyl (1-6C), or at most one R^1 is $R^2-C=NR^3$,

wherein R^2 is H, alkyl(1-6C) or is a substituted or unsubstituted phenyl or benzyl residue, or is NR^4_2 in which each R^4 is independently H or alkyl(1-6C), and R^3 is H, alkyl(1-6C), phenyl or benzyl, or $R^2-C=NR^3$ is a radical selected from the group consisting of:



20 where m is an integer of 2-3, and each R^5 is independently H or alkyl(1-6C);

and wherein one or two (CH_2) may be replaced by O or S provided said O or S is not adjacent to another heteroatom;

25 AA_1 is a small, neutral (polar or nonpolar) amino acid and n_1 is an integer of 0-3;

AA_2 is a neutral, nonpolar large (aromatic or nonaromatic) or a polar aromatic amino acid and n_2 is an integer of 0-3;

30 AA_3 is a proline residue or a modified proline residue (as defined below) and n_3 is an integer of 0-1;

AA_4 is a neutral, small amino acid or the N-alkylated form thereof and n_4 is an integer of 0-3;

35 each of X_1 and X_2 is independently a residue capable of forming a bond between X_1 and X_2 to obtain a cyclic compound as shown; and

each of Y_1 and Y_2 is independently a noninterfering substituent or may be absent;

wherein one or more peptide linkages may optionally be replaced by a linkage selected from the group consisting of $-\text{CH}_2\text{NH}-$, $-\text{CH}_2\text{S}-$, CH_2CH_2- , $-\text{CH}=\text{CH}-$ (cis and trans), $-\text{COCH}_2-$, $-\text{CH}(\text{OH})\text{CH}_2-$ and $-\text{CH}_2\text{SO}-$;

with the proviso that if n_3 is 0; either:

- 1) the sum of n_2 and n_4 must be at least 2; or
- 2) K^* must be other than Har or K; or
- 3) at least one of X_1 and X_2 must be other than cys (C), penicillamine (Pen), or 2-amino-3,3-cyclopentanemethylene-3-mercaptopropionic acid (APmp); or
- 4) Y_1 or Y_2 must comprise at least one amino acid residue; or
- 5) one or more peptide linkages is replaced by said alternate linkage.

Other aspects of the invention are concerned with recombinant methods and materials related to the synthesis of these and other related peptides, to methods of in vitro synthesis thereof, to pharmaceutical compositions containing these compounds, and to methods to inhibit platelet aggregation and thrombus formation using these compounds and compositions.

25 Brief Description of the Drawings

Figure 1 shows inhibition of the binding of fibrinogen to GP IIb-IIIa by partially purified snake venoms.

Figure 2 shows the dose-response adhesion inhibition of Centricon-10 ultrafiltrates of crude venoms in both fibrinogen/GP IIb-IIIa and vitronectin/vitronectin receptor assays.

Figure 3 shows the HPLC profile of crude PAI from Eristicophis macmahoni venom. The shaded area contains the biologically active fractions.

Figure 4 shows the HPLC profile of PAI fractions from Figure 3. The shaded area contains the bioactive fractions.

Figure 5 shows the analytical HPLC profile of PAI fractions from Figure 4.

Figure 6 shows the complete amino acid sequences of eristicophin, barbourin, tergeminin, cerastin, ruberin, lachesin, cotiarin, crotatroxin, horridin, lutosin, viridin, molossin, basilicin, durissin, jararacin, cereberin, and oreganin, and enzyme digestion fragments determined by automated Edman degradation.

Figure 7 depicts the HPLC profile of PAI obtained from G-50 fractions of crude Sistrurus c. tergeminus venom.

Figure 8 depicts the HPLC profile of PAI fractions from Figure 7.

Figure 9 shows the activity of the purified PAI of Figure 8 in inhibiting binding in several receptor assays.

Figure 10 depicts the HPLC profile of platelet aggregation inhibitor obtained from G-50 fractions of crude Sistrurus m. barbouri venom. The shaded areas contain the bioactive fractions.

Figure 11 depicts the HPLC profile of active PAI fractions of Figure 10.

Figure 12 compares the amino acid sequences of a number of PAIs to that of barbourin.

Figure 13 depicts the HPLC profile of crude PAI from Lachesis mutas venom. Shaded areas contain the biologically active fractions.

Figure 14 depicts the HPLC profile of the PAI active fractions from Figure 13. Shaded area contains the biologically active fractions.

Figure 15 depicts the analytical HPLC profile of PAI fractions of Figure 14 from Lachesis mutas.

Figure 16 depicts the HPLC profile of crude PAI from Crotalus viridis viridis venom. Shaded area contains the biologically active fractions.

Figure 17 depicts the HPLC profile of the PAI
5 fractions of Figure 16.

Figure 18 shows the dose-response effects of purified snake venom peptides to inhibit fibrinogen/GP IIb-IIIa binding as compared to echistatin.

Figure 19 shows the dose-response effects of
10 purified snake venom peptides to inhibit ADP (4 μ M) induced human platelet aggregation in platelet rich plasma (PRP), as compared to echistatin.

Figure 20 shows the activity profile from HPLC fractionation of C. c. cerastes venom.

Figure 21 shows the results of HPLC analysis of
15 the active fractions of Figure 20.

Figure 22 shows the activity profile of HPLC fractionation of PAI from C. ruber ruber.

Figure 23 shows the activity profile of an
20 analytical C-18 column on homogeneous peptide obtained from C. atrox.

Figure 24 shows the analytical HPLC profile of the homogeneous peptide isolated from Bothrops cotiara.

Figure 25 shows the dose-response effects of
25 purified cotiarin on inhibiting the binding of fibrinogen to GP IIb-IIIa and inhibition of the binding of vitronectin to the vitronectin receptor.

Figure 26 shows the effects of purified snake
30 venom peptides on binding of fibrinogen to GP IIb-IIIa and vitronectin to the vitronectin receptor.

Figure 27 shows the results of binding activity for analog #1, [$E^{28}L^{41}C^{64}$]barbourin(28-73), with regard to GP IIb-IIIa and vitronectin receptor.

Figure 28 shows the ability of synthetic
35 eristicophin analog to inhibit the binding of fibrinogen

to GP IIb-IIIa and inability to inhibit the binding of vitronectin to the vitronectin receptor.

Figure 29 shows the ability of linear and cyclic RGDW compounds and linear and cyclic KGDW compounds to
5 inhibit the binding of fibrinogen to GP IIb-IIIa.

Figure 30 shows the ability of various KGDW analogs to inhibit binding of fibrinogen to GP IIb-IIIa and inhibit binding of vitronectin to vitronectin receptor.

10 Figure 31 shows the ability of various native and synthetic platelet aggregation inhibitors to inhibit the attachment of M21 melanoma cells to vitronectin.

Figure 32 shows the ability of RGDS and a cyclic RGD compound to inhibit the attachment of M21 melanoma
15 cells to vitronectin and the lack of ability of a cyclic KGDW analog to inhibit the attachment of M21 melanoma cells to vitronectin.

Figure 33 shows the activity of analog number 60, Mpr-(Har)-G-D-W-P-C-NH₂, in inhibiting aggregation of
20 platelets and cell adhesion to vitronectin.

Figure 34 shows the activities of Figure 33 for analog 19, Mpr-K-G-D-W-P-C-NH₂.

Figure 35 shows the initiation of cyclic flow reductions (CFRs) in an open chest dog model of thrombosis
25 (Folts Model).

Figure 36 shows the effects of a 10 mg bolus dose administration on the CFRs initiated in the open chest dog model of thrombosis (Folts Model).

Figure 37 shows the effects of a 40 mg bolus
30 dose administration on the CFRs initiated in the open chest dog model of thrombosis (Folts Model).

Figure 38 shows the full-length DNA sequence encoding the amino acid sequence of barbourin(1-73).

Figure 39 shows the DNA sequence encoding
35 [M-1L41]barbourin(1-73) ligated to a PhoA leader sequence.

Figure 40 shows the DNA sequence encoding analog #1 linked to a PhoA leader sequence for expression in bacteria.

Figure 41 shows oligonucleotides utilized in a PCR reaction to obtain DNA encoding analog #1. The amino acids included in the analog per se are shown in boldface type.

Figure 42 shows the junction sequence of tandem repeats of the analog #1-encoding DNA.

Figure 43 shows a diagram of the truncated barbourin gene as tandem repeats.

Modes of Carrying Out the Invention

The invention provides platelet aggregation inhibitors (PAI) which may be isolated from snake venom which has been identified as active by the assay methods of the invention and compounds which have similar structures and are synthesized using standard in vitro techniques, such as solid phase peptide synthesis, or using recombinant methods, or combinations of these. Some of these inhibitors are uniquely specific for inhibition of platelet aggregation and do not inhibit alternate binding within the integrin family. Others have different ranges of specificity. The sections below describe the isolation of naturally occurring PAI from snake venom; the design of inhibitors which are of substantially higher potency in inhibiting platelet aggregation than in inhibiting, for example, vitronectin/vitronectin receptor interaction by incorporating a K^{*}GD sequence in preference to RGD; methods of synthesizing these peptides; methods of recombinant production; antibodies raised against the invention peptides; the assay method which permits identification of snake venoms which contain PAI; and the administration and utility of the PAI of the invention.

By PAI is meant a factor which is capable of preventing the aggregation of stimulated platelets in

standard assays, for example those described by Gan, Z.-R., et al., and Huang, T.-F., et al., (supra). In these assays, washed platelets are combined with fibrinogen, Ca^{+2} and the material to be tested. The platelets are stimulated with ADP (or other known stimulators or combinations thereof) and aggregation (or lack thereof) is observed using, for example, a commercially available aggregometer.

Some of the PAIs of the invention are identified as specific for the inhibition of binding of fibrinogen and/or vWF to GP IIb-IIIa. It is understood that specificity is a matter of degree; therefore, a PAI "specific for inhibition of Fg or vWF binding to GP IIb-IIIa inhibits this binding substantially more than it inhibits the binding of Fn to FnR, or Vn to VnR. By "substantially more" is meant that either the % inhibition is at least twofold greater at a given concentration of PAI or that the concentration of PAI that causes 50% inhibition is at least twofold less for Fg or vWF/GP IIb-IIIa binding inhibition than for alternate ligand/receptor binding.

Isolated Native PAI and Purification Methods

The platelet aggregation inhibitors (PAI) of the invention include low molecular weight peptides which can be prepared in isolated form, as described below, from snake venom which has been identified as "active", i.e., has been found to contain PAI using the method of the invention, which is described hereinbelow.

The invention method permits ready identification and characterization of the presence of an effective PAI in snake venom which selectively inhibits binding to GP IIb-IIIa as opposed to other integrins as, for example, the vitronectin receptor and the fibronectin receptor. Upon such identification, and, optionally and optimally, characterization, the PAI can be isolated and purified

using a variety of standard techniques illustrated herein and disclosed in the art. For example, a combination of separation based on molecular weight (typically recovery of substances of <10kd), ion exchange chromatography, and
5 reverse phase HPLC can be used. Other techniques can also be employed, but a workable procedure applicable to PAI from any active snake venom is as follows:

About 10-1000 mg venom is dissolved in dilute acetic acid and applied to a sizing column, such as
10 Sephadex G-50, and eluted in the same solvent. Fractions are assayed for activity using the Fg/GP IIb-IIIa binding assay of the invention, a standard platelet aggregation assay (PAA) or any similar assay relying on the adhesive protein binding activity of GP IIb-IIIa. Alternatively,
15 the <10kd fraction of the fraction of the venom can be recovered using ultrafiltration and similarly assayed.

The low MW fraction isolated by either procedure is then loaded onto a preparative C-18 HPLC column, such as a C-18 Delta Pak reverse phase HPLC column, available
20 from Waters, preequilibrated in 0.1% trifluoroacetic acid (TFA)/8% acetonitrile. The adsorbed PAI is then eluted using a gradient of 8%-60% acetonitrile in 0.1% TFA. The slope of the gradient and flow rate are optimized using routine procedures. Active fractions are determined by
25 PAA or by the invention receptor binding method. The active fractions are then pooled, concentrated, and tested for homogeneity using analytical HPLC or SDS-PAGE. Further reverse-phase HPLC gradient purification is applied until the recovered PAI is homogenous.

30 PAIs of the invention, obtainable by the foregoing or other purification methods include those from venoms selected from the group consisting of Echis colorata, Eristicophis macmahonii; A. hypnale, A. acutus, A. piscivorus leucostoma, A. piscivorus conanti; Bothrops asper; Bothrops cotiara, B. jararaca, B. jararacussu, B. lansbergi, B. medusa, B. nasuta, B. neuwiedi, B. pradoi,

B. schlegli; Crotalus atrox, C. basilicus, C. cerastes
cerastes, C. durissus durissus, C. durissus totonatacus,
C. horridus horridus, C. molossus molossus, C. ruber
ruber, C. viridis cereberus, Crotalus v. helleri, Crotalus
5 v. lutosus, Crotalus v. oreganus, Crotalus v. viridis;
Lachesis mutas; Sistrurus catenatus tergeminus, and
Sistrurus milarus barbouri.

Preferred are PAIs in isolated form prepared
from, or having the amino acid sequences of, those
10 obtained from Eristicophis macmahonii (eristicophin);
Bothrops cotiara (cotiarin); B. jararacussu; Crotalus
atrox (crotatroxin); Crotalus basilicus (basilicin); C.
cerastes cerastes (cerastin); C. durissus totonatacus
(durissin); Crotalus d. durissus (durissin); C. h.
15 horridus (horridin); Crotalus m. molossus (molossin); C.
ruber ruber (ruberin); Crotalus viridis lutosus (lutosin);
C. v. viridis (viridin); Crotalus v. oreganus (oreganin);
Crotalus v. helleri; Lachesis mutas (lachesin); Sistrurus
catenatus tergeminus (tergeminin); and S. milarus barbouri
20 (barbourin). Particularly preferred are PAI specific for
inhibiting Fg or vWF/GP IIb-IIIa binding, e.g., that from
Sistrurus m. barbouri.

Especially preferred are eristicophin; cotiarin,
crotatroxin, cerastin, durissin, horridin, ruberin,
25 lachesin, basilicin, lutosin, molossin, oreganin, viridin,
tergeminin and barbourin.

The purified PAI of the invention can be
sequenced using standard procedures, thus permitting
synthesis using standard solid phase techniques (in
30 particular for shorter forms of the PAI) or recombinant
production. For example, an Applied Biosystems Sequenator
can be used following carboxyamido methylation or
pyridylethylation of the peptide as described by Huang et
al., J Biol Chem (1987) 262:16157-16163 followed by
35 desalting of the sample on a C-18 Delta Pak column using
0.1% TFA and acetonitrile.

It is understood that the isolated PAI of determined sequence can, when synthesized in vitro, be modified by sequence alterations which do not destroy activity. In general, these modified forms will differ from the native forms by 1-10, preferably 1-4, amino acid substitutions or will be truncated forms. In addition, one or more peptide linkages may be replaced by alternate linkages as described hereinbelow. A particularly preferred substitution is replacement of RGD by K*GD to confer GP IIB-IIIa specificity as described below.

The PAI of Sistrurus m. barbouri has been purified to homogeneity and sequenced, and termed "barbourin". Unlike the adhesive proteins for GP IIB-IIIa so far identified and the peptides from snake venoms that block GP IIB-IIIa function, barbourin does not contain the standard Arg-Gly-Asp sequence of the adhesive proteins known in the art. The apparent binding sequence in barbourin is Lys-Gly-Asp-(Trp). The presence of the KGD sequence in the apparent binding region of this peptide is especially surprising in view of the observation that replacement of Lys for Arg in small synthetic peptides based on the RDGX sequence greatly decreases the ability of these peptides to bind to integrin receptors (Pierschbacher et al., Proc Natl Acad Sci (USA) (1984) 81:5985-5988; Williams et al., Thromb Res (1987) 46:457-471); Huang et al., J.Biol Chem (1987) 262:16157-16163. It is thought that this substitution may in part be responsible for the specificity of the barbourin peptide to inhibit Fg and vWF binding to GP IIB-IIIa, versus, for example, inhibition of vitronectin binding to the vitronectin receptor.

K*GDH-Containing Peptides

The "barbourin" peptide isolated by the method of the invention has been shown to have the binding sequence KGDH in contrast to the RGDH found in the PAI

compounds of the prior art. The presence of the KGDX in this PAI sequence appears to be associated with a preferential affinity for GP IIB-IIIa as opposed to the vitronectin or fibronectin receptors. The effect of the substitution of a lysyl residue for an arginine in the sequence appears to be associated with increased length of the sidechain along with retained basicity of the nitrogen as is further described hereinbelow. Surprisingly, it appears that it is not the lysyl residue per se which accounts for the enhanced activity and specificity, but rather the spacing provided by this homologous extension of the replaced arginine. Thus, the peptides of the invention which contain K*GDX in the binding sequence are substantially more potent in inhibiting the binding of Fg or vWF to GP IIB-IIIa as compared to their ability to inhibit the binding of vitronectin to the vitronectin receptor and the binding of fibronectin to the fibronectin receptor. As stated above, by "substantially more" potent in inhibiting the preferred binding is meant that the percent inhibition is at least 2-fold greater at a set concentration of inhibitor or that the concentration of PAI that causes 50% inhibition is at least 2-fold less for the binding of Fg or vWF to GP IIB-IIIa than for the binding of alternate ligands to other integrins.

As used herein K* refers to a lysyl residue which is unsubstituted, or which contains substitutions for the hydrogens on the epsilon amino group. The substituents must be sufficiently electron donating so as to maintain the basicity of the nitrogen to which they are attached. Thus, K* is defined as a lysyl residue of the formula $R^1_2 N(CH_2)_4 CHNHCO-$,

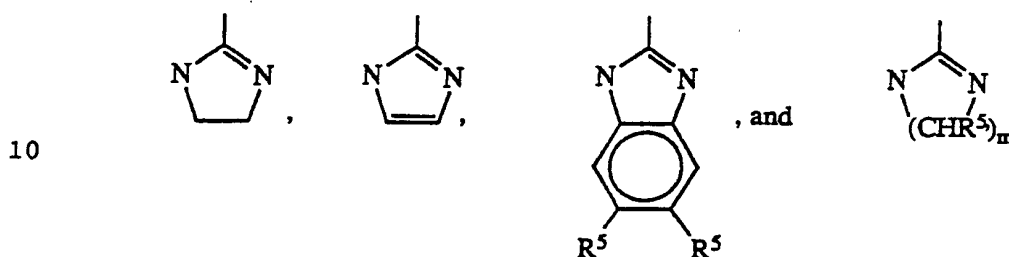
wherein each R^1 is independently H, alkyl (1-6) or at most one R^1 is $R^2-C=NR^3$,

wherein R^2 is H, alkyl(1-6C), or is a substituted or unsubstituted phenyl or benzyl residue, or

is NR^4_2 in which each R^4 is independently H or alkyl(1-6C), and

R^3 is H, alkyl(1-6C), phenyl or benzyl, or

$\text{R}^2\text{-C=NR}^3$ is a radical selected from the group
5 consisting of:



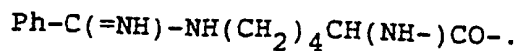
where m is an integer of 2-3, and each R^5 is
15 independently H or alkyl(1-6C);

and wherein one or two (CH_2) may be replaced by O or S provided said O or S is not adjacent to another heteroatom.

"Alkyl" is conventionally defined as a straight
20 or branched chain or cyclic hydrocarbyl residue of the indicated number of carbon atoms such as methyl, ethyl, isopropyl, N-hexyl, 2-methylbutyl, cyclohexyl and the like.

The benzyl and phenyl residues represented by R^2
25 may be unsubstituted, or may be substituted by noninterfering substituents. Preferred substitution patterns are those wherein only one substituent is bound to the aromatic nucleus, preferably in the 4-position. Preferred substituents are electron donating substituents
30 such as alkyl, especially ethyl or methyl, or phenyl.

Preferred embodiments of K^* include the residues of lysine, homoarginine, formylhomoarginine, ornithine, acetimidyl lysine, $\text{N}^{\text{G}}\text{N}^{\text{G}}$ ethylene-homoarginine, and phenylimidyl lysine. The phenylimidyl lysyl residue, for
35 example, has the formula:



As the essential feature of the preferential inhibition of binding appears to reside in the substitution of K* for R of RGD_X, one class of peptides or peptide-related compounds of the invention comprises naturally occurring platelet aggregation inhibitors which ordinarily contain RGD_X in the binding sequence whereby these forms are modified by substituting K* for R in this sequence. Included in the invention are the native peptides having this substitution, as well as their fragments of sufficient length to be effective in selectively inhibiting the binding of adhesive proteins to GP IIb-IIIa and fragments or full-length peptides which have irrelevant substitutions in positions of the peptide which do not destroy this activity. For the most part, the fragments will contain residues corresponding to the length of a peptide chain of at least 7 amino acids if the conformation is controlled by, for example, cyclization, and are of greater length if there is no such conformational control. In general, aside from the K*GD_X required sequence, there may be 1-10, preferably 1-4, and more preferably 1-3 amino acid substitutions in the non-K*GD_X portion of the peptides.

Additionally, the G of RGD_X or K*GD_X may be replaced by a sarcosine residue.

In addition, one or more of the peptide bonds may be optionally replaced by substitute linkages such as those obtained by reduction or elimination. Thus, one or more of the -CONH- peptide linkages can be replaced with other types of linkages such as -CH₂NH-, -CH₂S-, CH₂CH₂-, -CH=CH- (cis and trans), -COCH₂-, -CH(OH)CH₂- and -CH₂SO-, by methods known in the art. The following references describe preparation of peptide analogs which include these alternative-linking moieties: Spatola, A.F., Vega Data (March 1983), Vol. 1, Issue 3, "Peptide Backbone

- Modifications" (general review); Spatola, A.F. in "Chemistry and Biochemistry of Amino Acids, Peptides and Proteins," B. Weinstein, eds., Marcel Dekker, New York, p. 267 (1983) (general review); Morley, J.S., Trends Pharm Sci (1980) pp. 463-468 (general review); Hudson, D. et al. Int J Pept Prot Res (1979) 14:177-185 (-CH₂NH-, CH₂CH₂-); Spatola, A.F. et al., Life Sci (1986) 38:1243-1249 (-CH₂-S); Hann, M.M. J Chem Soc Perkin Trans I (1982) 307-314 (-CH-CH-, cis and trans); Almquist, R.G., et al., J Med Chem (1980) 23:1392-1398 (-COCH₂-); Jennings-White, C. et al. Tetrahedron Lett (1982) 23:2533 (-COCH₂-); Szelke, M., et al., European Appln. EP 45665 (1982) CA: 97:39405 (1982) (-CH(OH)CH₂-); Holladay, M.W. et al. Tetrahedron Lett (1983) 24:4401-4404 (-C(OH)CH₂-); and Hruby, V.J. Life Sci (1982) 31:189-199 (-CH₂-S-). Particularly preferred is -CH₂NH-.

Examples of fragments and/or modified forms of the naturally-occurring snake venom PAI include [E²⁸, L⁴¹, C⁶⁴]barbourin(28-73) of the sequence

20

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1
ECADGLCCDQCRFLKKGTVCRVAKGDWNDDTCTGQSCDCPRNGLYG 46
28                                                    73

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and [K²⁹]eristicophin(4-51) of the sequence

25

```

4
EEPCATGPCCRRCKFKRAGKVCRVAKGDWNNDYCTGKSCDCPRNPWNG 51
4                                                    51

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- In this notation, the size of the fragment is noted in parentheses after the name by the numbers of the amino acids which are included in the fragment, and the bracketed prefix letters and numbers indicate amino acid substitutions at the numbered positions in the native full-length peptide. Thus, for the barbourin fragment above, the length of the fragment spans residues 28-73 inclusive of the native sequence and the amino acids

30

35

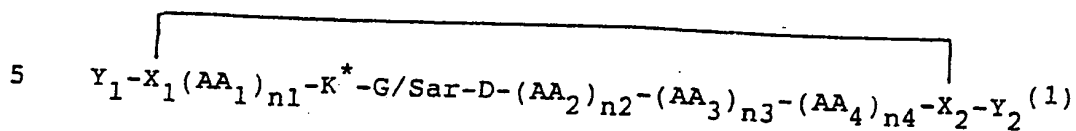
originally in positions 28, 41 and 64 of the numbered native sequence have been replaced by Glu (E), Leu (L), and Cys (C), respectively.

As additional examples, the arginine of the RGD sequence appearing in trigramin, elegantin, albolabrin, crotatroxin, flavoviridin, echistatin, bitistatin, viridin, molossin, lutosin, basilicin, applagin, halysin, horridin, tergeminin, lachesin, cotiarin, cereberin, jararacin, kistrin, eristicophin, bitan-a, and ruberin/oreganin can be replaced by a K^{*} residue to provide specifically active PAIs with a preferential affinity for GP IIB-IIIa. In addition, shortened forms of these peptides, containing at least 20, preferably at least 30, and more preferably at least 40, amino acids, can be prepared from the native peptide or in this modified form. In addition, or in the alternative, 1-10, preferably 1-4, amino acids irrelevant to the RGD/K^{*}GD sequence can be substituted or modified, preferably with conservative amino acid substitutions. By conservative amino acid substitutions is meant, for example, substitution of an acidic amino acid residue for an acidic amino acid residue, neutral for neutral, basic for basic, etc., as is further described hereinbelow.

Still an additional group of examples includes that wherein the glycyl residue of RGD or K^{*}GD can be replaced by a sarcosyl residue with retention of activity. Thus, the active PAIs which are isolated and/or modified in other ways as described above may further be modified by this substitution.

While fragments and/or modified PAIs from snake venom can be included among the Fg/vWF/GP IIB-IIIa binding-specific compounds of the invention by replacing RGD by K^{*}GD, in additional embodiments of the invention specifically active peptides are based on compatible extensions of the K^{*}GD sequence per se. In this regard, a preferred group of peptides or peptide-related compounds

of the invention are cyclic peptides of the general formula:



wherein K^* is substituted or unsubstituted lysyl as above defined;

AA_1 is a small, neutral (polar or nonpolar) amino acid and $n1$ is an integer of 0-3;

AA_2 is a neutral, nonpolar large (aromatic or nonaromatic) or a polar aromatic amino acid and $n2$ is an integer of 0-3;

AA_3 is a proline residue or a modified proline residue (as defined below) and $n3$ is an integer of 0-1;

AA_4 is a neutral, small amino acid or the N-alkylated form thereof and $n4$ is an integer of 0-3;

each of X_1 and X_2 is independently a residue capable of forming a bond between X_1 and X_2 to obtain a cyclic compound as shown; and

each of Y_1 and Y_2 is independently a noninterfering substituent or may be absent;

wherein one or more peptide linkages may optionally be replaced by a linkage selected from the group consisting of $-CH_2NH-$, $-CH_2S-$, CH_2CH_2- , $-CH=CH-$ (cis and trans), $-COCH_2-$, $-CH(OH)CH_2-$ and $-CH_2SO-$;

with the proviso that if $n3$ is 0; either:

- 1) the sum of $n2$ and $n4$ must be at least 2; or
- 2) K^* must be other than Har or K; or
- 3) at least one of X_1 and X_2 must be other than cys (C), penicillamine (Pen), or 2-amino-3,3-cyclopentanemethylene-3-mercaptopropionic acid (APmp); or
- 4) Y_1 or Y_2 must comprise at least one amino acid residue; or

5) one or more peptide linkages is replaced by said alternate linkage.

Y_1 and Y_2 can be peptide extensions of 0-25 amino acid residues and may be in derivatized form. The Y_1 N-terminal extension may, for example, be acetylated or otherwise acylated; the Y_2 C-terminal extension may be
5 amidated with NH_2 or with a primary or secondary amine of the formula $R-NH_2$ or R_2NH wherein each R is independently a lower alkyl of 1-4C such as methyl, n-butyl, or t-butyl. Y_1 can also be (H) or acyl; Y_2 can be (OH), NH_2 or an amine as above. Where the compound of formula (1) is a
10 simple cyclic peptide, Y_1 and Y_2 are absent.

X_1 and X_2 are typically amino acid residues capable of cyclization such as, for example and most preferably, cysteine residues capable of forming a disulfide ring. However, other residues capable of forming
15 disulfide or other linkages may also be used--for example, the Pen (penicillamine) residue described by Pierschbacher et al. (supra) or the Mpr (mercapto propionyl) or Mvl (mercaptovaleryl) residue. Other types of covalent linkages for cyclization envisioned include
20 peptide linkages, as for example, an amide formed between the side-chain amino group of a lysyl residue with a side-chain carboxyl group of a glutamyl residue and ester linkages, such as would be formed between a side-chain alcohol of a threonine residue with a side-chain carboxyl
25 of an aspartyl residue. Any compatible residue capable of forming peptide bonds with the remainder of the chain (or modified peptide bonds as described above) and capable of covalent bond formation to effect cyclization can be used. This includes, for example, simple cyclic peptides,
30 wherein a peptide bond is directly formed between the NH_2 at the N-terminus and the COOH at the C-terminus.

As described above, one or more of the indicated peptide bonds may be replaced by a substitute linkage such as $-CH_2NH-$, $-CH_2S-$, CH_2CH_2- , $-CH=CH-$ (cis and trans),
35 $-COCH_2-$, $-CH(OH)CH_2-$ and $-CH_2SO-$.

In the designation of the amino acid residues AA₁-AA₄ above, description has been made on the basis of a classification method, wherein amino acid residues can be generally subclassified into four major subclasses. This
5 classification is also shown diagrammatically hereinbelow.

Acidic: The residue has a negative charge due to loss of H ion at physiological pH and the residue is attracted by aqueous solution so as to seek the surface positions in the conformation of a peptide in which it is
10 contained when the peptide is in aqueous medium at physiological pH.

Basic: The residue has a positive charge due to association with H ion at physiological pH and the residue is attracted by aqueous solution so as to seek the surface
15 positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium at physiological pH.

Neutral/nonpolar: The residues are not charged at physiological pH and the residue is repelled by aqueous
20 solution so as to seek the inner positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium. These residues are also designated "hydrophobic" herein.

Neutral/polar: The residues are not charged at
25 physiological pH, but the residue is attracted by aqueous solution so as to seek the outer positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium.

It is understood, of course, that in a statistical
30 collection of individual residue molecules some molecules will be charged, and some not, and there will be an attraction for or repulsion from an aqueous medium to a greater or lesser extent. To fit the definition of "charged", a significant percentage (at least ap-
35 proximately 25%) of the individual molecules are charged at physiological pH. The degree of attraction or repul-

sion required for classification as polar or nonpolar is arbitrary, and, therefore, amino acids specifically contemplated by the invention have been specifically classified as one or the other. Most amino acids not specifically named can be classified on the basis of known behavior.

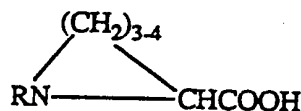
Amino acid residues can be further subclassified as cyclic or noncyclic, and aromatic or nonaromatic, self-explanatory classifications with respect to the side chain substituent groups of the residues, and as small or large. The residue is considered small if it contains a total of 4 carbon atoms or less, inclusive of the carboxyl carbon. Small residues are, of course, always nonaromatic.

For the naturally occurring protein amino acids, subclassification according to the foregoing scheme is as follows (see also the diagram below).

- 20 Acidic: Aspartic acid and Glutamic acid;
- Basic/noncyclic: Arginine, Lysine;
- Basic/cyclic: Histidine;
- 25 Neutral/polar/small: Glycine, Serine and Cysteine;
- Neutral/polar/large/nonaromatic: Threonine, Asparagine, Glutamine;
- 30 Neutral/polar/large/aromatic: Tyrosine;
- Neutral/nonpolar/small: Alanine;
- 35 Neutral/nonpolar/large/nonaromatic: Valine, Isoleucine, Leucine, Methionine;

Neutral/nonpolar/large/aromatic: Phenylalanine,
and Tryptophan.

The gene-encoded amino acid proline, although
5 technically within the group neutral/nonpolar/large/cyclic
and nonaromatic, is a special case due to its known ef-
fects on the secondary conformation of peptide chains, and
is not, therefore, included in this defined group, but is
classified separately. AA₃ is designated a proline
10 residue or a "modified proline residue." Proline, as is
understood, is a five-membered nitrogen heterocycle with a
carboxyl group in the 2-position. Modified proline
residues are all nitrogen five or six-membered
heterocycles with carboxyl groups in the position alpha to
15 the nitrogen; additional heterocyclic atoms may also be
included in the ring. Thus, modified proline residues
include residues of pipecolic acid (2-carboxypiperidine,
abbreviated Pip) and thiazolidine (Thz). Thus, proline or
modified proline residues are of the formula



25 wherein one or two of the methylene groups may be replaced
by NR, S, or O and where any ring nitrogen may optionally
be substituted with a noninterfering substituent such as
alkyl.

Certain commonly encountered amino acids, which
30 are not encoded by the genetic code, include, for example,
beta-alanine (beta-ala), or other omega-amino acids, such
as 3-amino propionic, 4-amino butyric and so forth,
alpha-aminoisobutyric acid (Aib), sarcosine (Sar),
ornithine (Orn), citrulline (Cit), homoarginine (Har),
35 t-butylalanine (t-BuA), t-butylglycine (t-BuG),
N-methylisoleucine (N-MeIle), phenylglycine (Phg), and

cyclohexylalanine (Cha), norleucine (Nle), cysteic acid (Cya); pipecolic acid (Pip), thiazolidine (Thz), 2-naphthyl alanine (2-Nal) and methionine sulfoxide (MSO). These also fall conveniently into particular categories.

5

Based on the above definition,

Sar and beta-ala are neutral/nonpolar/small;

t-BuA, t-BuG, N-MeIle, Nle and Cha are neutral/nonpolar/large/nonaromatic;

10

Har and Orn are basic/noncyclic;

Cya is acidic;

Cit, Acetyl Lys, and MSO are neutral/polar/large/nonaromatic;

2-Nal and Phg are neutral/nonpolar/large/aromatic; and

15

Pip and Thz are modified proline residues.

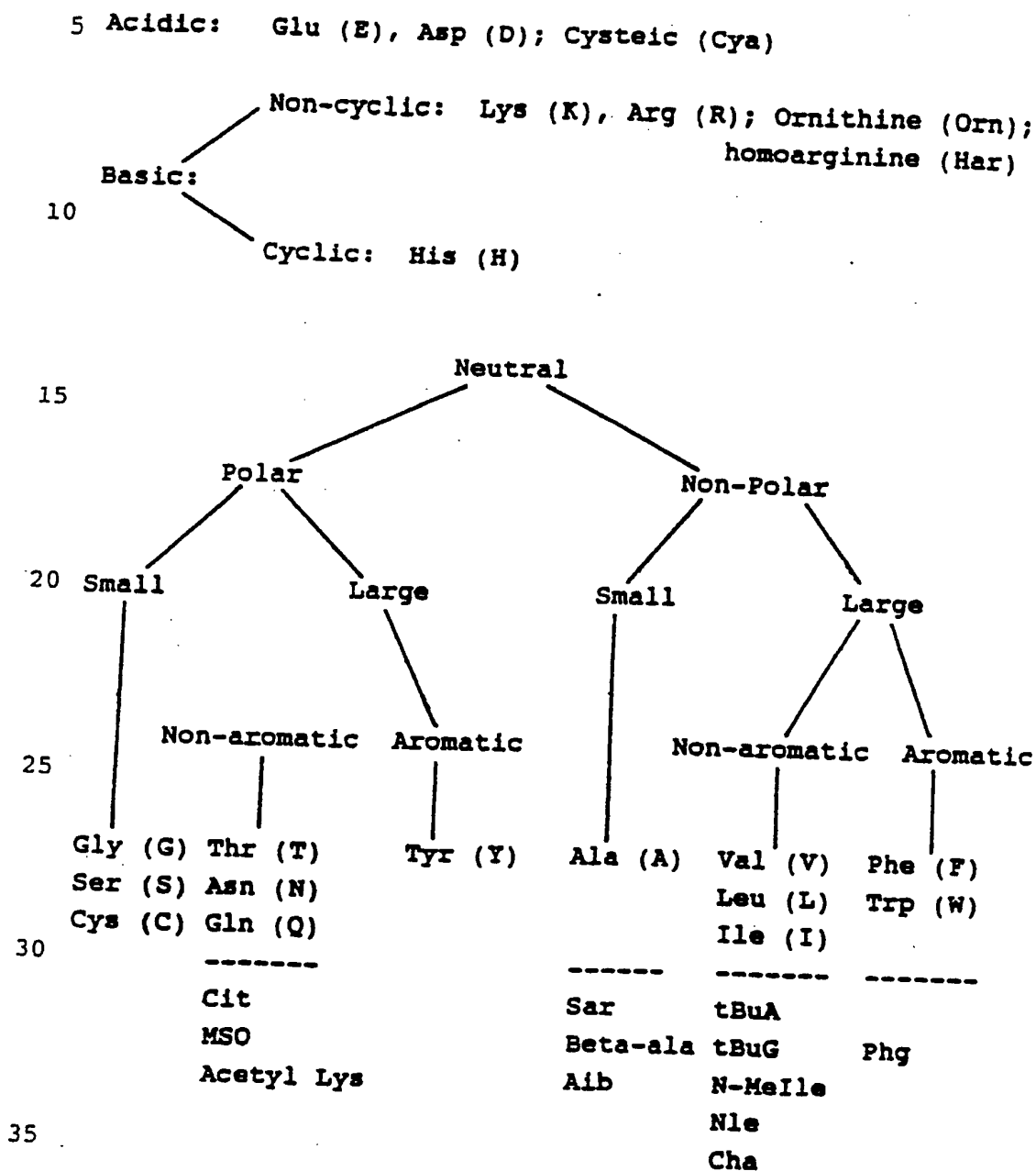
The foregoing may be shown diagrammatically as follows:

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Amino Acid Classification Scheme

The various omega-amino acids are classified according to size as neutral/nonpolar/small (beta-ala, i.e., 3-aminopropionic, 4-aminobutyric) or large (all others).

5 Other amino acid substitutions for those encoded in the gene can also be included in peptide compounds within the scope of the invention and can be classified within this general scheme.

10 In the formulas representing selected specific embodiments of the present invention, the amino- and carboxy-terminal groups, although often not specifically shown, will be understood to be in the form they would assume at physiological pH values, unless otherwise specified. Thus, the N-terminal H^+ and C-terminal- O^- at 15 physiological pH are understood to be present though not necessarily specified and shown, either in specific examples or in generic formulas. Of course, the basic and acid addition salts including those which are formed at nonphysiological pH values are also included in the 20 compounds of the invention. Unless otherwise noted, the residues are in the L-form; in generic formulas, the specified residues can be either L- or D-. Generally, the peptides of the invention have 0, 1, or 2 D-residues included, preferably 0 or 1, most preferably 0. In the 25 peptides shown, each encoded residue where appropriate is represented by a single letter designation, corresponding to the trivial name of the amino acid, in accordance with the following conventional list:

30	<u>Amino Acid</u>	<u>One-Letter Symbol</u>
	Alanine	A
	Arginine	R
35	Asparagine	N

	Aspartic acid	D
	Cysteine	C
	Glutamine	Q
	Glutamic acid	E
5	Glycine	G
	Histidine	H
	Isoleucine	I
	Leucine	L
	Lysine	K
10	Methionine	M
	Phenylalanine	F
	Proline	P
	Serine	S
	Threonine	T
15	Tryptophan	W
	Tyrosine	Y
	Valine	V
	Pyroglutamic acid	Z

20 The amino acids not encoded genetically are abbreviated as indicated above.

In the specific peptides shown in the present application, the L-form of any amino acid residue having an optical isomer is intended unless otherwise expressly
25 indicated by a dagger superscript ([†]). While the residues of the invention peptides are normally in the natural L optical isomer form, one or two, preferably one, amino acid may be replaced with the optical isomer D form.

Free functional groups, including those at the
30 carboxy- or amino-terminus, can also be modified by amidation, acylation or other substitution, which can, for example, change the solubility of the compounds without affecting their activity.

In forming amidated peptides of the present
35 invention, the analog compounds can be synthesized directly, for example using Boc-AA_x-PMBHA-Resin or Boc-

AA_x-BHA-Resin, wherein AA_x is the selected carboxy-terminal amino acid of the desired peptide as described in further detail below. Alternatively, the peptides of the present invention can be chemically or enzymatically
 5 amidated subsequent to peptide synthesis using means well known to the art, or prepared by standard solution-phase peptide synthesis protocols.

Certain embodiments of the de novo peptides of the invention are preferred. In the K*(G/Sar)D sequence,
 10 G/Sar is preferably G. AA₁ and AA₄ are preferably Gly, Ala or Ser; n₁ is preferably 0-2, n₄ is preferably 1-2. Preferred for AA₂ are neutral/nonpolar/aromatic amino acids, especially tryptophan and phenylalanine, particularly tryptophan, n₂ is preferably 1. X₁ and X₂ are
 15 preferably Cys, Mpr, or Pen (penicillamine) residues. Y₁ is preferably H, acetyl, or Gly; Y₂ is preferably -NH₂ or -A-NH₂. Also preferred generally are C-terminal amidated forms of Y₂.

Thus, preferred embodiments of the PAI analogs
 20 of the invention include peptides of the following formulas. Although all of these are capable of provision in cyclic form through formation of disulfide linkages, these linkages are not specifically shown; other cyclic forms are noted by "cyclo."

Preferred Peptides

PAI 1: E-C-A-D-G-L-C-C-D-Q-C-R-F-L-K-K-G-T-V-
 C-R-V-A-K-G-D-W-N-D-D-T-C-T-G-Q-S-C-D-C-P-R-N-G-L-Y-G

30 PAI 2: E-E-P-C-A-T-G-P-C-C-R-R-C-K-F-K-R-A-G-
 K-V-C-R-V-A-K-G-D-W-N-N-D-Y-C-T-G-K-S-C-D-C-P-R-N-P-W-N-G

PAI 3: G-C-G-K-G-D-W-P-C-A-NH₂;

PAI 4: G-C-K-G-D-W-P-C-A-NH₂

PAI 5: C-G-K-G-D-W-P-C-NH₂

35 PAI 7: C-K-G-D-W-C-A-NH₂;

PAI 9: Mpr-K-G-D-Pen-NH₂

	PAI 10:	C-K-G-D-W-P-C-NH ₂
	PAI 12:	C-K-G-D-Y-P-C-NH ₂
	PAI 13:	C-K-G-D-F-P-C-NH ₂
5	PAI 14:	C-K-G-D-L-P-C-NH ₂
	PAI 15:	C-K-G-D-V-P-C-NH ₂
	PAI 16:	C-K-G-D-Y(OMe)-P-C-NH ₂
10	PAI 17:	C-K-G-D-(2-Nal)-P-C-NH ₂
	PAI 18:	C-K-G-D-(Cha)-P-C-NH ₂
	PAI 19:	Mpr-K-G-D-W-P-C-NH ₂
	PAI 20:	Mpr-K-G-D-Y-P-C-NH ₂
15	PAI 21:	Mpr-K-G-D-F-P-C-NH ₂
	PAI 22:	Mpr-K-G-D-L-P-C-NH ₂
	PAI 23:	Mpr-K-G-D-V-P-C-NH ₂
	PAI 24:	Mpr-K-G-D-Y(OMe)-P-C-NH ₂
20	PAI 25:	Mpr-K-G-D-(2-Nal)-P-C-NH ₂
	PAI 26:	Mpr-K-G-D-(Cha)-P-C-NH ₂
	PAI 27:	cyclo(G-K-G-D-W-P)
	PAI 28:	cyclo(A-K-G-D-W-P)
25	PAI 29:	cyclo(D-Ala-K-G-D-W-P)
	PAI 30:	cyclo(F-K-G-D-W-P)
	PAI 31:	cyclo(beta-Ala-K-G-D-W-P)
30	PAI 32:	cyclo(gamma-Abu-K-G-D-W-P)
	PAI 33:	cyclo(R-K-G-D-W-P)
	PAI 34:	C-K-G-D-W-G-C-NH ₂
	PAI 37:	C-K-A-D-W-P-C-NH ₂
35	PAI 39:	C-K-G-D-W-(Sar)-C-NH ₂
	PAI 41:	C-K-G-D-I-P-C-NH ₂

- 5 PAI 42: C-K-G-D-(4-Cl-Phe)-P-NH₂
PAI 43: C-K-(Sar)-D-W-P-C-NH₂
PAI 44: C-K-G-D-(4-NO₂-Phe)-P-C-NH₂
PAI 47: Acetyl-C-K-G-D-W-P-C-NH₂
PAI 48: Mpr-K-G-D-W(Formyl)-P-C-NH₂
PAI 49: Mvl-K-G-D-W-P-C-NH₂
10 PAI 51: Mpr-K-G-D-W-P-Pen-NH₂
PAI 52: Mpr-K-G-D-W-P-Pen[†]-NH₂
PAI 54: Mpr-K-G-D[†]-W-P-Pen-NH₂
PAI 55: Mpr-K-G-D-W-(Thz)-C-NH₂
15 PAI 56: Mpr-K-G-D-H(2,4-DNP)-P-C-NH₂
PAI 57: Mpr-K-G-D-(2-Nal)-P-Pen-NH₂
PAI 58: Mvl-K-G-D-W-P-Pen-NH₂
PAI 59: Mpr-K-G-D-W-(Pip)-Pen-NH₂
20 PAI 60: Mpr-(Har)-G-D-W-P-C-NH₂
PAI 61: Mpr-K-G-D-W-P-C[†]-NH₂
PAI 62: Mpr-K[†]-G-D-W-P-Pen-NH₂
PAI 63: Mpr-(Har)-G-D-W-P-Pen-NH₂
25 PAI 64: Mpr-(Acetimidyl-Lys)-G-D-W-P-C-NH₂
PAI 65: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
PAI 66: Mpr-(N^G,N^{G'}-ethylene-Har)-G-D-W-P-C-NH₂
PAI 67: Mpr-(N^G,N^{G'}-ethylene-Har)-G-D-W-P-Pen-NH₂
30 PAI 68: Mpr-Har-Sar-D-W-P-C-NH₂
PAI 69: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
PAI 70: Mpr-(Phenylimidyl-Lys)-G-D-W-P-C-NH₂
35 PAI 71: Mpr-Har-Sar-D-W-P-PenNH₂
PAI 72: Mpr-(Phenylimidyl-Lys)-G-D-W-P-PenNH₂

PAI 73: Mpr-Har-G-D-W-(3,4-dehydro-Pro)-C-NH₂

PAI 74: Mpr-Har-G-D-Pen-NH₂

PAI 75: Mpr-(Phenylimidyl-Lys)-G-D-Pen-NH₂

5

formulas

PAI 3: G-C-G-K-G-D-W-P-C-A-NH₂;

PAI 4: G-C-K-G-D-W-P-C-A-NH₂;

10

PAI 5: C-G-K-G-D-W-P-C-NH₂;

PAI 9: Mpr-K-G-D-Pen-NH₂; and

PAI 10: C-K-G-D-W-P-C-NH₂

PAI 12: C-K-G-D-Y-P-C-NH₂

15

PAI 13: C-K-G-D-F-P-C-NH₂

PAI 19: Mpr-K-G-D-W-P-C-NH₂

PAI 25: Mpr-K-G-D-(2-Nal)-P-C-NH₂

20

PAI 34: C-K-G-D-W-G-C-NH₂

PAI 39: C-K-G-D-W-(Sar)-C-NH₂

PAI 42: C-K-G-D-(4-Cl-Phe)-P-NH₂

PAI 43: C-K-(Sar)-D-W-P-C-NH₂

25

PAI 44: C-K-G-D-(4-NO₂-Phe)-P-C-NH₂

PAI 47: Acetyl-C-K-G-D-W-P-C-NH₂

PAI 48: Mpr-K-G-D-W(Formyl)-P-C-NH₂

PAI 49: Mvl-K-G-D-W-P-C-NH₂

30

PAI 51: Mpr-K-G-D-W-P-Pen-NH₂

PAI 52: Mpr-K-G-D-W-P-(D-Pen)-NH₂

PAI 55: Mpr-K-G-D-W-(Thz)-C-NH₂

PAI 56: Mpr-K-G-D-H(2,4-DNP)-P-C-NH₂

35

PAI 57: Mpr-K-G-D-(2-Nal)-P-Pen-NH₂

PAI 58: Mvl-K-G-D-W-P-Pen-NH₂

- PAI 59: Mpr-K-G-D-W-(Pip)-Pen-NH₂
 PAI 60: Mpr-(Har)-G-D-W-P-C-NH₂
 PAI 61: Mpr-K-G-D-W-P-C[†]-NH₂
 5 PAI 62: Mpr-K[†]-G-D-W-P-Pen-NH₂
 PAI 63: Mpr-(Har)-G-D-W-P-Pen-NH₂
 PAI 64: Mpr-(Acetimidyl-Lys)-G-D-W-P-C-NH₂
 PAI 65: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
 10 PAI 66: Mpr (N^G,N^{G'}-ethylene-Har)-G-D-W-P-C-NH₂
 PAI 67: Mpr (N^G,N^{G'}-ethylene-Har)-G-D-W-P-Pen-NH₂
 PAI 68: Mpr-Har-Sar-D-W-P-C-NH₂
 15 PAI 69: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
 PAI 70: Mpr-(Phenylimidyl-Lys)-G-D-W-P-C-NH₂
 PAI 71: Mpr-Har-Sar-D-W-P-PenNH₂
 PAI 72: Mpr-(Phenylimidyl-Lys)-G-D-W-P-PenNH₂
 20 PAI 73: Mpr-Har-G-D-W-(3,4-dehydro-Pro)-C-NH₂

Chemical Synthesis of the Invention Peptides

Compounds within the scope of the present invention can be synthesized chemically by means well known in the art such as, e.g., solid-phase peptide synthesis. The synthesis is commenced from the carboxy-terminal end of the peptide using an alpha-amino protected amino acid. t-Butyloxycarbonyl (Boc) protective groups can be used for all amino groups even though other protective groups such as fluorenylmethyloxycarbonyl (Fmoc), are suitable. For example, Boc-Gly-OH, Boc-Ala-OH, Boc-His (Tos)-OH, (i.e., selected carboxy-terminal amino acids) can be esterified to chloromethylated polystyrene resin supports, p-methyl benzhydrylamine (pMBHA) or PAM resins. The polystyrene resin support is preferably a copolymer of styrene with about 0.5 to 2% divinyl benzene as a cross-linking agent

which causes the polystyrene polymer to be completely insoluble in certain organic solvents. See Stewart, et al., Solid-Phase Peptide Synthesis (1969) W.H. Freeman Co., San Francisco and Merrifield, J Am Chem Soc (1963) 85:2149-2154. These and other methods of peptide synthesis are also exemplified by U.S. Patent Nos. 3,862,925, 3,842,067, 3,972,859, and 4,105,602.

The synthesis may use manual synthesis techniques or automatically employ, for example, an Applied BioSystems 430A or 431A Peptide Synthesizer (Foster City, California) following the instructions provided in the instruction manual supplied by the manufacturer. Cleavage of the peptides from the resin can be performed using the "low-high" HF deprotection protocols as described in Lu, G.-S., et al., Int J Peptide & Protein Res (1987) 29:545-557. Refolding of analogs of the snake venom PAIs can be performed using the procedure outlined in Garsky, V., et al., Proc Natl Acad Sci USA (1989) 86:4022-4026 which describes the solid-phase synthesis of echistatin.

The cyclic peptides of this invention which do not have disulfide bonds can be conveniently prepared by a combination of solid phase synthesis and formation of the cyclic ring structure in solution using the general methods as outlined in U.S. Patent 4,612,366 to Nutt. Thus, linear peptides prepared on standard Merrifield resin can be cleaved from the resin with hydrazine, followed by cyclization of the corresponding azide to form the cyclic peptides.

It will be readily appreciated by those having ordinary skill in the art of peptide synthesis that the intermediates which are constructed in accordance with the present disclosure during the course of synthesizing the present analog compounds are themselves novel and useful compounds and are thus within the scope of the invention.

Recombinant Production

Alternatively, selected compounds of the present invention can be produced by expression of recombinant DNA constructs prepared in accordance with well-known methods. Such production can be desirable to provide large quantities or alternative embodiments of such compounds. Since the peptide sequences are relatively short, recombinant production is facilitated; however, production by recombinant means is particularly preferred over standard solid phase peptide synthesis for peptides of at least 8 amino acid residues.

The DNA encoding the sequenced PAI is preferably prepared using commercially available nucleic acid synthesis methods. Methods to construct expression systems for production of PAI in recombinant hosts are also generally known in the art.

Expression can be effected in either procaryotic or eucaryotic hosts. Procaryotes most frequently are represented by various strains of E. coli. However, other microbial strains may also be used, such as bacilli, for example Bacillus subtilis, various species of Pseudomonas, or other bacterial strains. In such procaryotic systems, plasmid vectors which contain replication sites and control sequences derived from a species compatible with the host are used. For example, a workhorse vector for E. coli is pBR322 and its derivatives. Commonly used procaryotic control sequences, which contain promoters for transcription initiation, optionally with an operator, along with ribosome binding-site sequences, include such commonly used promoters as the beta-lactamase (penicillinase) and lactose (lac) promoter systems, the tryptophan (trp) promoter system, and the lambda-derived P_L promoter and N-gene ribosome binding site. However, any available promoter system compatible with procaryotes can be used.

Expression systems useful in eucaryotic hosts comprise promoters derived from appropriate eucaryotic genes. A class of promoters useful in yeast, for example, includes promoters for synthesis of glycolytic enzymes, e.g., those for 3-phosphoglycerate kinase. Other yeast promoters include those from the enolase gene or the Leu2 gene obtained from YEp13.

Suitable mammalian promoters include the early and late promoters from SV40 or other viral promoters such as those derived from polyoma, adenovirus II, bovine papilloma virus or avian sarcoma viruses. Suitable viral and mammalian enhancers are cited above. In the event plant cells are used as an expression system, the nopaline synthesis promoter, for example, is appropriate.

The expression systems are constructed using well-known restriction and ligation techniques and transformed into appropriate hosts.

Transformation is done using standard techniques appropriate to such cells. The cells containing the expression systems are cultured under conditions appropriate for production of the PAI, and the PAI is then recovered and purified.

Antibodies

The availability of the purified PAI of the invention also permits the production of antibodies specifically immunoreactive with these forms of the active peptide.

The compositions containing purified PAI isolated from snake venom or otherwise synthesized can be used to stimulate the production of antibodies which immunoreact with the PAI peptide. Standard immunization protocols involving administering PAI to various vertebrates, such as rabbits, rats, mice, sheep, and chickens result in antisera which are immunoreactive with the purified peptide. PAI may be advantageously

conjugated to a suitable antigenically neutral carrier, such as an appropriate serum albumin or keyhole limpet hemocyanin, in order to enhance immunogenicity. In addition, the free peptide can be injected with methylated BSA as an alternative to conjugation. Furthermore, the antibody-secreting cells of the immunized mammal can be immortalized to generate monoclonal antibody panels which can then be screened for reactivity with PAI.

The resulting polyclonal or monoclonal antibody preparations are useful in assays for levels of the corresponding PAI in biological samples using standard immunoassay procedures.

The Invention Assay

The identification of snake venom starting material which contains active PAI, and which PAI has known specificity, is made possible by the assay of the invention. The assay rests on the observation that compounds which block the binding of fibrinogen to the GP IIB-IIIA complex in vitro also are capable of inhibiting thrombin or ADP-induced aggregation of human platelets and the formation of platelet-thrombi in vivo. This observation provides the basis for obtaining potent PAI by evaluating the ability of test materials to disrupt fibrinogen-GP IIB-IIIA interactions.

In the assay, GP IIB-IIIA, prepared in purified form, for example as described by Fitzgerald, L.A., et al., Anal Biochem (1985) 151:169-177, incorporated herein by reference, is coated onto a solid support such as beads, test tubes, or microtiter plates. The coated support is then contacted with fibrinogen and with the test material and incubated for a sufficient time to permit maximal binding of fibrinogen to the immobilized GP IIB-IIIA. Fibrinogen is typically provided at a concentration of about 5-50 nM and the test material can, if desired, be added at a series of dilutions. Typical incubations are

2-4 hr at 35°C, the time and temperature being inter-dependent.

After incubation, the solution containing the fibrinogen and test material is removed and the level of binding of fibrinogen measured by quantitating bound fibrinogen to GP IIB-IIIa. Any suitable means of detection may be used, but it is convenient to employ labeled fibrinogen, for example using radioactive, fluorescent or biotinylated labels. Such methods are well known and need not be elaborated here.

Assessment of the results is aided by employing a control sample, usually identical to the test sample except that the test substance is absent. In this case, percent inhibition may be calculated using the amount of Fg bound in the control as representing the basis, so that

$$\% \text{ inhibition} = \frac{\text{control} - \text{test}}{\text{control}} \times 100.$$

Other measures of inhibition effectiveness, such as IC₅₀, may also be used.

The assay systems of the invention further include characterization of the PAI specificity by binding inhibition assays identical to that above but substituting other adhesive proteins for Fg and other receptors for GP IIB-IIIa. In particular, inhibition of the binding of vitronectin to the vitronectin receptor; fibronectin to the fibronectin receptor; fibronectin to GP IIB-IIIa and fibrinogen and/or vWF to GP IIB-IIIa may be assessed. The adhesive protein and receptors for these assays are available in the art.

Other Assays

In addition to the plate assays of the invention, other assays for platelet aggregation inhibition activity and related activities are also available, as set

forth above. In summary, a list of commonly employed assays is as follows:

1. The plate assays utilizing specific receptors described in the previous paragraphs;
2. Standard assays directly applied to platelet aggregation, such as those described by Gann, Z.-R., et al., J Biol Chem (1988) 263:19827-19832; Huang, T.F., et al., J Biol Chem (1987) 262:16157-16163; Biochemistry (1989) 28:661-666, cited above and incorporated herein by reference;
3. An in vivo thrombosis model in dogs as described hereinbelow in Example 1, and by Folts, J.D., et al., Circulation (1976) 54:365; and
4. Effect on cell adhesion using S35 methionine-labeled cells as described hereinbelow in Example 19.

Administration and Utility

The PAIs of the invention are useful therapeutically to prevent thrombus formation. Indications appropriate to such treatment include, without limitation, atherosclerosis and arteriosclerosis, acute myocardial infarction, chronic unstable angina, transient ischemic attacks and strokes, peripheral vascular disease, arterial thrombosis, preeclampsia, embolism, restenosis and/or thrombosis following angioplasty, carotid endarterectomy, anastomosis of vascular grafts, and chronic cardiovascular devices (e.g., in-dwelling catheters or shunts "extracorporeal circulating devices"). These syndromes represent a variety of stenotic and occlusive vascular disorders thought to be initiated by platelet activation on vessel walls.

The PAIs may be used for prevention or abortion of arterial thrombus formation, in unstable angina and arterial emboli or thrombosis, as well as treatment or prevention of myocardial infarction (MI) and mural

thrombus formation post MI. For brain-related disorders, treatment or prevention of transient ischemic attack and treatment of thrombotic stroke or stroke-in-evolution are included.

5 The PAIs may also be used for prevention of platelet aggregation, embolization, or consumption in extracorporeal circulations, including improving renal dialysis, cardiopulmonary bypasses, hemoperfusions, and plasmapheresis.

10 PAIs prevent platelet aggregation, embolization, or consumption associated with intravascular devices, and administration results in improved utility of intraaortic balloon pumps, ventricular assist devices, and arterial catheters.

15 The PAIs will also be useful in treatment or prevention of venous thrombosis as in deep venous thrombosis, IVC, renal vein or portal vein thrombosis, and pulmonary venous thrombosis.

20 Various disorders involving platelet consumption, such as thrombotic thrombocytopenic purpura are also treatable.

 In addition, the PAIs of the present invention may be used in numerous nontherapeutic applications where inhibiting platelet aggregation is desired. For example, improved platelet and whole blood storage can be obtained by adding sufficient quantities of the peptides, the amount of which will vary depending upon the length of proposed storage time, the conditions of storage, the ultimate use of the stored material, etc.

25 The PAI dosage can range broadly depending upon the desired affects and the therapeutic setting. Typically, dosages will be between about 0.01 and 10 mg/kg, preferably between about 0.01 to 0.1 mg/kg, body weight. Administration is preferably parenteral, such as intravenous on a daily basis for up to a week or as much as one or two months or more, all of which will vary with

the peptide's size. If the peptides are sufficiently small (e.g., less than about 8-10 amino acid residues) other routes of administration can be utilized, such as intranasally, sublingually, or the like.

5 Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution or suspension in liquid prior to injection, or as emulsions. Suitable excipients are, for example, water, saline, dextrose, mannitol, lactose, 10 lecithin, albumin, sodium glutamate, cysteine hydrochloride or the like. In addition, if desired, the injectable pharmaceutical compositions may contain minor amounts of nontoxic auxiliary substances, such as wetting agents, pH buffering agents, and the like. If desired, 15 absorption enhancing preparations (e.g., liposomes) may be utilized.

Example 1

20 Assay for Snake Venom Platelet Adhesion Inhibitors

A. Description of Assays--Plate Assays

Purified platelet GP IIb-IIIa receptor was prepared as described by Fitzgerald, L.A., et al., Anal 25 Biochem (1985) 151:169-177. Vitronectin receptor was prepared as described by Smith, J.W., J Biol Chem (1988) 263:18726-18731. After purification, the receptors were stored in 0.1% Triton X-100 at 0.1-1.0 mg/ml.

The receptors were coated to the wells of 96- 30 well flat-bottom ELISA plates (Linbro EIA-Plus microtiter plate, Flow Laboratories) after diluting 1:200 with a solution of 20 mM Tris-HCl, 150 mM NaCl, 1 mM CaCl₂, pH 7.4, to reduce the Triton X-100 concentration to below its critical micellar concentration and adding an aliquot of 35 100 ul to each well. The wells were all allowed to incubate overnight at 4°C, and then aspirated to dryness.

Additional sites were blocked by the addition of bovine serum albumin (BSA) at 35 mg/ml in the above buffer for 2 hr at 30° C. to prevent nonspecific binding. The wells were then washed once with binding buffer (50 mM Tris-HCl, 100 mM NaCl, 2 mM CaCl₂, 1 mg/ml BSA).

The corresponding ligands (fibrinogen, von Willebrand Factor, or vitronectin) were labeled with ¹²⁵I or conjugated to biotin using commercially available reagents and standard protocols. The labeled ligands were added to the receptor-coated wells at final concentration of 10 nM (100 ul/well) and incubated for 3 h at 30°C in the presence or absence of the test samples. After incubation, the wells are aspirated to dryness and bound ligand is quantitated.

For ¹²⁵I-labeled ligands, the protein is solubilized with 250 ul SDS. For biotinylated ligands, the bound protein is detected by the addition of antibiotin antibody conjugated to alkaline phosphatase followed by addition of substrate (p-nitrophenyl phosphate), and determination of the optical density of each well at 405 nm. Decreased color development or decreased ¹²⁵I content is observed in wells incubated with test samples which inhibit binding of ligand to receptor.

B. Determination of Adhesion Inhibition in Crude Venom

Sixty-eight crude, lyophilized snake venoms obtained from either Sigma Chemical Company (St. Louis, MO) or Miami Serpentarium Labs (Salt Lake City, UT) were dissolved at 1 mg/ml in buffer (50 mM Tris, 100 mM NaCl, 0.02% azide, 2 mM CaCl₂). One ml aliquots of the solutions were subjected to ultrafiltration through Centrocon-10 (YM membrane) microconcentrators (Amicon, Danvers, MA). The filtrates were used as test samples in the receptor/ligand assay of paragraph A using the GP IIb-IIIa/fibrinogen system, and detecting binding using

biotinylated fibrinogen. The results are shown in Table 1.

It is seen that the activity is present in some, but not all, species of Viperinae, but absent in all species tested of Elapidae.

Figure 1 shows the results at various dilutions of the filtrate for four species. Even at the greatest dilution, 25 ul/0.5 ml, the three active venoms showed maximal inhibition.

C. Determination of the Activity of Peptides in an in vivo Model of Thrombosis

Purified peptides were tested for their ability to prevent thrombi formation in dog coronary arteries in the model described by Folts (Folts J.D., et al., Circulation (1976) 54:365. In this model, flow reductions in a constricted coronary artery have been shown to be due to the formation of platelet aggregates, and agents which block the binding of fibrinogen to GP IIb-IIIa have been shown to prevent these flow reductions (Coller, B.S., et al., Blood (1986) 68:783. The peptides were dissolved in normal saline and administered into a peripheral vein as a single bolus.

TABLE 1

CENTRICON 10 PURIFIED VENOMS SCREENED IN IIB-IIIa PLATE ASSAY		
5	Elapids	Activity
	<i>Austrelaps superba</i> (Australian Copperhead)	-
	<i>Acanthopis antarcticus</i> (Death Adder)	-
10	<i>Dendroaspis jamesonii</i> (Jameson's Mamba)	-
	<i>Notechis scutatus</i> (Mainland Tiger)	-
	<i>Pseudechis colleti guttatus</i> (Blue-bellied)	-
	<i>Pseudechis textillis textillis</i> (Common Brown)	-
15	<i>Oxyuranus scutellatus</i> (Papuan Taipan)	-
	Viperinae (True Vipers)	Activity
	<i>Atheris squamigera</i> (Green Bush Viper)	-
	<i>Bitis nasicornus</i> (River Jack)	-
20	<i>Causus rhombeatus</i> (Rhombic Night Adder)	-
	<i>Cerastes cerastes</i> (Desert Horned Viper)	-
	<i>Cerastes vipera</i> (Sahara Horned Viper)	-
	<i>Echis carinatus</i> (Saw-scaled Viper)	+
25	<i>Echis colorata</i> (Carpet Viper)	+
	<i>Eristicophis macmahonii</i> (Macmahons Viper)	++
	<i>Pseudocerastes fieldi</i> (Persian Horned Viper)	-
	<i>Vipera xanthina xanthina</i> (Ottomans Viper)	-
30	<i>Vipera ammodytes</i> (Long-nosed Viper)	-
	<i>Vipera r. russelli</i> (Russells Viper)	-
	<i>Vipera r. siamensis</i>	-
35	<i>Vipera palaestinae</i> (Palestine Viper)	-

TABLE 1 (Continued)

5	Crotalinae (Pit Vipers)	Activity
	Agkistrodon rhodostoma (Malayan Pit Viper)	+
	Agkistrodon halys blomhoffi (Mamushi)	+
10	Agkistrodon hypnale (Hump-nosed Viper)	+
	Agkistrodon acutus (Sharp-nosed Viper)	++
	Agkistrodon bilineatus (Mexican Moccasin)	-
	Agkistrodon contortrix contortrix	-
15	Agkistrodon c. laticinctus	-
	Agkistrodon c. pictigaster	-
	Agkistrodon contortrix mokasen (Northern Copperhead)	-
	Agkistrodon piscivorous piscivorous (Eastern Cottonmouth)	-
20	Agkistrodon piscivorus leucostoma (Western Cottonmouth)	+
	Agkistrodon piscivorous conanti	+
	Bothrops asper	+
	Bothrops nummifer (Jumping Viper)	-
25	Bothrops cotiara (Cotiara)	+
	Bothrops jararacussu (Jararacussu)	+
	Bothrops jararaca (Jararaca)	+
	Bothrops lansbergi	+
30	Bothrops alternata (Urutu)	-
	Bothrops medusa	+
	Bothrops neuwiedi	+
	Bothrops nasuta	+
35	Bothrops pradoi	+

TABLE 1 (Continued)

5	<i>Bothrops schlegli</i> (Schlegel's Viper)	-
	<i>Trimeresurus gramineus</i> (Formosan Green Habu)	-
	<i>Trimeresurus flavoviridis</i> (Okinawa Habu)	-
10	<i>Trimeresurus wagleri</i>	-
	<i>Lachesis muta</i> (Bushmaster)	-
	<i>Crotalus durissus terrificus</i> (Tropical Rattlesnake)	-
	<i>Crotalus durissus totonatacus</i>	-
15	<i>Crotalus durissus durissus</i>	-
	<i>Crotalus scutalatus</i> (Mojave rattlesnake)	-
	<i>Crotalus horridus horridus</i> (Timber Rattlesnake)	-
	<i>Crotalus horridus atricaudatus</i> (Canebrake RS)	-
20	<i>Crotalus atrox</i> (Western Diamondback)	-
	<i>Crotalus adamanteus</i> (Eastern Diamondback)	-
	<i>Crotalus basiliscus</i> (Mexican West-coast RS)	-
	<i>Crotalus molossus molossus</i> (Black-tailed RS)	-
25	<i>Crotalus ruber ruber</i> (Red diamondback RS)	-
	<i>Crotalus cerastes cerastes</i> (Mojave sidewinder)	-
	<i>Crotalus viridis viridis</i> (Prairie Rattlesnake)	+
	<i>Crotalus v. helleri</i> (Southern Pacific RS)	+
30	<i>Crotalus v. oregonus</i> (Northern Pacific RS)	+
	<i>Crotalus v. cereberus</i> (Arizona black RS)	+
	<i>Crotalus v. lutosus</i> (Great Basin RS)	+
	<i>Crotalus v. concolor</i> (Midget-faded RS)	-
35	<i>Sistrurus catenatus teryeminus</i> (Western massasauga)	+
	<i>Sistrurus miliaris barbouri</i> (Southeastern Pigmy Rattlesnake)	+

D. Effects of Purified Snake Venom Peptides on Cell Attachment to Adhesive Proteins

M21 melanoma cells, which express high levels of the vitronectin receptor, were metabolically labelled with ³⁵S-methionine, and then added to 24-well tissue culture plates coated with vitronectin. An incubation period of 1 hr at 37°C was allowed for cell attachment, and this was followed by a wash to remove non-adherent cells. After washing, the adherent cells were solubilized, and the supernatants placed in a liquid scintillation counter. The fraction of cells remaining adherent was calculated by dividing the cpm in the solubilized supernatants by the cpm in the total number of cells added to each well. The effects of purified snake venom peptides and synthetic cyclic peptides on cell adhesion was determined by including them with the M21 cells during the incubation period.

E. Specificity of Adhesion Inhibition

Ultrafiltrates from three species of snake venom, Sistrurus m. barbouri, Crotalus ruber ruber, and Crotalus basilicus, were tested in both the fibrinogen/GP IIB-IIIA and vitronectin/vitronectin receptor assays of paragraph A. The results were evaluated at various dilutions. As shown in Figure 2, the venom from Sistrurus m. barbouri preferentially inhibits the binding of fibrinogen to GP IIB-IIIA; the venom of Crotalus ruber ruber inhibits binding in both systems approximately equally; and the venom from Crotalus basilicus preferentially inhibits vitronectin/vitronectin receptor binding.

In the purifications described in Examples 2-6 and 8-12, PAI activity was assayed using a direct inhibition of platelet aggregation. Platelet rich plasma (PRP) was obtained from a healthy human volunteer. Aggregation was induced by the addition of 4 uM ADP to 0.5 ml PRP in an aggregometer (Chrono-log Corp.).

A table showing results of amino acid composition analysis of purified PAIs of Examples 2-6 will be found after Example 6; that showing the results for Examples 8-11 is shown after Example 8.

- 5 This analysis was obtained by hydrolysis of peptides using 6 N HCl and analyzing the hydrolysate using a Beckman 121 HC analyzer equipped with a Model 126 data system. Cysteic acid was determined according to the method of Moore, J Biol Chem (1969) 230:235-237.
- 10 Tryptophan was not determined.

Example 2

Purification of Platelet Aggregation Inhibitor (PAI) From Eristocophis macmahoni Venom

- 15 A solution of 45 mg of Eristocophis macmahoni venom (Miami Serpentarium Labs, Lot #EM23SZ) in 1.0 ml of 0.5% trifluoroacetic acid (TFA) was cooled on ice for 20 min, spun at 14,000 rpm for 3 min to remove insoluble material and loaded onto a 3.9 mm x 30 cm, C-18 Delta Pak
- 20 reverse-phase HPLC column (Waters, Milford, MA) equilibrated with 5% acetonitrile containing 1% TFA. A gradient running from 5% to 15% acetonitrile over 5 min (2%/min) followed by a gradient from 15% to 30% acetonitrile over 35 min and then to 50% acetonitrile over
- 25 20 min, was run using a Waters 600E liquid chromatograph. A flow rate of 1.5 ml/min was maintained throughout the gradient and column effluent was collected in 2 min fractions into polypropylene tubes.

- 30 The column effluent was monitored at 220 nm/2.5 absorbance units full scale (AUFS).

- Fractions were concentrated to one-half their original volume using a Speed-Vac concentrator (Savant) followed by lyophilization. Samples were then re-
- 35 constituted in 1 ml distilled water and aliquots (10-50 ul) assayed for their ability to inhibit human platelet aggregation in platelet-rich plasma induced by 20 uM ADP

using a whole blood aggregometer (Chrono-Log Corp., Havertown, PA).

As shown in Figure 3, activity was found in fractions that eluted at 21-25% acetonitrile concentration. These fractions were then lyophilized and rerun on the C-18 HPLC column using shallower acetonitrile gradient as follows: Initial conditions consisted of 8% acetonitrile followed by a gradient to 25% acetonitrile over 68 min (0.25%/min), then to 60% acetonitrile in 10 min. One-minute fractions were collected, dried and reassayed for inhibitory activity in platelet aggregation of human platelets as above.

As shown in Figure 4, the activity eluted at 24% acetonitrile. The active fractions were then subjected to analytical HPLC with detection at 220 nm and eluted as a single symmetric bioactive component as shown in Figure 5. Amino acid analysis of the HPLC-purified material showed that the peptide contains 49 residues including 7-8 cysteines, as set forth in Table 2.

Attempts at automated Edman degradation of the carboxyamidomethylated peptide did not yield any detectable sequence. Therefore, digestion of this material was performed with Lys-C and Asp-N endoproteinases yielding fragments which were sequenced and are shown in Figure 6. This analysis revealed a sequence of 48 residues. However, since two tryptophan residues are apparent from this sequence analysis which were not determined in the amino acid composition, the intact peptide contains 51 amino acid residues. Thus, two Glx and one Arg residues missing from the determined sequence were presumably present at the blocked amino terminus of the peptide. Since it was quite likely that one of the Glx residues was a pyroglutamyl residue at the amino terminus leading to the blocked nature of the intact peptide, we removed this group from the intact, carboxyamidomethylated peptide with the enzyme pyroglutamyl aminopeptidase (L-pyroglutamyl

peptide hydrolase, EC 3.4.11.8, Boehringer Mannheim Biochemicals, Indianapolis, IN). Protocols described by Podell and Abraham, Biochem Biophys Res Commun (1978) 81:176-185 were used. Digestion of 100 ug of peptide with the peptidase at a substrate-to-enzyme ratio of 100:1, followed by reversed-phase HPLC purification of the mixture on a Waters analytical C-18 column gave material suitable for automated Edman degradation. The results of this analysis and the assignment of the entire sequence of this peptide which was named "eristicophin," is shown in Figure 6.

The complete amino acid sequence of this PAI, is shown in Figure 6. This peptide has RGD in the binding region and shows considerable homology to echistatin.

Example 3

Purification of PAI from Sistrurus catenatus tergeminus Venom

Three hundred sixty mg of Sistrurus c. tergeminus venom (Miami Serpentarium Labs, Lot #ST6SZ) was dissolved in 7.0 ml of 0.5 M acetic acid and applied to a column of Sephadex G-50 fine (Pharmacia, 2.5 x 100 cm) equilibrated and eluted with 0.5 M acetic acid. The column was run at a flow rate of approximately 25 ml/hr and 5-ml fractions collected. Twenty-five ul of each fraction was pooled in groups of 10 fractions (i.e., fractions 1-10, 11-20, etc.) and lyophilized for analysis. The dried pooled fractions were redissolved in water and aliquots assayed for inhibitory activity in ADP-stimulated aggregation of human platelets. Active fractions (31-40) were pooled and lyophilized.

This material was dissolved in 2 ml of 0.5% TFA and loaded onto a 19 mm x 30 cm C-18 Delta Pak reverse-phase HPLC column (Waters) equilibrated with 8% acetonitrile containing 0.1% TFA. A gradient from 8% to 30% acetonitrile concentration over 30 min and then to 60%

acetonitrile over twenty min was run at a flow rate of 18 ml/min. The column effluent was collected into polypropylene tubes in 0.2 min fractions and monitored at 220 nm/2.2 AUFS. Fractions were concentrated on a Speed-
5 Vac concentrator (Savant), lyophilized and assayed for antiaggregation activity with human platelets as previously described.

Figure 7 shows that the PAI-containing fraction elutes at 24-25% acetonitrile. Analysis of these active
10 fractions using HPLC with detection at 220 nm showed a symmetric bioactive component, as shown in Figure 8. The amino acid analysis of this material showed a peptide of 71-72 residues, including 12 cysteines, as shown in Table 2.

A portion of the purified peptide was reduced and alkylated with iodoacetamide and purified on a C-18 reverse-phase HPLC column. N-terminal sequence analysis of this material revealed the following amino acid sequence for 23 cycles of Edman degradation: Glu-Ala-Gly-
20 Glu-Glu-Cys-Asp-Cys-Gly-Ser-Pro-Ala-Asn-Pro-Cys-Cys-Asp-Ala-Ala-Thr-Cys-Lys-Leu.

The complete amino acid sequence for this PAI, which was named "tergeminin" is shown in Figure 6.

The purified peptide was tested in the receptor-
25 based assays described in Example 1, paragraph A. Concentrations of pure peptide at less than 100 nM inhibited the binding of Fg and vWF to GP IIb-IIIa and of Vn and vWF to the vitronectin receptor, as shown in Figure 9.

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Example 4

Purification of Platelet Aggregation Inhibitor from Sistrurus milarus barbouri Venom

Two hundred mg of Sistrurus m. barbouri venom
35 (Miami Serpentarium Labs, Lot #SM13SZ) was dissolved in 7.0 ml of 0.5 M acetic acid and applied to a column of

Sephadex G-50 fine (Pharmacia, 2.5 x 100 cm) equilibrated and eluted with 0.5 M acetic acid. The column was run at a flow rate of 26 ml/hr and 5 ml fractions were collected and analyzed for antiplatelet aggregation activity as previously described. Active fractions (41-50) were pooled and lyophilized. This material was redissolved in 2.0 ml 0.5% TFA and loaded onto the preparative C-18 HPLC column as in Example 3 and eluted employing the same gradient conditions. Two-tenths-min fractions from the column were collected into polypropylene tubes, concentrated, lyophilized and analyzed for platelet aggregation inhibitory activity.

Figure 10 shows the activity profile from this HPLC column. The active fractions were subjected to analytical HPLC, which showed several fractions (45-47) which were more than 90% homogeneous. The peptide of fraction 46 (150 ug) was purified to homogeneity on an analytical C-18 column with manual collection of the symmetric peak, as shown in Figure 11. Amino acid analysis of this material showed a peptide of 71-72 amino acids, including 12 cysteine residues, as set forth in Table 2.

The purified peptide (150 ug) was dissolved in 300 ul reaction buffer (6 M guanidine HCl, 0.25 M Tris-HCl, 20 mM EDTA, 20 mM dithiothreitol (DTT), pH 7.5) for 1.5 hours at room temperature to reduce the peptide. This was followed by reaction of 3 ul of 4-vinylpyridine (Aldrich) at room temperature for an additional hour. The reaction was stopped by addition of 200 ul 1% TFA and loaded onto an analytical C-18 HPLC column and eluted with an acetonitrile gradient in water containing 0.1% TFA, starting at 8% acetonitrile and running to 25% acetonitrile in 20 minutes, then to 60% acetonitrile in 10 minutes.

A portion of this pyridylethylated material was submitted to N-terminal sequence analysis, as described above, and exhaustive proteolytic cleavage of the reduced

and alkylated peptide was performed using endoproteinase Lys-C and endoproteinase Asp-N with peptide fragments isolated on either C-3 or C-18 reverse-phase HPLC columns using acetonitrile/water/TFA gradient elution. The amino acid sequence of the N-terminus of the intact peptide and isolated proteolytic fragments were determined as described by Yarden, Y., et al., Nature (1986) 323:226, using automated Edman degradation on a gas-phase sequencer.

The complete amino acid sequence of this isolated peptide, designated "barbourin" is shown in Figure 6, along with the sequences for the proteolytic fragments. A comparison of this sequence with those of other snake venom adhesion inhibitors is shown in Figure 12.

Example 5

Purification of PAI from Lachesis mutas venom

99 mg of Lachesis mutas venom (Miami Serpentarium Labs, Lot #LM15FZ) was dissolved in 2.0 ml of 0.5% trifluoroacetic acid was cooled on ice for 20 min, spun at 14,000 rpm for 3 min to remove insoluble material and loaded onto a 3.9 mm x 30 cm, C-18 Delta Pak reversed-phase HPLC column (Waters) equilibrated with 5% acetonitrile containing 0.1% trifluoroacetic acid. A gradient from 5% to 15% acetonitrile over 5 min and then to 30% over 35 min (2%/min) and continued to 60% acetonitrile over 20 min was run. The flow rate was maintained at 1.5 ml/min and the column effluent monitored at 220 nm/3.0 AUFS. Two minute fractions were collected, concentrated by Speed-Vac and lyophilized. Fractions were assayed for platelet aggregation inhibitory activity.

Figure 13 shows the active fractions which elute at 18% acetonitrile. These fractions were rerun on the C-18 column using a shallower gradient consisting of a 40 min gradient from 5-28% acetonitrile. One-min fractions

were collected, concentrated, lyophilized and assayed for platelet aggregation inhibition activity, with the results shown in Figure 14. These active fractions were run on an analytical C-18 column, and the eluted center peak fraction collected by hand. The eluted material, which is in a single symmetric peak, as shown in Figure 15, was subjected to amino acid analysis and showed a peptide of 72-73 amino acids containing 12 cysteines, as shown in Table 2.

The complete amino acid sequence of this PAI, called "lachesin" is shown in Figure 6.

Example 6

Purification of PAI from *Crotalus viridis viridis* venom

47 mg of *Crotalus viridis viridis* venom (Sigma Chemical Co., Lot #24F-0534) was dissolved in 1 ml of 0.5% trifluoroacetic acid, cooled on ice for 20 min, spun at 14,000 rpm for 3 min to remove insoluble material and loaded onto a 3.9 mm x 30 cm C-18 Delta Pak reverse-phase HPLC column (Waters) equilibrated with 5% acetonitrile containing 0.1% trifluoroacetic acid. A gradient from 5% to 15% acetonitrile over 5 min (2%/min) followed by a gradient from 15% to 30% acetonitrile in 35 min and then to 60% acetonitrile in 60 min was run. A flow rate of 1.5 ml/min was maintained throughout the gradient and the column effluent was collected into polypropylene tubes in 2 min fractions. The column effluent was monitored at 220 nm/3.0 AUFS. Fractions were concentrated, lyophilized and assayed for platelet aggregation inhibitory activity.

The active fractions, shown in Figure 16 as 18-19% acetonitrile, were run on the C-18 HPLC column using a gradient of 8%-20% acetonitrile over 48 min (0.25%/min). The fractions were concentrated and lyophilized and tested for activity; the active fractions were run on a C-18 column using 8-16% acetonitrile over 10 min, 16-20% acetonitrile over 15 min, and then to 60% over 10 min.

The effluent was monitored at 220 nm with individual peaks collected by hand into polypropylene tubes. Reanalysis of the active peak on analytical HPLC gave the results shown in Figure 17. The amino acid analysis conducted on this peak showed a 72-73-residue peptide containing 12 cysteines, as set forth in Table 2. The complete amino acid sequence of this PAI called "viridin" is shown in Figure 6 and is compared to other PAI in Figure 12.

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Table 2 **Amino acid Compositions of Purified Peptides**

	Amino acid	<i>Sistrurus</i> <i>m. bartbouri</i>	<i>Sistrurus</i> <i>c. tergeminus</i>	<i>Lachesis</i> <i>mutas</i>	<i>Crotalus</i> <i>v. viridis</i>	<i>Eristocophis</i> <i>macmahoni</i>
10	Lys	4	3	4	3-4	4
	His	0	0	0-1	1	0
	Arg	4	5	7	5	7
15	Asx	11	11	10	11	7
	Thr	4	4	2	4	2
	Ser	2	2	1	2	1
20	Glx	6-7	5-6	7	6	4
	Pro	4	4	5	6	5
	Gly	9	9	9	10	5
	Ala	7	8	9	7	3
25	Cys	12	12	12	12	7
	Val	2	2	0	1	2
	Met	1	1	0	0	0
	Ile	0	0	2	1	0
30	Leu	3	3	2	3	0
	Tyr	1	1	1	1	1
	Phe	1	1	1	1	1
35		<u>71-72</u>	<u>71-72</u>	<u>72-73</u>	<u>74-75</u>	<u>40</u>

Example 7

Comparison of Purified PAI to Echistatin

The peptides purified as described in Examples 2 and 4, eristicophin and barbourin, were compared to the 49-residue peptide echistatin in inhibiting fibrinogen binding to GP IIB-IIIA, as described in Example 1, paragraph A. Figure 18 shows that these purified PAIs are 2-3 times more potent in this assay than the standard echistatin.

Peptides purified to homogeneity from Echis carinatus, Sistrurus m. barbouri, and Eristicophis macmahoni venoms were compared to echistatin in the ADP-stimulated platelet aggregation assay. Increasing concentrations of purified snake venom peptides were added (without preincubation) at the indicated concentrations (Figure 19). Snake venom peptides from Eristicophis macmahoni and Sistrurus m. barbouri were at least twofold more potent than echistatin, in agreement with their order of potency observed for inhibiting fibrinogen binding to GP IIB-IIIA as presented above.

Example 8

Purification of PAI from

Crotalus cerastes cerastes venom

One gram of Crotalus c. cerastes venom (Miami Serpentarium Labs, Lot #CE4SZ) was dissolved in 7.0 ml of 0.5 M acetic acid and applied to a column of Sephadex G-50 fine (Pharmacia, 2.5 x 100 cm) equilibrated and eluted with 0.5 M acetic acid. The column was run at a flow rate of 25 ml/hr with 5 ml fractions collected into polypropylene tubes. Aliquots of these fractions were assayed for aggregation activity inhibitory activity as previously described. Active fractions (71-80) were pooled and lyophilized. The dried material was resuspended in 2.0 ml of 0.5% TFA, insoluble material

removed by centrifugation and loaded onto the preparative C-18 Waters HPLC column as described in Example 3 and eluted employing the gradient elution conditions described in Example 3. Fractions from the column were collected into polypropylene tubes, concentrated and analyzed for platelet aggregation inhibitory activity. Figure 20 shows the activity profile from this HPLC fractionation.

Active fractions with platelet aggregation inhibitory activity were pooled and lyophilized and rerun on the preparative C-18 HPLC column eluted with the same gradient. Fractions were collected by hand into polypropylene tubes and again assayed for platelet aggregation inhibitory activity as before. Active fractions were analyzed on an analytical C-18 column using the conditions described in Example 4 and homogeneous fractions were pooled and lyophilized. Analytical HPLC analysis of this material is shown in Figure 21.

Purified peptide was subjected to amino acid analysis revealing that it was a peptide of 73-74 amino acids containing 12 cysteine residues, as set forth in Table 3.

The purified peptide (450 ug) was dissolved in 750 ul reaction buffer (6 M guanidine-HCl, 0.25 M Tris-HCl, 20 mM EDTA, 20 mM dithiothreitol (DTT), pH 7.50) for 1.5 hr at room temperature to fully reduce the peptide followed by reaction at room temperature for 1 hr with excess iodoacetamide (Fluka, 16 mg). The reaction was stopped by addition of 500 ul of 1% TFA and loaded onto an analytical C-18 HPLC column and eluted with a gradient of acetonitrile from 8% to 25% in 20 minutes, then to 60% acetonitrile in 10 minutes. The UV absorbing peak was collected by hand into 1.5 ml eppendorf tubes and dried.

A portion of this carboxyamidomethylated peptide was submitted to N-terminal sequence analysis. Exhaustive proteolytic cleavage of the carboxyamidomethylated peptide was performed using endoproteinase Lys-C and

endoproteinase Asp-N. Peptide fragments from these digests were isolated on either C-3 or C-18 reversed-phase HPLC columns using acetonitrile/water/TFA gradient elution conditions. Amino acid sequence was determined as described in Example 4. The complete amino acid sequence determined for "cerastin" is shown in Figure 6, and is compared to that of other PAI in Figure 12.

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Table 1
Amino Acid Compositions

Amino acid	Crotalus c. cerastes	Crotalus atrox	Crotalus d. durissus	Crotalus d. totontatus	Crotalus h. horridus	Bothrops gotiera
Lys	3	3	3	3	4	3
His	0	0	0	0	0-1	0
Arg	3	5	5	5	4	8
Asx	11	10	12	12	10	10
Thr	5	4	4	4	3	2
Ser	1	2	2	2	2	1
Glx	7	6	5-6	5-6	6	8
Pro	5	6-7	4-5	5	7	6
Gly	9	8	10	10	8	8
Ala	7	6	8	8	8	9
Cys	12	12	12	12	10	12
Val	2	2	1	1	3	0
Met	1	0	0	0	1	0
Ile	0	1	1	1	0	1
Leu	3	3	3	3	2-3	2
Tyr	1	1	1	1	1	0
Phe	1	1	1	1	1	2
Trp	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
	73-74	70-71	72-74	73-74	70-72	72

Example 9Purification of PAIfrom Crotalus ruber ruber venom

One gram of Crotalus ruber ruber venom (Miami
5 Serpentarium labs, Lot #CF17SZ) was dissolved in 8 ml of
0.5 M acetic acid and applied to a column of Sephadex G-50
fine (Pharmacia, 2.5 x 100cm) equilibrated at room
temperature and eluted with 0.5 M acetic acid. The column
was run at a flow rate of 25 ml/hr with 5 ml fractions
10 collected into polypropylene tubes. Aliquots of fractions
were assayed for platelet aggregation inhibitory activity
as described. Active fractions (61-70) were pooled and
lyophilized. The dried material was resuspended in 2.0 ml
of 0.5% TFA. Insoluble material was removed by
15 centrifugation and loaded onto a preparative C-18 Water
HPLC column as described and eluted employing the gradient
conditions described in Example 3. Fractions collected
into polypropylene tubes were concentrated on a Speed-Vac
concentrator and analyzed for platelet aggregation
20 inhibitory activity.

Figure 22 shows the activity profile for this
HPLC fractionation. Individual active fractions were
lyophilized. Fractions 49 and 50 were pooled and loaded
onto the analytical C-18 reversed phase column and eluted
25 using conditions described in Example 4 which consisted of
an acetonitrile gradient running from 8% acetonitrile to
25% in twenty minutes followed in ten minutes to 69%
acetonitrile to yield homogeneous peptide which we have
called "ruberin." Automated Edman degradation of
30 carboxyamidomethylated peptide give the sequence shown in
Figure 6.

Example 10Purification of PAI from Crotalus atrox

One gram of Crotalus atrox venom (Miami Serpentarium Labs, Lot #CX16AZ) was dissolved in 10 ml of 0.5 M acetic acid and applied to a column of Sephadex G-50 fine (Pharmacia, 2.5 x 110 cm) equilibrated and run at room temperature with 0.5 M acetic acid. The column was run at a flow rate of 25 ml/hr with 5 ml fractions collected into polypropylene tubes. Aliquots of fractions were assayed for platelet aggregation inhibitory activity as previously described. Active fractions (81-100) were pooled and lyophilized. The dried material was dissolved in 2.0 ml of 0.5% TFA and loaded onto the preparative C-18 HPLC column and run as described in Example 3. Fractions from the column were collected into polypropylene tubes, concentrated on a Speed-vac concentrator and assayed for platelet aggregation inhibitory activity as before. Active fractions were rerun on the analytical C-18 column to yield homogeneous peptide (Figure 23). Amino acid analysis of this material revealed that the peptide contains 72 amino acids including 12 cysteine residues, as shown in Table 3. The amino acid sequence of the isolated peptide, crotatroxin, is shown in Figure 6.

Example 11Purification of PAI from Bothrops cotiara

Six hundred eighty milligrams of Bothrops cotiara venom (Miami Serpentarium Labs, Lot #B05SZ) was dissolved in 10 ml of 0.5 M acetic acid and loaded onto a column of Sephadex G-50 fine (Pharmacia, 2.5 x 110 cm) equilibrated and eluted with 0.5 M acetic acid. The column was run at a flow rate of 25 ml/hr with 5 ml fractions collected into polypropylene tubes. Aliquots of fractions were assayed for platelet aggregation inhibitory activity as described previously. Active fractions (71-90) were pooled and lyophilized. Dried material was

resuspended into 2.0 ml of 0.5% TFA and loaded onto the Waters preparative C-18 reverse-phase column. The column was eluted using the conditions described in Example 3. Fractions were collected into polypropylene tubes, concentrated on a Speed-Vac concentrator and assayed for platelet aggregation inhibitory activity. Active fractions were individually lyophilized. Several peak fractions were rerun on the analytical C-18 column as described in Example 4. The analytical HPLC profile of homogenous peptide is shown in Figure 24. Amino acid analysis of this material reveals this peptide to contain 72 amino acids including 12 cysteine residues, as shown in Table 3. Complete amino acid sequence of this peptide which we have called "cotiarin" is shown in Figures 6 and 12.

The purified peptide was tested in the receptor assays described in Example 1. Initial determinations showed that low concentrations of cotiarin (1-4 nM) selectively inhibited vitronectin binding to vitronectin receptor, whereas the same concentrations had significantly lower inhibiting activity in binding of fibrinogen to GP IIb-IIIa as shown in Figure 25; however, subsequent experiments failed to verify this result.

Example 12

Purification of PAI

from *Crotalus viridis lutosus*

A. One gram of *Crotalus viridis lutosus* venom (Miami Serpentarium Labs, Lot #CL18SZ) was dissolved in 8 ml of 0.5 M acetic acid and applied to a column of Sephadex G-50 fine (Pharmacia, 2.5 x 110 cm) which was equilibrated and eluted with 0.5 M acetic acid. The column was run at a flow rate of 25 ml/hr and 5 ml fractions collected into polypropylene tubes. Aliquots of fractions were assayed for platelet aggregation inhibitory activity. Fractions (71-100) were pooled and lyophilized.

Dried material was resuspended in 2.0 ml of 0.5% TFA. Insoluble material was removed by centrifugation and loaded onto the preparative C-18 Waters reversed-phase column and eluted using the gradient elution conditions described in Example 3. Fractions from the column were collected into polypropylene tubes, concentrated on a Speed-Vac concentrator and analyzed for platelet aggregation inhibitory activity. Active fractions were lyophilized in their individual tubes. Fractions with peak activity were rerun on the Waters analytical C-18 column using the acetonitrile gradient described in Example 4. Fractions were collected by hand into 1.5 ml Eppendorf tubes. Homogeneous fractions were pooled and lyophilized. Analytical HPLC of this material showed a single symmetric peak. The complete amino acid sequence of this peptide which we have called "lutosin" is shown in Figures 6 and 12.

B. In a similar manner to that set forth in paragraph A, the PAIs from B. jararacussu, C. basilicus, C. durissus durissus, C. v. oreganus, C. h. horridus, C. v. helleri, C. durissus totonactus and from C. m. molossus, were isolated and purified. Amino acid compositions for several of these peptides are shown in Table 3. The amino acid sequences of the PAI from C. h. horridus, C. basilicus, C. m. molossus, C. v. oreganus, and C. d. durissus, designated horridin, basilicin, molossin, oreganin, and durissin, respectively, are shown in Figure 6. Receptor binding data for the purified peptides of Examples 1-12 are shown in Figure 26.

In Examples 13-16 below, peptides were synthesized by solid-phase techniques on an Applied Biosystems 431A Peptide Synthesizer using t-Boc amino acids activated as HOBt active esters in accordance with the instructions of the manufacturer, which are briefly as follows for the preparation of Boc-AA1.....AA(n-1)-AA(n)-O-PAM-polystyrene resin.

One-half mmol of selected Boc-AA(n)-O-PAM-polystyrene resin is treated according to the following schedule for incorporation of the Boc-AA(n-1)-OH:

- 1) TFA deprotection: 30% TFA in DCM, 3 min,
5 50% TFA in DCM, 16 min.
- 2) Washes and neutralizations: DCM washes
(5X), 3 min, 5% DIEA in DCM, 2 min, 5% DIEA in NMP, 2 min.
NMP wash (6X), 5 min.
- 3) Coupling: 4 equivalents Boc-AA-HOBt ester
10 in NMP (preactivate 55 min), 38 min, DMSO to make 15%
DMSO/85% NMP, 16 min, 3.8 equiv DIEA, 5 min.
- 4) Wash and resin sample: NMP wash, 3 min.
- 5) Capping: 10% acetic anhydride, 5% DIEA in
NMP, 8 min.
- 15 6) Washes: DCM washes (6X) 4 min.

Example 13

Preparation of Analog #1

[E²⁸L⁴¹C⁶⁴]barbourin (28-73):

20 E-C-A-D-G-L-C-C-D-Q-C-R-F-L-K-K-G-T-V-C-R-V-
A-K-G-D-W-N-D-D-T-C-T-G-Q-S-C-D-C-P-R-N-G-L-Y-G

One-half mmol of PAM-Gly resin (0.6 meq/g, Applied Biosystems, Foster City, CA) was subjected to Procedure A with the required amino acids (introduced in
25 order). The Boc-protected amino acids had the following side-chain protection: Arg(Tos), Asp(OcHex), Cys(4-MeBzl), Glu(OcHex), Lys (Cl-Z), Thr(OBzl), Trp(CHO), and Tyr(Br-Z). Following assembly of the completed protected peptide-resin chain, the amino terminal Boc- group was
30 removed with TFA and the resin dried as its TFA-salt form. The resin (1.3 g) was subjected to "low-high" HF deprotection protocols followed by removal of HF "in vacuo". The dried peptide-resin mixture was transferred to a fritted funnel (coarse) with ethyl ether and was
35 washed several times with alternate washes of ether and

chloroform to remove most of the organic protecting groups and scavengers used in the deprotection.

The peptide mixture was transferred to 2 L of 0.4% acetic acid and the pH adjusted to 7.99 with concentrated NH_4OH . The resin was filtered from this solution and the solution allowed to sit at 4°C without stirring for 20 hr. This was followed by warming the solution to room temperature and storing for 3 days again without stirring. Precipitated material was removed by filtration and the supernatant pH adjusted to 3.0 with acetic acid and lyophilized.

The crude material was dissolved in 8.0 ml of 0.5M acetic acid and loaded onto a Sephadex G-50 fine column (2.5 x 100 cm) equilibrated with 0.5M acetic acid. The column was run at 20 ml/hr and fractions (4 ml) were collected into polypropylene tubes. Aliquots of fractions were dried, resuspended in water and tested for platelet aggregation inhibitory activity as previously described. Active fractions (71-90) were pooled and lyophilized.

Dried material (66 mg) was redissolved in 2.0 ml of 0.1M acetic acid and loaded onto the Waters Preparative C-18 column equilibrated with 8% acetonitrile containing 0.1% TFA. A gradient running from 8% acetonitrile to 20% in 10 minutes followed by a slow gradient to 30% acetonitrile in 40 min was performed. The column was eluted at 18 ml/min and fractions (12 sec) were collected into polypropylene tubes. Fractions were concentrated on a Speed-Vac concentrator to 1.0 ml volume and 10 μl aliquots were tested in the platelet aggregation assay.

Active fractions (29-32) were individually lyophilized and analyzed on the analytical C-18 HPLC column with an 8-30% acetonitrile gradient. Fractions 29 and 30 were pooled and loaded onto the analytical column in 1.0 ml of 0.5% TFA. The major peak was collected manually and lyophilized to yield 1.6 mg of pure peptide.

Amino acid analysis of this material confirmed the identity of the peptide. Assay of this material for its ability to inhibit the binding of fibrinogen to GP IIb-IIIa and vitronectin to VnR is displayed in Figures 26 and 27. These data demonstrate the high affinity of this analog for GP IIb-IIIa and the relative lack of affinity for VnR at concentrations up to 1 uM.

Example 14

Preparation of Analog #2, [K^{29}]eristicophin (4-51):
E-E-P-C-A-T-G-P-C-C-R-R-C-K-F-K-R-A-G-K-V-C-R-
V-A-K-G-D-W-N-N-D-Y-C-T-G-K-S-C-D-C-P-R-N-P-W-N-G

One-half mmol of PAM-Gly resin (0.6 meq/g, Applied Biosystems, Foster City, CA) was subjected to Procedure A with the required amino acids (introduced in order). The Boc-protected amino acids had the following side-chain protection: Arg(Tos), Asp(OcHex), Cys(4-MeBzl), Glu(O-cHex), Lys(Cl-Z), Ser(OBzl), Thr(OBzl), Trp(CHO) and Tyr(Br-Z). Cleavage, refolding and purification of this peptide was identical to the previous examples. Receptor binding data for this analog are shown in Figures 26 and 28.

Example 15

Preparation of Analog #3:
G-C-G-K-G-D-W-P-C-A-NH₂

One-half mmol of PMBHA resin (0.72 meq/g, Applied Biosystems, Foster City, CA) was subjected to Procedure A with the required amino acids (introduced in order). The Boc-protected amino acids had the following side-chain protection: Asp(O-cHex), Cys(4-MeBzl), and Lys(Cl-Z). Following completion of the assembly of the protected peptide-resin, the amino terminal Boc group was removed with TFA and the resin dried as its TFA-salt form. The resin (1.54 g) was treated with anhydrous hydrogen fluoride (HF) containing 10% anisole, 2% ethyl methyl

sulfide for 30 min at -10°C , and an additional 30 min at 0°C . The HF was removed in vacuo and the peptide/resin mixture was suspended in diethyl ether followed by alternately washing with chloroform and ether 3X. After a
5 final ether wash, the peptide was extracted from the resin with 2.0M acetic acid, diluted with distilled water and lyophilized.

The crude peptide (370 mg) was dissolved in deoxygenated 10 mM NH_4OAc , pH 8, to 0.5 mg/ml and allowed
10 to oxidize by dropwise addition of a slight excess of 0.01M potassium ferricyanide ($\text{K}_3\text{Fe}(\text{CN})_6$) solution, stirred an additional 20 min, and adjusted to pH 5 with acetic acid. The peptide solution was treated with DOWEX AG3x4 anion-exchange resin for 15 min with stirring and the
15 resin filtered, diluted with H_2O and lyophilized to yield the crude cyclized peptide. The crude cyclized peptide (392 mg) was purified by desalting on Sephadex G-25F using 0.5M acetic acid as eluent, followed by ion-exchange chromatography on CM-Sepharose (Pharmacia) using an
20 elution gradient generated by addition of 100 mM NH_4OAc to a solution of 10 mM NH_4OAc , pH 4.5. Fractions which had a minimum purity of 90% by HPLC analysis were pooled and lyophilized from H_2O several time to yield 175 mg. Final purification consisted of preparative HPLC purification on
25 a Water C-18 reverse-phase column with an acetonitrile/water/TFA gradient to yield purified peptide. Receptor binding data for this analog are shown in Figures 26, 29 and 30.

30

Example 16

Preparation of Additional Analogs

The following analogs were synthesized; in most cases in a manner similar to that set forth in Example 15. However, analog 60, shown below, was prepared in solution
35 via guanidation of the side chain of the lysine residue of

analog #19 using the procedure of Bajusz, S., et al., FEBS Letts (1980) 110:85-87.

One mg of analog #19 was reacted with 1 mg of 1-amidino-3,5-dimethylpyrazole nitrate (Aldrich) in 1 ml of absolute ethanol in the presence of diisopropylethylamine (DIEA) at room temperature for 4 days. The product analog 60 was purified from excess reagent and starting materials by reversed-phase HPLC on a C-18 column using a gradient of acetonitrile in 0.1% trifluoroacetic acid. Nine hundred ug of this material was isolated in purified form.

- #4 G-C-K-G-D-W-P-C-A-NH₂
- #5 C-G-K-G-D-W-P-C-NH₂
- #6 G-C-G-K-G-D-W-C-A-NH₂
- 15 #7 G-C-K-G-D-W-C-A-NH₂
- #8 Acetyl-C-K-G-D-C-NH₂
- #9 Mpr-K-G-D-Pen-NH₂
- #10 C-K-G-D-W-P-C-NH₂
- #11 Acetyl-C-R-G-D-Pen-NH₂
- 20 #12 C-K-G-D-Y-P-C-NH₂
- #13 C-K-G-D-F-P-C-NH₂
- #19 Mpr-K-G-D-W-P-C-NH₂
- #34 C-K-G-D-W-G-C-NH₂
- #35 C-K-G-E-W-P-C-NH₂
- 25 #36 C-Orn-G-D-W-P-C-NH₂
- #37: C-K-A-D-W-P-C-NH₂
- #38: C-K-A[†]-D-W-P-C-NH₂
- #39: C-K-G-D-W-(Sar)-C-NH₂
- #40: C-K(Formyl)-G-D-W-P-C-NH₂
- 30 #41: C-K-G-D-I-P-C-NH₂
- #42: C-K-G-D-(4-Cl-Phe)-P-NH₂
- #43: C-K-(Sar)-D-W-P-C-NH₂
- #44: C-K-G-D-(4-NO₂-Phe)-P-C-NH₂
- #45: C-K-G-D-(NMePhe)-P-C-NH₂
- 35 #46: C-H-G-D-W-P-C-NH₂
- #47: Acetyl-C-K-G-D-W-P-C-NH₂

- #48: Mpr-K-G-D-W(Formyl)-P-C-NH₂
 #49: Mvl-K-G-D-W-P-C-NH₂
 #50: Mpr-K-G-D-W[†]-P-Pen-NH₂
 #51: Mpr-K-G-D-W-P-Pen-NH₂
 5 #52: Mpr-K-G-D-W-P-Pen[†]-NH₂
 #53: Mpr-K-G-D-W-P[†]-Pen-NH₂
 #54: Mpr-K-G-D[†]-W-P-Pen-NH₂
 #55: Mpr-K-G-D-W-(Thz)-C-NH₂
 #56: Mpr-K-G-D-H(2,4-DNP)-P-C-NH₂
 10 #57: Mpr-K-G-D-(2-Nal)-P-Pen-NH₂
 #58: Mvl-K-G-D-W-P-Pen-NH₂
 #59: Mpr-K-G-D-W-(Pip)-Pen-NH₂
 #60: Mpr-(Har)-G-D-W-P-C-NH₂
 #61: Mpr-K-G-D-W-P-C[†]-NH₂
 15 #62: Mpr-(D-Lys)-G-D-W-P-Pen-NH₂
 #63: Mpr-(Har)-G-D-W-P-Pen-NH₂
 #64: Mpr-(Acetimidyl-Lys)-G-D-W-P-C-NH₂
 #65: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
 #66: Mpr-(N^G,N^{G'}-ethylene-Har)-G-D-W-P-C-NH₂
 20 #67: Mpr-(N^G,N^{G'}-ethylene-Har)-G-D-W-P-Pen-NH₂
 #68: Mpr-Har-Sar-D-W-P-C-NH₂
 #69: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
 #70: Mpr-(Phenylimidyl-Lys)-G-D-W-P-C-NH₂
 #71: Mpr-Har-Sar-D-W-P-PenNH₂
 25 #72: Mpr-(Phenylimidyl-Lys)-G-D-W-P-PenNH₂
 #73: Mpr-Har-G-D-W-(3,4-dehydro P)-C-NH₂

Example 17

30 PAI Activity of Peptides

When tested in the standard aggregation inhibition assays described above, analogs #3-5 had IC₅₀ values of 5 uM for ability to inhibit ADP-induced human platelet aggregation. However, analog #6 has an IC₅₀ of
 35 more than 200 uM, and analog #7, 100 uM. IC₅₀ values for the analogs of the invention in this assay are as follows:

<u>Analog</u>		<u>Sequence</u>	<u>Appr. IC₅₀ (uM)</u>
	#3	G-C-G-K-G-D-W-P-C-A-NH ₂	5
	#4	G-C-K-G-D-W-P-C-A-NH ₂	5
	#5	C-G-K-G-D-W-P-C-NH ₂	5
5	#6	G-C-G-K-G-D-W-C-A-NH ₂	>200
	#7	G-C-K-G-D-W-C-A-NH ₂	100
	#8	Acetyl-C-K-G-D-C-NH ₂	200
	#9	Mpr-K-G-D-Pen-NH ₂	25
	#10	C-K-G-D-W-P-C-NH ₂	5
10	#11	Acetyl-C-R-G-D-Pen-NH ₂	5
	#12	C-K-G-D-Y-P-C-NH ₂	12
	#13	C-K-G-D-F-P-C-NH ₂	20
	#19	Mpr-K-G-D-W-P-C-NH ₂	1
	#34	C-K-G-D-W-G-C-NH ₂	100
15	#35	C-K-G-E-W-P-C-NH ₂	>300
	#36	C-Orn-G-D-W-P-C-NH ₂	150-200
	#37	C-K-A-D-W-P-C-NH ₂	100
	#38	C-K-A [†] -D-W-P-C-NH ₂	>200
	#39	C-K-G-D-W-(Sar)-C-NH ₂	5
20	#40	C-K(Formyl)-G-D-W-P-C-NH ₂	>200
	#41	C-K-G-D-I-P-C-NH ₂	100
	#42	C-K-G-D-(4-Cl-Phe)-P-NH ₂	20
	#43	C-K-(Sar)-D-W-P-C-NH ₂	50
	#44	C-K-G-D-(4-NO ₂ -Phe)-P-C-NH ₂	75
25	#45	C-K-G-D-(NMePhe)-P-C-NH ₂	>200
	#46	C-H-G-D-W-P-C-NH ₂	200
	#47	Acetyl-C-K-G-D-W-P-C-NH ₂	2.5
	#48	Mpr-K-G-D-W(Formyl)-P-C-NH ₂	1
	#49	Mvl-K-G-D-W-P-C-NH ₂	1.5
30	#50	Mpr-K-G-D-W [†] -P-Pen-NH ₂	>200
	#51	Mpr-K-G-D-W-P-Pen-NH ₂	0.75
	#52	Mpr-K-G-D-W-P-Pen [†] -NH ₂	5
	#53	Mpr-K-G-D-W-P [†] -Pen-NH ₂	>200
	#54	Mpr-K-G-D [†] -W-P-Pen-NH ₂	>100
35	#55	Mpr-K-G-D-W-(Thz)-C-NH ₂	2
	#56	Mpr-K-G-D-H(2,4-DNP)-P-C-NH ₂	5

	#57	Mpr-K-G-D-(2-Nal)-P-Pen-NH ₂	1
	#58	Mvl-K-G-D-W-P-Pen-NH ₂	1
	#59	Mpr-K-G-D-W-(Pip)-Pen-NH ₂	1
	#60	Mpr-(Har)-G-D-W-P-C-NH ₂	0.15
5	#61	Mpr-K-G-D-W-P-C [†] -NH ₂	15
	#62	Mpr-K [†] -G-D-W-P-Pen-NH ₂	2.5
	#63	Mpr-(Har)-G-D-W-P-Pen-NH ₂	0.10
	#64	Mpr-(Acetimidyl-Lys)-G-D-W-P-C-NH ₂	0.25
	#68	Mpr-Har-Sar-D-W-P-C-NH ₂	3.0
10	#69	Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH ₂	0.5
	#70	Mpr-(Phenylimidyl-Lys)-G-D-W-P-C-NH ₂	0.5
	#71:	Mpr-Har-Sar-D-W-P-PenNH ₂	2.5
	#72:	Mpr-(Phenylimidyl-Lys)-G-D-W-P-PenNH ₂	0.5

15

Example 18Activity of Linear versus Cyclic Peptides

When tested for inhibition of fibrinogen binding to GP IIb-IIIa in the plate assay, linear RGDW-NH₂ was very similar in activity to cyclic GCGRGDWPCA-NH₂ (Figure 29). In contrast, the linear KGDW-NH₂ was much less potent than cyclic GCGKGDWPCA-NH₂ (Figure 29). For the KGDW compounds, but not the RGDW compounds, cyclization resulted in a marked increase in the ability of the peptide to inhibit the binding of fibrinogen to GP IIb-IIIa.

Example 19Results of Plate BindingAssays for Synthetic Peptides

The peptides synthesized in Example 17, in addition to being assessed for the ability to inhibit platelet aggregation directly, were also tested in the plate assays of the invention as described above. The results for analogs 4-8 are shown in Figure 30. As indicated in the figure, these analogs are differentially

capable, to varying degrees, of inhibiting the binding of fibrinogen to GP IIB-IIIA as compared to vitronectin to vitronectin receptor. Analog #4 appears, among this group, to have the highest differential. Analogs #7 and #5, on the other hand, are also quite specific, and have excellent platelet aggregation inhibition activities.

Example 20

Effects of Purified Peptides on Cell Adhesion

M21 melanoma cells were labelled with ³⁵S-methionine, and then added to vitronectin-coated plates in the presence of the indicated concentrations of purified snake venom peptides. Cell attachment was measured by solubilizing the cells remaining after an incubation and wash, as described in Section C, on page 40. As shown in Figure 31, neither barbourin nor Peptide 1 (truncated barbourin) had a significant effect on cell adhesion to vitronectin, although both are potent inhibitors of platelet aggregation as shown in Examples 2 and 3. In contrast, cotiarin, which is a potent inhibitor of vitronectin binding to the vitronectin receptor, was very potent in inhibiting cell attachment to vitronectin. In similar experiment, Peptide #3, Peptide #3 with K replaced by R (GCGRGDWPCA-NH₂) and RGDS were examined on M21 cell attachment to vitronectin. As shown in Figure 32, RGDS and GCGRGDWPCA-NH₂ are potent inhibitors of cell attachment whereas GCGKGDWPCA-NH₂ was ineffective up to 60 uM.

Example 21

Comparison of Analogs 60 and 19

Analogs 60 and 19 described above are peptides of the invention containing the sequence K*GD_X and are identical except for the embodiment of K*. Analog 60 is of the formula:

Mpr-(Har)-G-D-W-P-C-NH₂;

analog 19 is of the formula:

5 Mpr-K-G-D-W-P-C-NH₂.

These analogs were tested by standard platelet aggregation inhibition assays and using the cell adhesion assay of Example 20 above. The results are shown in 10 Figures 33 and 34. As shown in Figure 33, analog #60 is efficient at vanishingly small concentrations in inhibiting platelet aggregation, and is relatively less effective in preventing cell adhesion to vitronectin. Figure 34 shows analog #19 has good platelet aggregation 15 inhibition activity as well as specificity; however, it is less active in the platelet aggregation inhibition assay than its analog #60 counterpart. Analog #60 has an IC₅₀ in platelet aggregation of approximately 0.15 nM; analog #19 has an IC₅₀ of approximately 1 nM.

20

Example 22

Folt's Model of Thrombosis in Dog Coronary Artery

25 A. Initiation of cyclic flow reductions (CFRs)
in open chest dog. An occluder placed on the left anterior descending (LAD) coronary artery of a 20 kg dog, as previously described was performed. The phasic and mean blood flows as measured by an electromagnetic (EM) flow probe, and Doppler flow probe are shown in Figure 35.

30

B. Effect of Cyclic GCGKGDWPCA-NH₂ (Analog #3)
on the CFRs in the open chest dog. A dose of 10 mg of this peptide was infused into a peripheral vein in the dog. Shown in Figure 36 are the blood flow patterns in 35 the LAD, as described above. Note the partial ablation of the CFRs, as seen in the decreased slope of the flow

reductions. Note also that flow is not reduced to the same degree as in the control (A).

C. A second infusion of 40 mg of Analog #3 was given into a peripheral vein. As shown in Figure 37 the complete ablation of the CFRs indicates that full flow has been restored in the LAD.

Example 23

10 Construction of Expression Vectors for Barbourin Peptides

A gene encoding the full length [L^{41}] barbourin peptide (1-73) was assembled from synthetic oligonucleotides as shown in Figure 38, which were kinased, annealed and ligated into EcoRI-HindIII digested M13mp18 using standard procedures. The bacterial alkaline phosphatase gene (phoA) signal sequence (Watson, M.E.E., Nucleic Acids Research (1984) 12:5145) was added to the barbourin construct by ligating synthetic oligonucleotides into the EcoRI/NcoI sites of the [L^{41}] barbourin (1-73) construct as shown in Figure 39. The nucleotide sequences of all constructs were verified by the Sanger dideoxy chain termination method.

A truncated version of this peptide was also constructed from synthetic oligonucleotides which would encode only amino acids 28-73 of the full length molecule. Two alterations, Q^{28} to E^{28} and A^{64} to C^{64} were introduced using site directed mutagenesis as described by Kunkel et al. Meth Enzymol (1987) 154:367. The phoA signal sequence was added to the truncated version as described above (Figure 40). In addition, the signal sequence for the E. coli heat-stable enterotoxin II (Picken, R.W., et al. Infect Immun (1983) 42:269) was added to the truncated version using synthetic oligonucleotides with EcoRI and NcoI compatible ends. All bacterial secretion constructs were subcloned into the bacterial expression vector pPROK-1 (Brosius, J., Gene (1984) 27:151, ibid:161), available

commercially from CLONTECH Lab, Inc. using EcoRI and HindIII restriction endonucleases.

A gene encoding tandem repeats of the desired title peptide was prepared using the polymerase chain reaction (PCR) to produce the multimerization unit from the full-length barbourin peptide 1-73 containing L41 and C64.

Figure 41 shows the oligonucleotides used for the PCR synthesis. The PCR reaction was conducted according to the method of Saiki, R.K., et al. Science (1988) 239:487. The resulting polymer junction contains methionines at either end of the sequence as shown in Figure 42 and provides desirable restriction sites for the construct.

The tandem repeats are formed from the individual multimer-forming components by, for example, ligating an EcoRI/BamHI fragment to a BglII/HindIII fragment in an M13mpl8 vector cut with EcoRI/HindIII to form a dimer. The resultant dimer is excised with EcoRI and BamHI and religated to a BglII/HindIII fragment to produce a trimer, and so on until the desired size is obtained. This construction is diagramed in Figure 43.

The multimer was then ligated into the E. coli vector pKK233-2, Amann, E., et al., Gene (1985) 40:183, available from Clontech, by digesting the vector with NcoI/HindIII and ligating a monomer subfragment of NcoI/BamHI and multimer subfragments of BglII/HindIII.

For expression as a fusion protein, the above digested vector was used along with an NcoI-EcoRI subfragment containing a slightly modified amino-terminal portion (amino acids 1 to 72) of the chloramphenicol acetyltransferase gene (Chang, C.N., et al. Gene (1987) 55:189) and EcoRI-HindIII subfragments of the multimer constructions.

Example 24Expression of Recombinant Genes

Protein expression from all of the recombinant plasmids described above is induced according to Kanamari et al. Gene (1988) 66:295 after transfection into appropriate E. coli host strains. Products are characterized by sodium dodecyl sulfate polyacrylamide gel electrophoresis and by their ability to inhibit ADP-induced platelet aggregation in platelet-rich plasma.

10 Following purification, the multimeric proteins are converted to monomer units with cyanogen bromide cleavage and the products assayed as above.

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CLAIMS

1. A method to determine the presence or absence of platelet aggregation inhibitor (PAI) activity
5 in a biological fluid, which method comprises:

contacting a sample of said fluid with purified platelet GP IIB-IIIa in the presence of a solution of fibrinogen (Fg) or von Willebrand Factor (vWF) under conditions wherein Fg or vWF binds to said GP IIB-IIIa,
10 and

detecting the decrease or lack of decrease in the binding of Fg or vWF to GP IIB-IIIa in comparison to the binding of Fg or vWF to GP IIB-IIIa in a control which does not contain biological fluid.
15

2. The method of claim 1 which further includes at least one assay selected from the group consisting of:

determining the inability or ability of said fluid to inhibit binding of vitronectin to vitronectin
20 receptor;

determining the inability or ability of said fluid to inhibit the binding of fibronectin to fibronectin receptor;

determining the inability or ability of said
25 fluid to inhibit the binding of fibronectin to GP IIB-IIIa receptor; and

determining the inability or ability of said fluid to inhibit the binding of von Willebrand Factor to the vitronectin receptor.
30

3. A purified platelet aggregation inhibitor (PAI) obtainable from snake venom characterized by the specific ability of said PAI to inhibit binding of fibrinogen and/or von Willebrand Factor to GP IIB-IIIa and
35 relative inability to inhibit binding of vitronectin to

vitronectin receptor or to inhibit binding of fibronectin to fibronectin receptor.

4. A purified and isolated PAI or a modified
5 and/or truncated form thereof which is obtainable from a
snake venom identified as active by the method of claim 1
with the proviso that said venom is not Echis carinatus,
Trimeresurus gramineus, T. flavoviridis, T. Elegans, T.
albolabris, Bitis arietans, Bothrops atrox, Agkistrodon
10 halys, Agkistrodon p. piscivorus, or Agkistrodon
rhodostoma.

5. The PAI of claim 4 wherein said snake venom
is selected from the group consisting of Echis colorata,
15 Eristicophis macmahonii; A. hypnale, A. acutus, A.
piscivorus leucostoma, A. piscivorus conanti; Bothrops
asper; Bothrops cotiara, B. jararaca, B. jararacussu, B.
lansbergi, B. medusa, B. nasuta, B. neuwiedi, B. pradoi,
B. schlegli; Crotalus atrox, C. basilicus, C. cerastes
20 cerastes, C. durissus durissus, C. durissus totonatacus,
C. horridus horridus, C. molossus molossus, C. ruber
ruber, C. viridis cereberus, Crotalus v. helleri, Crotalus
v. lutosus, Crotalus v. oreganus, Crotalus v. viridis;
Lachesis mutas; Sistrurus catenatus tergeminus, and
25 Sistrurus milarus barbouri.

6. The PAI of claim 5 wherein the snake venom
is selected from the group consisting of Eristicophis
macmahonii (eristicophin); Bothrops cotiara (cotiarin); B.
30 jararacussu; Crotalus atrox (cotratroxin); Crotalus
basilicus (basilicin); C. cerastes cerastes (cerastin); C.
durissus totonatacus; C. h. horridus (horridin); Crotalus
m. molossus (molossin); C. v. helleri; C. ruber ruber
(ruberin); Crotalus viridis lutosus (lutosin); C. v.
35 viridis (viridin); C. v. oreganus (oreganin); C. durissus
durissus (durissin); Lachesis mutas (lachesin); Sistrurus

catenatus tergeminus (tergemin); and S. milarus barbouri (barbourin).

7. The PAI of claim 4 which has been isolated
5 and purified from snake venom by a process which comprises the steps of:

applying said venom to a sizing column to obtain
a solution containing components of molecular weight
10 <10kd, followed by

subjecting said fraction to reverse phase HPLC
and obtaining a multiplicity of fractions, and

recovering those fractions which inhibit the
binding of fibrinogen to GP IIB-IIIA.

8. The PAI of claim 7 wherein the <10kd frac-
15 tion is obtained by subjecting the venom to column
chromatography using a sizing gel and eluting a multiplicity of fractions, and

isolating the fractions which inhibit fibrinogen
20 binding to GP IIB-IIIA.

9. A method to purify PAI from snake venom,
which method comprises

applying said venom to a sizing column to obtain
25 a solution containing components of molecular weight
<10kd, followed by

subjecting said fraction to reverse phase HPLC
and obtaining a multiplicity of fractions, and

recovering those fractions which inhibit the
30 binding of fibrinogen to GP IIB-IIIA.

10. The method of claim 9 wherein the <10kd
fraction is obtained by subjecting the venom to column
chromatography using a sizing gel and eluting a multiplicity of fractions, and
35

isolating the fractions which inhibit fibrinogen binding to GP IIb-IIIa.

11. A pharmaceutical composition which
5 comprises an amount of the PAI of claim 3 effective to prevent thrombus formation, in admixture with a pharmaceutically acceptable excipient.
12. A method to inhibit thrombus formation in
10 animal subjects which method comprises administering to a subject in need of such treatment an effective amount of the PAI of claim 3 or a pharmaceutical composition thereof.
13. A method for treating a patient suspected
15 of having a platelet-associated ischemic syndrome comprising administering to the patient a therapeutically effective dose of the PAI of claim 3.
14. A method for preventing platelet loss during
20 extracorporeal circulation of the blood which comprises contacting the PAI of claim 3 with the blood as it is withdrawn from a subject.
15. A pharmaceutical composition which
25 comprises an amount of the PAI of claim 4 effective to prevent thrombus formation, in admixture with a pharmaceutically acceptable excipient.
16. A method to inhibit thrombus formation in
30 animal subjects which method comprises administering to a subject in need of such treatment an effective amount of the PAI of claim 4 or a pharmaceutical composition thereof.

35

17. A method for treating a patient suspected of having a platelet-associated ischemic syndrome comprising administering to the patient a therapeutically effective dose of the PAI of claim 4.

5

18. A method for preventing platelet loss during extracorporeal circulation of the blood which comprises contacting the PAI of claim 4 with the blood as it is withdrawn from a subject.

10

19. A DNA in isolated and purified form which encodes the PAI of claim 4.

20. A recombinant expression system capable, when transformed into compatible host, of expressing the DNA of claim 19.

21. A recombinant host transformed with the expression system of claim 20.

20

22. A method to produce PAI which method comprises:

culturing the host of claim 21 under conditions effective to express the PAI-encoding DNA to produce PAI; and recovering the PAI from the culture

25

23. A platelet aggregation inhibitor in purified and isolated form which PAI can be isolated from snake venom and which contains, as the inhibitory sequence, the amino acid sequence Lys-Gly-Asp (KGD).

30

24. The platelet aggregation inhibitor of claim 23 comprising the following amino acid sequence:

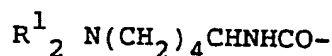
35

1
 Glu-Ala-Gly-Glu-Glu-Cys-Asp-Cys-Gly-Ser-Pro-Glu-Asn-Pro-
 20
 Cys-Cys-Asp-Ala-Ala-Thr-Cys-Lys-Leu-Arg-Pro-Gly-Ala-Gln-
 30
 Cys-Ala-Asp-Gly-Leu-Cys-Cys-Asp-Gln-Cys-Arg-Phe-Met-Lys-
 40
 5 Lys-Gly-Thr-Val-Cys-Arg-Val-Ala-Lys-Gly-Asp-Trp-Asn-Asp-
 50
 60
 Asp-Thr-Cys-Thr-Gly-Gln-Ser-Ala-Asp-Cys-Pro-Arg-Asn-Gly-
 70
 Leu-Tyr-Gly

10 or a modified and/or truncated form thereof.

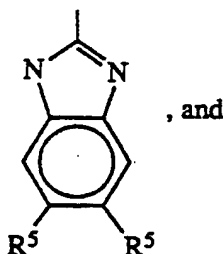
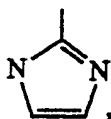
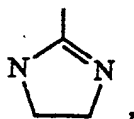
25. A composition of matter which consists
 essentially of a specific platelet aggregation inhibitor
 (PAI) capable of inhibiting binding of Fg or vWF to GP
 15 IIb-IIIa with substantially more potency than of
 inhibiting binding of vitronectin to vitronectin receptor
 or fibronectin to fibronectin receptor, which PAI
 comprises the amino acid sequence $K^*(Sar/G)D$ wherein K^* is
 a lysyl residue of the formula

20

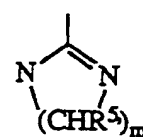


wherein each R^1 is independently H, alkyl(1-6C)
 or at most one R' is $R^2-C=NR^3$,

25 wherein R^2 is H, alkyl(1-6C) or is a substituted
 or unsubstituted phenyl or benzyl residue, or is NR^4_2 in
 which each R^4 is independently H or alkyl(1-6C), and
 R^3 is H, alkyl(1-6C), phenyl or benzyl, or
 $R^2-C=NR^3$ is a radical selected from the group consisting
 30 of:



, and



35

where m is an integer of 2-3, and each R⁵ is independently H or alkyl(1-6C); and

wherein one or two (CH₂) may be replaced by O or S provided said O or S is not adjacent to another heteroatom; and

wherein said K* (Sar/G)D is contained in a cyclic peptide of at least 8 amino acid residues.

26. The PAI of claim 25 which has the primary structure of a naturally occurring PAI containing RGD or KGD or a truncated and/or modified form thereof, wherein the R residue in the sequence RGD occurring in said PAI or truncated and/or modified form thereof is replaced by K*, or wherein the K in the sequence KGD occurring in said PAI or truncated and/or modified form thereof is replaced by an embodiment of K* which is not K.

27. The composition of claim 26 wherein said naturally occurring PAI is present in snake venom.

28. The composition of claim 27 wherein said PAI is selected from the group consisting of trigramin, echistatin, elegantin, albolabrin, lachesin, flavoviridin, eristicophin, tergeminin, rhodastomin, applaggin, halysin, bitistatin, ruberin, cerastin, cotiarin, crotatroxin, horridin, basilicin, lutosin, molossin, durissin, jararacin, cereberin, oreganin, viridin, and barbourin.

29. A recombinant expression system capable, when transformed into a compatible host, of expressing a DNA encoding a specific platelet aggregation inhibitor (PAI) capable of inhibiting binding of Fg or vWF to GP IIb-IIIa with substantially more potency than of inhibiting binding of vitronectin to vitronectin receptor or fibronectin to fibronectin receptor, which PAI comprises

the amino acid sequence KGD contained in a peptide sequence of at least 8 amino acid residues.

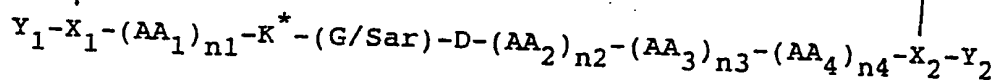
30. A recombinant host transformed with the expression system of claim 29.

31. A method to produce a GP IIb-IIIa specific PAI which method comprises:

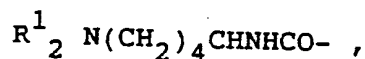
culturing the host of claim 30 under conditions effective to express the PAI-encoding DNA to produce PAI; and

recovering the PAI from the culture.

32. A composition of matter which consists essentially of a specific platelet aggregation inhibitor (PAI) capable of inhibiting binding of Fg or vWF to GP IIb-IIIa with substantially more potency than of inhibiting binding of vitronectin to vitronectin receptor or fibronectin to fibronectin receptor, which PAI has the formula



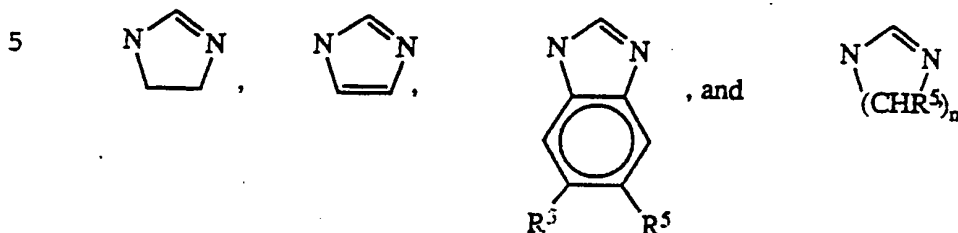
wherein K^* is a lysyl residue of the formula



wherein each R^1 is independently H, alkyl (1-6C), or at most one R^1 is $R^2-C=NR^3$,

wherein R^2 is H, alkyl(1-6C) or is a substituted or unsubstituted phenyl or benzyl residue, or is NR^4_2 in which each R^4 is independently H or alkyl(1-6C), and R^3 is H, alkyl(1-6C), phenyl or benzyl, or

$R^2-C=NR^3$ is a radical selected from the group consisting of:



where m is an integer of 2-3, and each R^5 is independently H or alkyl(1-6C);

and wherein one or two (CH_2) may be replaced by O or S provided said O or S is not adjacent to another
15 heteroatom;

AA_1 is a small, neutral (polar or nonpolar) amino acid and n_1 is an integer of 0-3;

AA_2 is a neutral, nonpolar large (aromatic or nonaromatic) or a polar aromatic amino acid and n_2 is an
20 integer of 0-3;

AA_3 is a proline residue or a modified proline residue (as defined below) and n_3 is an integer of 0-1;

AA_4 is a neutral, small amino acid or the N-alkylated form thereof and n_4 is an integer of 0-3;
25

each of X_1 and X_2 is independently a residue capable of forming a bond between X_1 and X_2 to obtain a cyclic compound as shown; and

each of Y_1 and Y_2 is independently a noninterfering substituent or may be absent;
30

wherein one or more peptide linkages may optionally be replaced by a linkage selected from the group consisting of $-CH_2NH-$, $-CH_2S-$, CH_2CH_2- , $-CH=CH-$ (cis and trans), $-COCH_2-$, $-CH(OH)CH_2-$ and $-CH_2SO-$;

with the proviso that if n_3 is 0; either:
35

1) the sum of n_2 and n_4 must be at least 2; or

- 2) K^* must be other than Har or K; or
- 3) at least one of X_1 and X_2 must be other than cys (C), penicillamine (Pen), or 2-amino-3,3-cyclopentanemethylene-3-mercaptopropionic acid (APmp); or
- 5 4) Y_1 or Y_2 must comprise at least one amino acid residue; or
- 5) one or more peptide linkages is replaced by said alternate linkage.
- 10 33. The composition of claim 32 wherein Y_1 is H, acyl, or a peptide residue or derivatized form thereof or is absent and Y_2 is OH, NH_2 or a peptide residue or derivatized form thereof or is absent.
- 15 34. The composition of claim 33 wherein Y_2 is NH_2 -A- NH_2 or is absent.
- 20 35. The composition of claim 33 wherein Y_1 is H, acetyl, G or is absent.
36. The composition of claim 32 wherein X_1 and X_2 are selected from the group consisting of cysteine (C), mercaptopropionyl (Mpr) and penicillamine (Pen).
- 25 37. The composition of claim 32 wherein AA_1 is G and n_1 is 0 or 1.
- 30 38. The composition of claim 32 wherein AA_2 is selected from the group consisting of W, F, L, Y, and V.
39. The composition of claim 38 wherein AA_2 is W.
- 35 40. The composition of claim 32 wherein K^* is K, Har, acetimidyl-Lys or phenylimidyl-Lys.

41. The composition of claim 40 which is selected from the group consisting of

- PAI 1: E-C-A-D-G-L-C-C-D-Q-C-R-F-L-K-K-G-T-V-
5 C-R-V-A-K-G-D-W-N-D-D-T-C-T-G-Q-S-C-D-C-P-R-N-G-L-Y-G
- PAI 2: E-E-P-C-A-T-G-P-C-C-R-R-C-K-F-K-R-A-G-
K-V-C-R-V-A-K-G-D-W-N-N-D-Y-C-T-G-K-S-C-D-C-P-R-N-P-W-N-G
- PAI 3: G-C-G-K-G-D-W-P-C-A-NH₂;
- 10 PAI 4: G-C-K-G-D-W-P-C-A-NH₂
- PAI 5: C-G-K-G-D-W-P-C-NH₂
- PAI 7: C-K-G-D-W-C-A-NH₂;
- PAI 9: Mpr-K-G-D-Pen-NH₂
- 15 PAI 10: C-K-G-D-W-P-C-NH₂
- PAI 12: C-K-G-D-Y-P-C-NH₂
- PAI 13: C-K-G-D-F-P-C-NH₂
- PAI 14: C-K-G-D-L-P-C-NH₂
- 20 PAI 15: C-K-G-D-V-P-C-NH₂
- PAI 16: C-K-G-D-Y(OMe)-P-C-NH₂
- PAI 17: C-K-G-D-(2-Nal)-P-C-NH₂
- PAI 18: C-K-G-D-(Cha)-P-C-NH₂
- 25 PAI 19: Mpr-K-G-D-W-P-C-NH₂
- PAI 20: Mpr-K-G-D-Y-P-C-NH₂
- PAI 21: Mpr-K-G-D-F-P-C-NH₂
- 30 PAI 22: Mpr-K-G-D-L-P-C-NH₂
- PAI 23: Mpr-K-G-D-V-P-C-NH₂
- PAI 24: Mpr-K-G-D-Y(OMe)-P-C-NH₂
- PAI 25: Mpr-K-G-D-(2-Nal)-P-C-NH₂
- 35 PAI 26: Mpr-K-G-D-(Cha)-P-C-NH₂
- PAI 27: cyclo(G-K-G-D-W-P)

	PAI 28:	cyclo(A-K-G-D-W-P)
	PAI 29:	cyclo(A [†] -K-G-D-W-P)
	PAI 30:	cyclo(F-K-G-D-W-P)
5	PAI 31:	cyclo(beta-Ala-K-G-D-W-P)
	PAI 32:	cyclo(gamma-Abu-K-G-D-W-P)
	PAI 33:	cyclo(R-K-G-D-W-P)
10	PAI 34:	C-K-G-D-W-G-C-NH ₂
	PAI 37:	C-K-A-D-W-P-C-NH ₂
	PAI 39:	C-K-G-D-W-(Sar)-C-NH ₂
	PAI 41:	C-K-G-D-I-P-C-NH ₂
15	PAI 42:	C-K-G-D-(4-Cl-Phe)-P-NH ₂
	PAI 43:	C-K-(Sar)-D-W-P-C-NH ₂
	PAI 44:	C-K-G-D-(4-NO ₂ -Phe)-P-C-NH ₂
	PAI 47:	Acetyl-C-K-G-D-W-P-C-NH ₂
20	PAI 48:	Mpr-K-G-D-W(Formyl)-P-C-NH ₂
	PAI 49:	Mvl-K-G-D-W-P-C-NH ₂
	PAI 51:	Mpr-K-G-D-W-P-Pen-NH ₂
	PAI 52:	Mpr-K-G-D-W-P-Pen [†] -NH ₂
25	PAI 54:	Mpr-K-G-D [†] -W-P-Pen-NH ₂
	PAI 55:	Mpr-K-G-D-W-(Thz)-C-NH ₂
	PAI 56:	Mpr-K-G-D-H(2,4-DNP)-P-C-NH ₂
30	PAI 57:	Mpr-K-G-D-(2-Nal)-P-Pen-NH ₂
	PAI 58:	Mvl-K-G-D-W-P-Pen-NH ₂
	PAI 59:	Mpr-K-G-D-W-(Pip)-Pen-NH ₂
	PAI 60:	Mpr-(Har)-G-D-W-P-C-NH ₂
35	PAI 61:	Mpr-K-G-D-W-P-C [†] -NH ₂
	PAI 62:	Mpr-K [†] -G-D-W-P-Pen-NH ₂

- PAI 63: Mpr-(Har)-G-D-W-P-Pen-NH₂
PAI 64: Mpr-(Acetimidyl-Lys)-G-D-W-P-C-NH₂
PAI 65: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
PAI 66: Mpr (N^G,N^{G'}-ethylene-Har)-G-D-W-P-C-NH₂
PAI 67: Mpr (N^G,N^{G'}-ethylene-Har)-G-D-W-P-Pen-NH₂
PAI 68: Mpr-Har-Sar-D-W-P-C-NH₂
PAI 69: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
PAI 70: Mpr-(Phenylimidyl-Lys)-G-D-W-P-C-NH₂
PAI 71: Mpr-Har-Sar-D-W-P-PenNH₂
PAI 72: Mpr-(Phenylimidyl-Lys)-G-D-W-P-PenNH₂
PAI 73: Mpr-Har-G-D-W-(3,4-dehydro-Pro)-C-NH₂
PAI 74: Mpr-Har-G-D-Pen-NH₂
PAI 75: Mpr-(Phenylimidyl-Lys)-G-D-Pen-NH₂

42. The composition of claim 41 which is selected from the group consisting of

- PAI 3: G-C-G-K-G-D-W-P-C-A-NH₂;
PAI 4: G-C-K-G-D-W-P-C-A-NH₂;
PAI 5: C-G-K-G-D-W-P-C-NH₂;
PAI 9: Mpr-K-G-D-Pen-NH₂; and
PAI 10: C-K-G-D-W-P-C-NH₂
PAI 12: C-K-G-D-Y-P-C-NH₂
PAI 13: C-K-G-D-F-D-C-NH₂
PAI 19: Mpr-K-G-D-W-P-C-NH₂
PAI 25: Mpr-K-G-D-(2-Nal)-P-C-NH₂
PAI 34: C-K-G-D-W-G-C-NH₂
PAI 39: C-K-G-D-W-(Sar)-C-NH₂

- PAI 42: C-K-G-D-(4-Cl-Phe)-P-NH₂
PAI 43: C-K-(Sar)-D-W-P-C-NH₂
PAI 44: C-K-G-D-(4-NO₂-Phe)-P-C-NH₂
PAI 47: Acetyl-C-K-G-D-W-P-C-NH₂
PAI 48: Mpr-K-G-D-W(Formyl)-P-C-NH₂
PAI 49: Mvl-K-G-D-W-P-C-NH₂
PAI 51: Mpr-K-G-D-W-P-Pen-NH₂
PAI 52: Mpr-K-G-D-W-P-Pen[†]-NH₂
PAI 55: Mpr-K-G-D-W-(Thz)-C-NH₂
PAI 56: Mpr-K-G-D-H(2,4-DNP)-P-C-NH₂
PAI 57: Mpr-K-G-D-(2-Nal)-P-Pen-NH₂
PAI 58: Mvl-K-G-D-W-P-Pen-NH₂
PAI 59: Mpr-K-G-D-W-(Pip)-Pen-NH₂
PAI 60: Mpr-(Har)-G-P-W-P-C-NH₂
PAI 61: Mpr-K-G-D-W-P-C[†]-NH₂
PAI 62: Mpr-K[†]-G-D-W-P-Pen-NH₂
PAI 63: Mpr-(Har)-G-D-W-P-Pen-NH₂
PAI 64: Mpr-(Acetimidyl-Lys)-G-D-W-P-C-NH₂
PAI 65: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
PAI 66: Mpr (N^G,N^{G'}-ethylene-Har)-G-D-W-P-C-NH₂
PAI 67: Mpr (N^G,N^{G'}-ethylene-Har)-G-D-W-P-Pen-NH₂
PAI 68: Mpr-Har-Sar-D-W-P-C-NH₂
PAI 69: Mpr-(Acetimidyl-Lys)-G-D-W-P-Pen-NH₂
PAI 70: Mpr-(Phenylimidyl-Lys)-G-D-W-P-C-NH₂
PAI 71: Mpr-Har-Sar-D-W-P-PenNH₂
PAI 72: Mpr-(Phenylimidyl-Lys)-G-D-W-P-PenNH₂
PAI 73: Mpr-Har-G-D-W-(3,4-dehydro P)-C-NH₂.

43. A pharmaceutical composition which comprises an amount of the PAI of claim 25 effective to prevent thrombus formation, in admixture with a pharmaceutically acceptable excipient.

44. A method to inhibit thrombus formation in animal subjects which method comprises administering to a subject in need of such treatment an effective amount of the PAI of claim 25 or a pharmaceutical composition thereof.

45. A method for treating a patient suspected of having a platelet-associated ischemic syndrome comprising administering to the patient a therapeutically effective dose of the PAI of claim 25.

46. A method for preventing platelet loss during extracorporeal circulation of the blood which comprises contacting the PAI of claim 25 with the blood as it is withdrawn from a subject.

47. A method to inhibit metastases in a carcinoma patient which method comprises administering to said patient a therapeutically effective dose of cotiarin or Vn/VnR inhibiting fragment thereof.

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INHIBITION OF FIBRINOGEN BINDING TO GPIIb-IIIa BY SNAKE VENOM

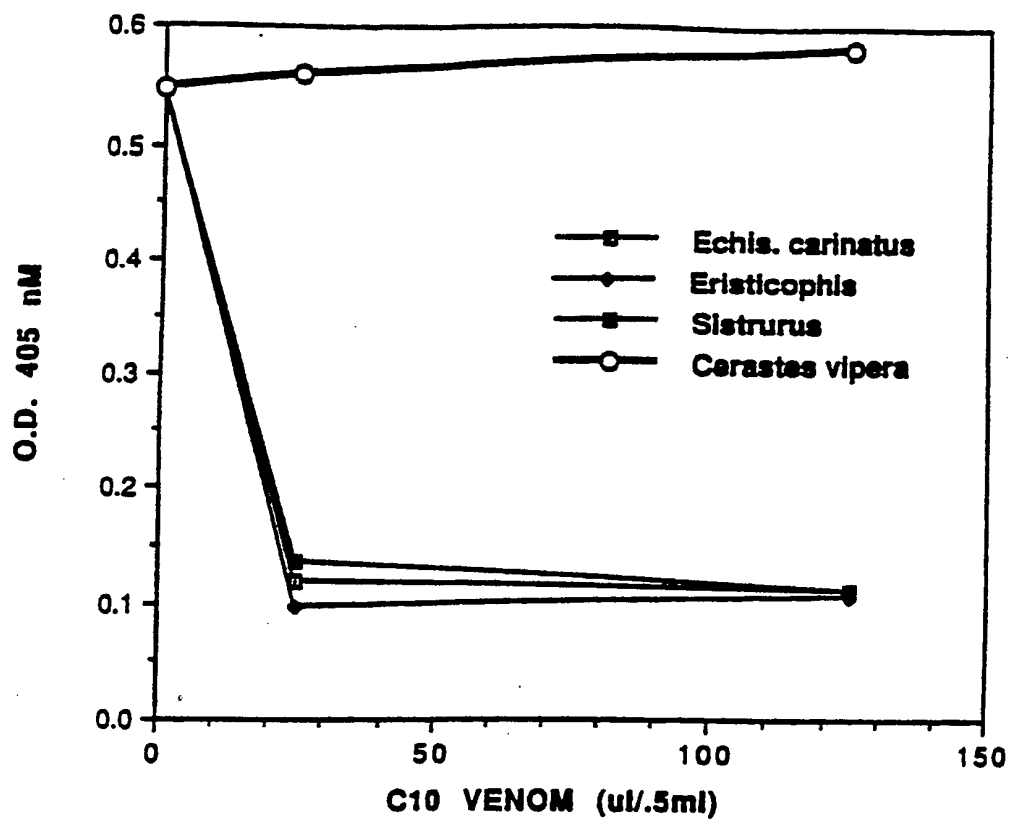


Figure 1

Selectivity of Snake Venom Peptides In
GPIIb-IIIa and Vitronectin Receptor Binding Assays

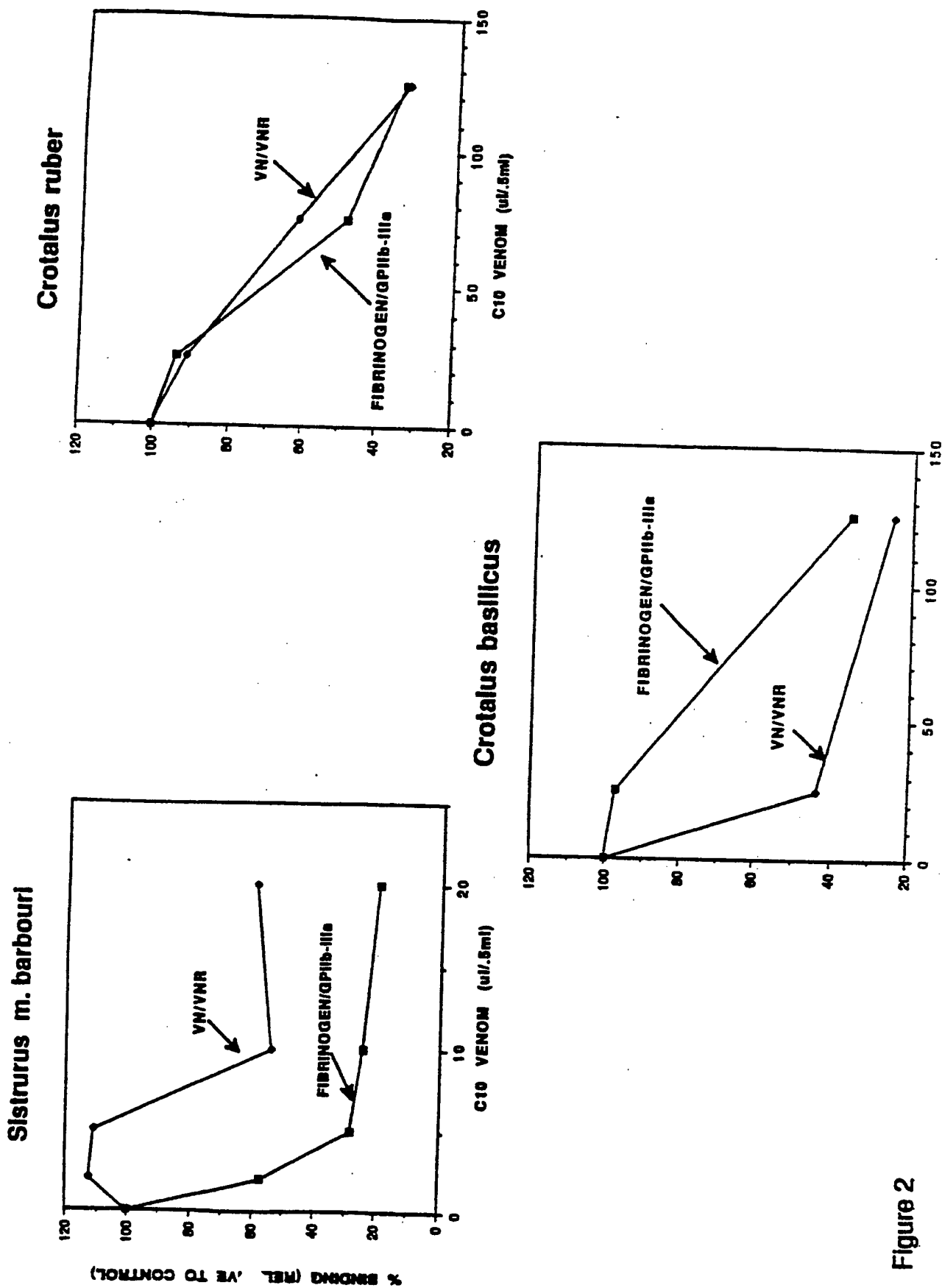
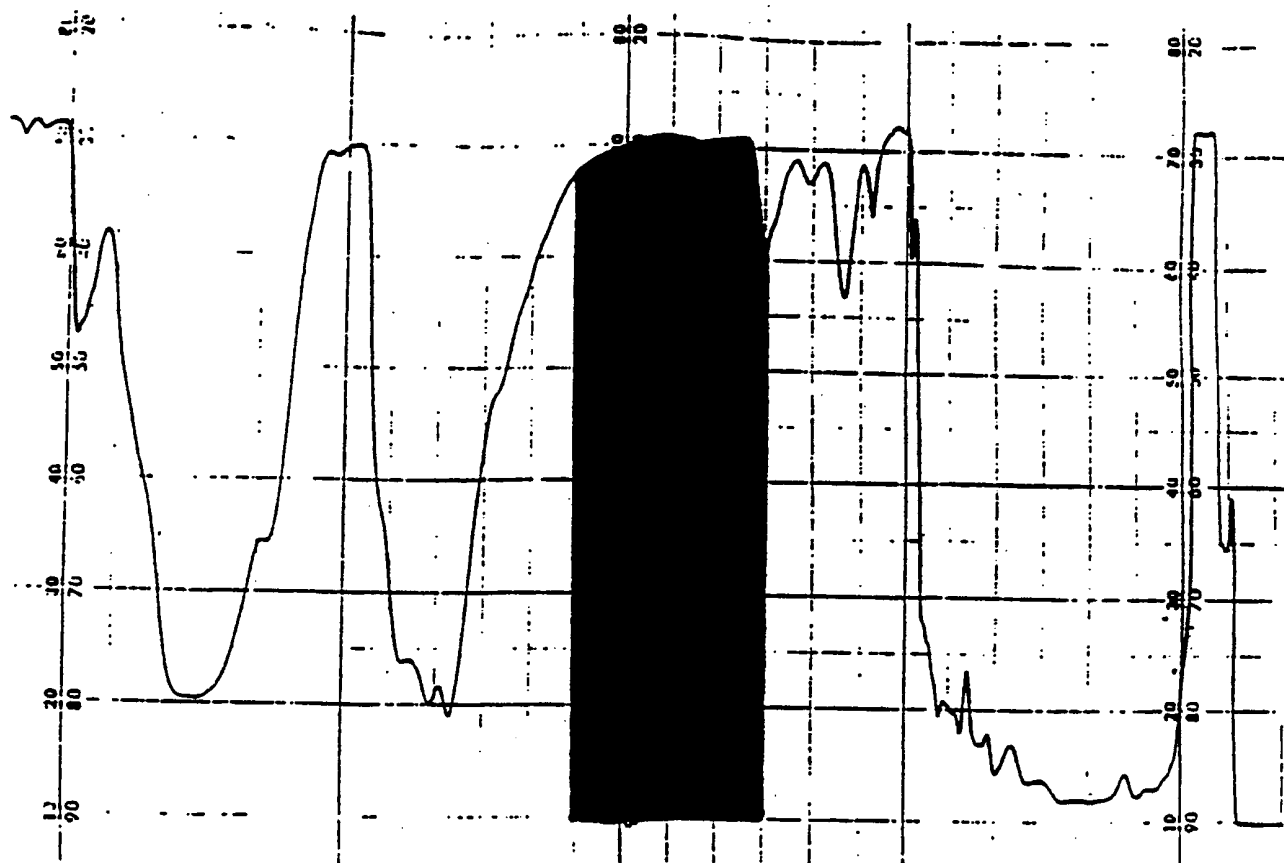


Figure 2

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HPLC Purification of Eristicophis Macdonaldi Platelet Aggregation Inhibitor**Figure 3**

- 4 / 47

**Slow gradient HPLC purification of Erisilcophis Macmahoni
Platelet Aggregation Inhibitor**

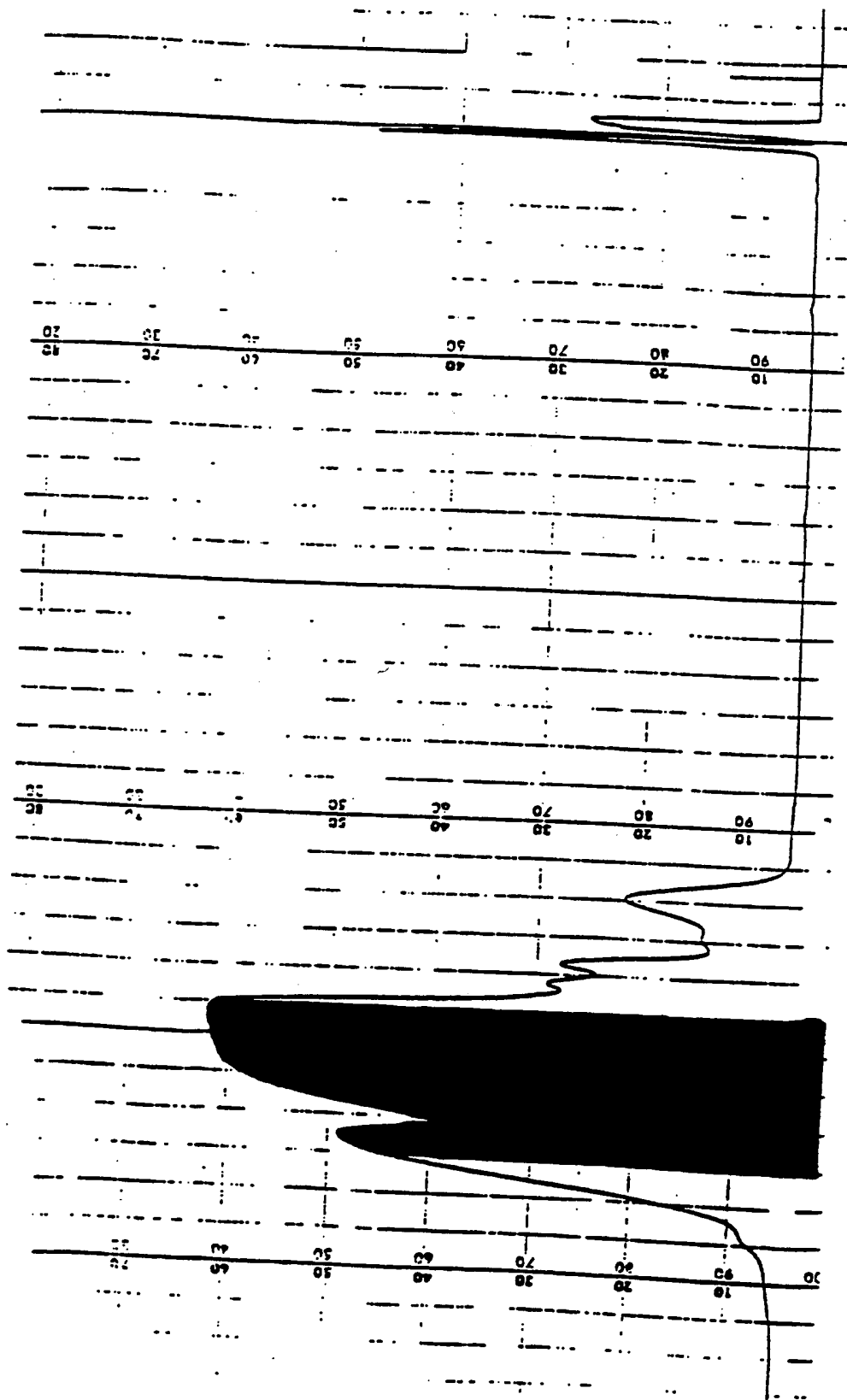
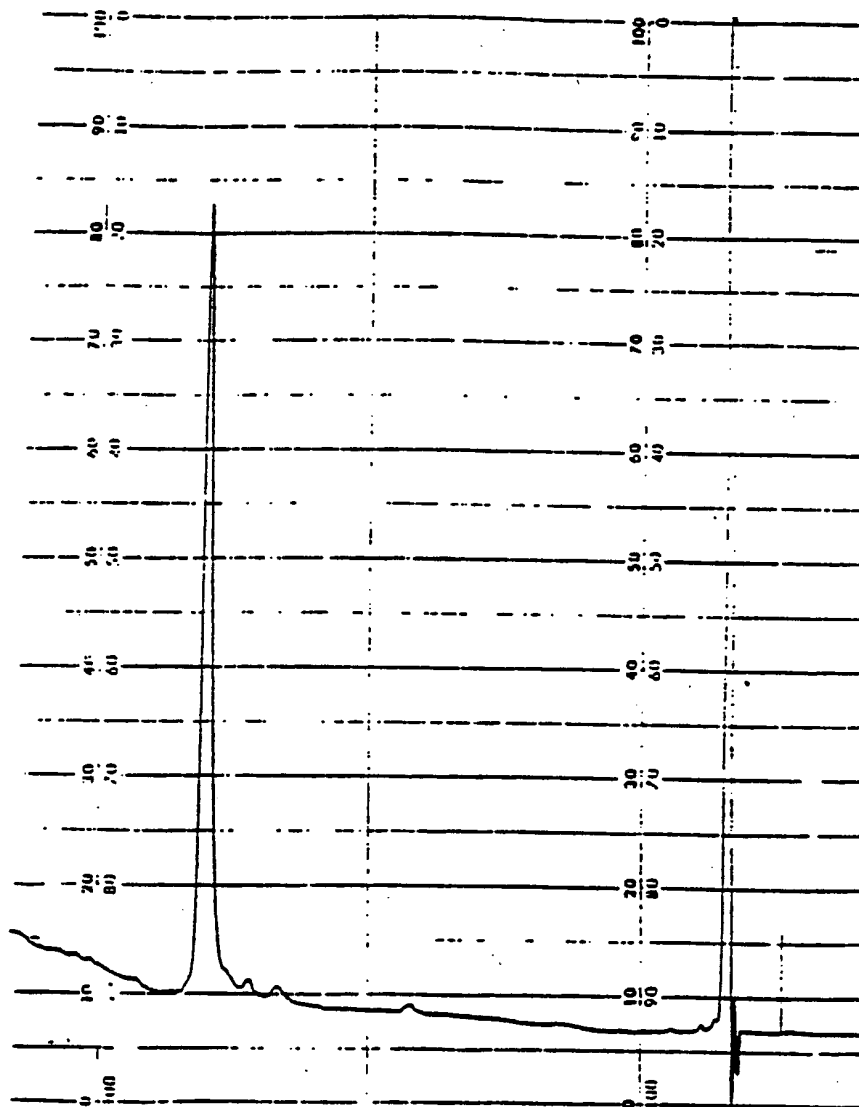


Figure 4

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**Purified Platelet Aggregation Inhibitor From
Eristicophis Macmahoni Venom****Figure 5**

Amino Acid Sequence and Fragmentation of Snake Venom PAIs

(K)= fragments from endoproteinase Lys-C digestion

(D)= fragments from endoproteinase Asp-N digestion

[Z]= fragment from pyroglutamyl aminopeptidase digestion

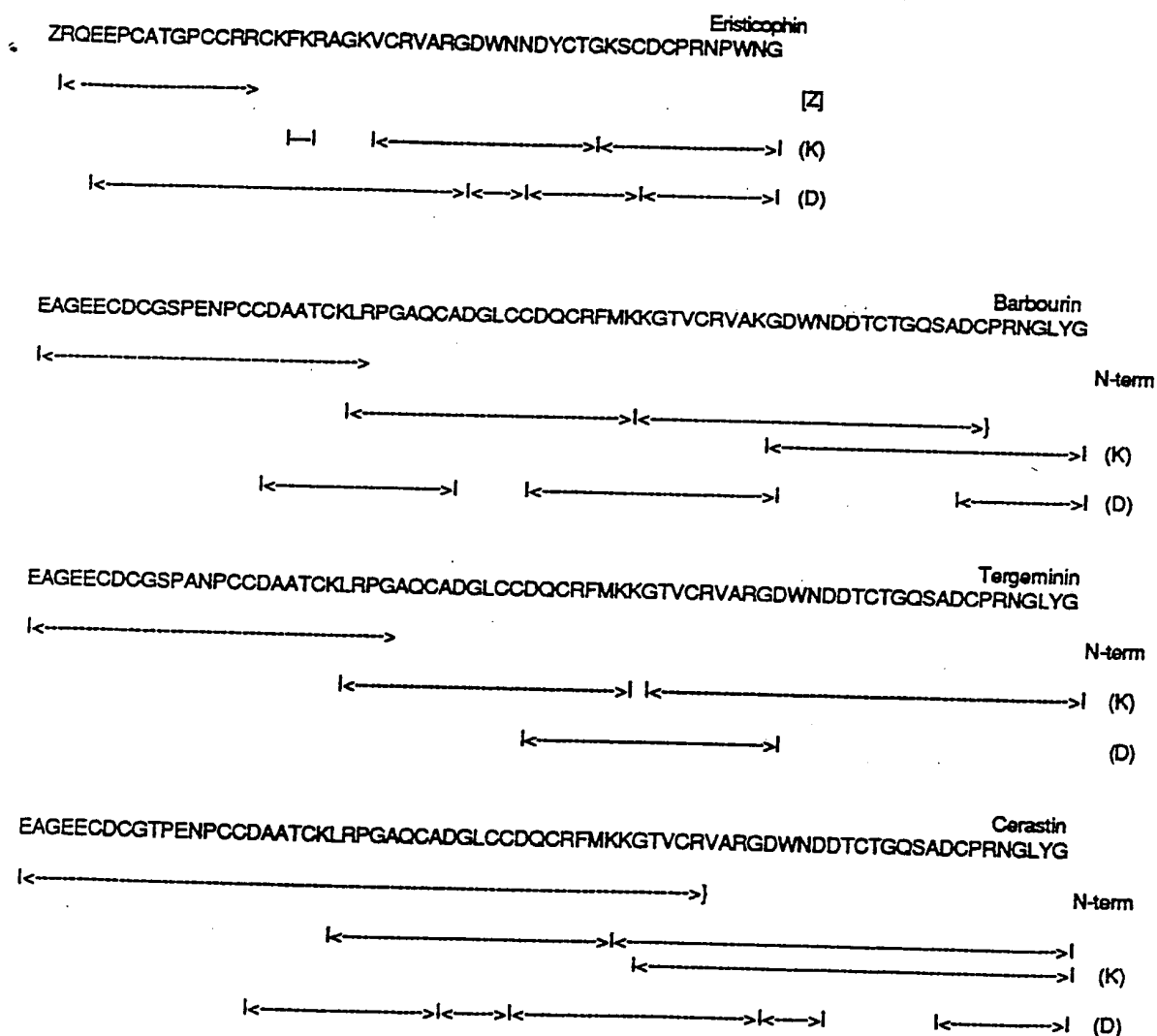


Figure 6-1

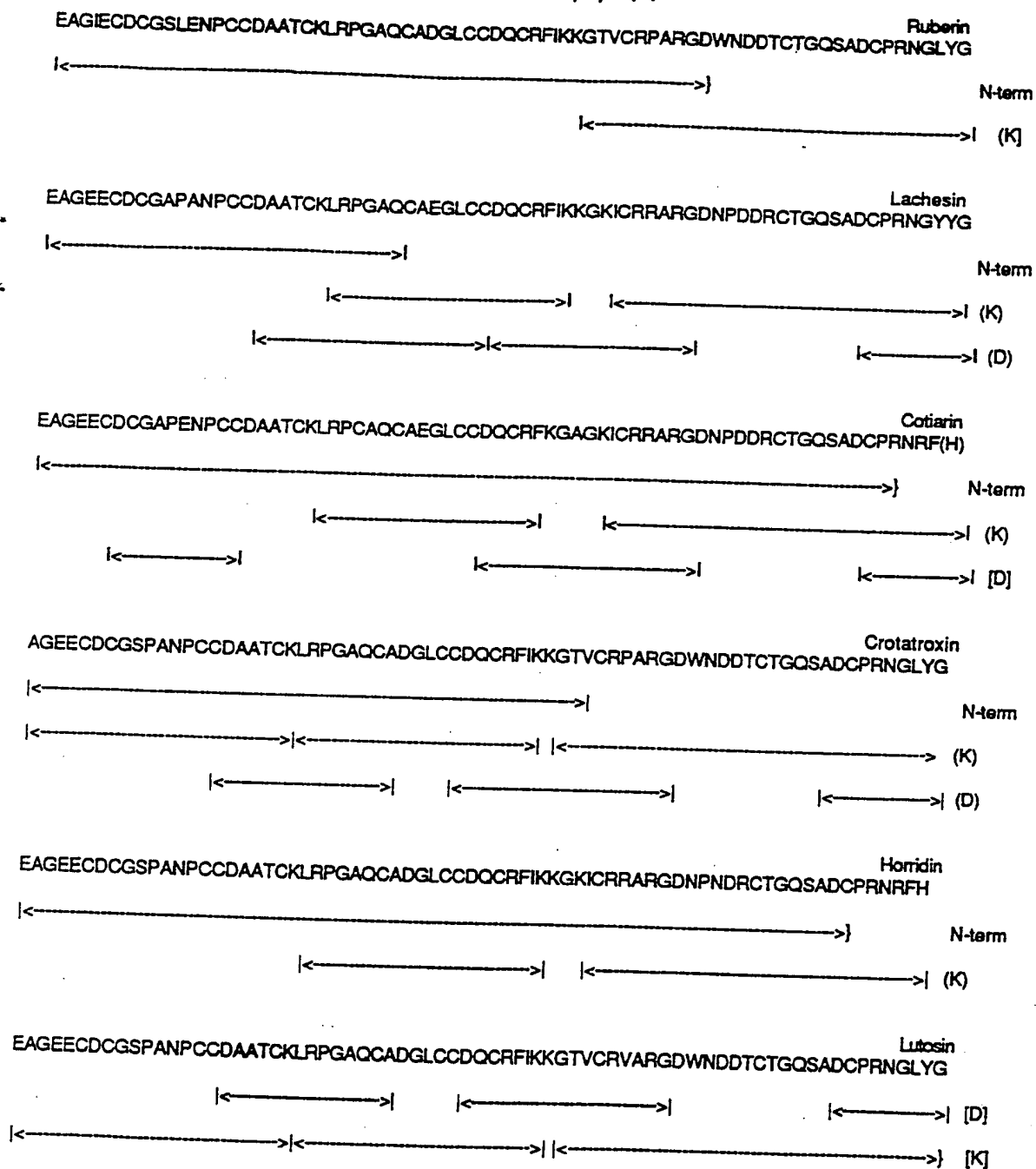


Figure 6-2

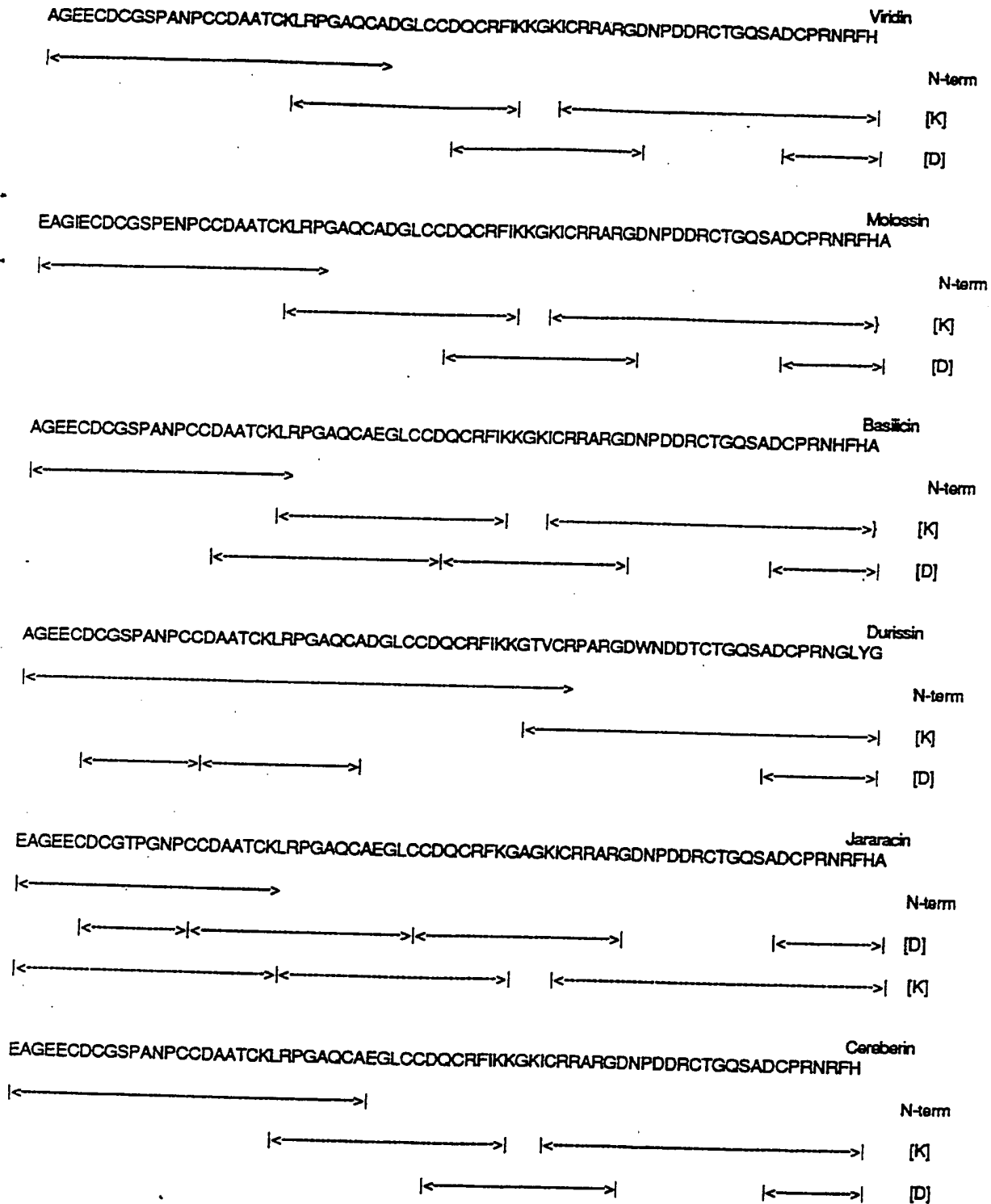


Figure 6-3

- 9 / 47

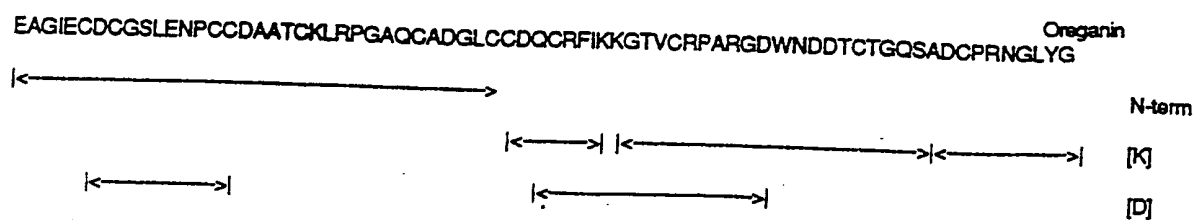
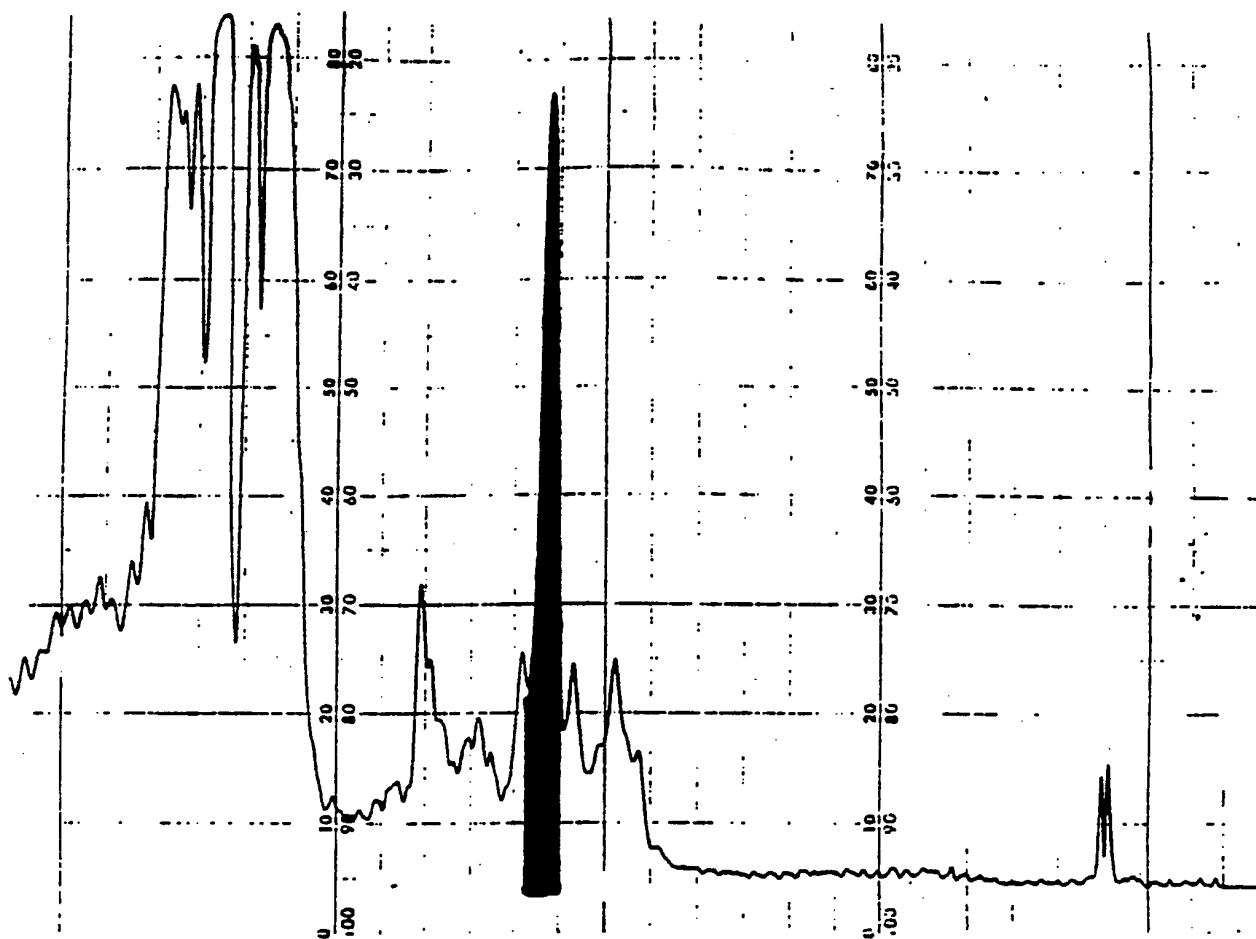


Figure 6-4

**HPLC Purification of Platelet Aggregation Inhibitor From
*Sistrurus catenatus tergeminus*****Figure 7**

**Purified Platelet Aggregation Inhibitor From
*Sistrurus catenatus tergeminus***

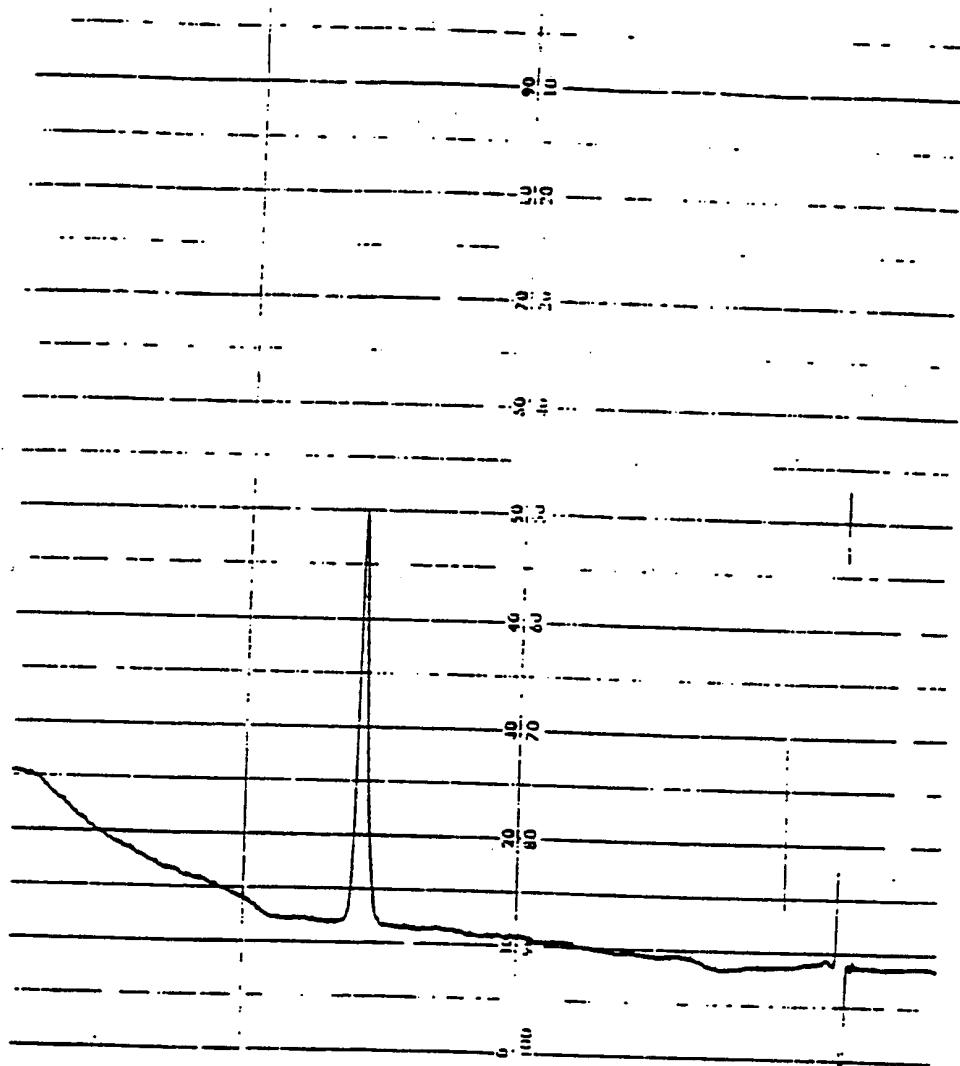


Figure 8

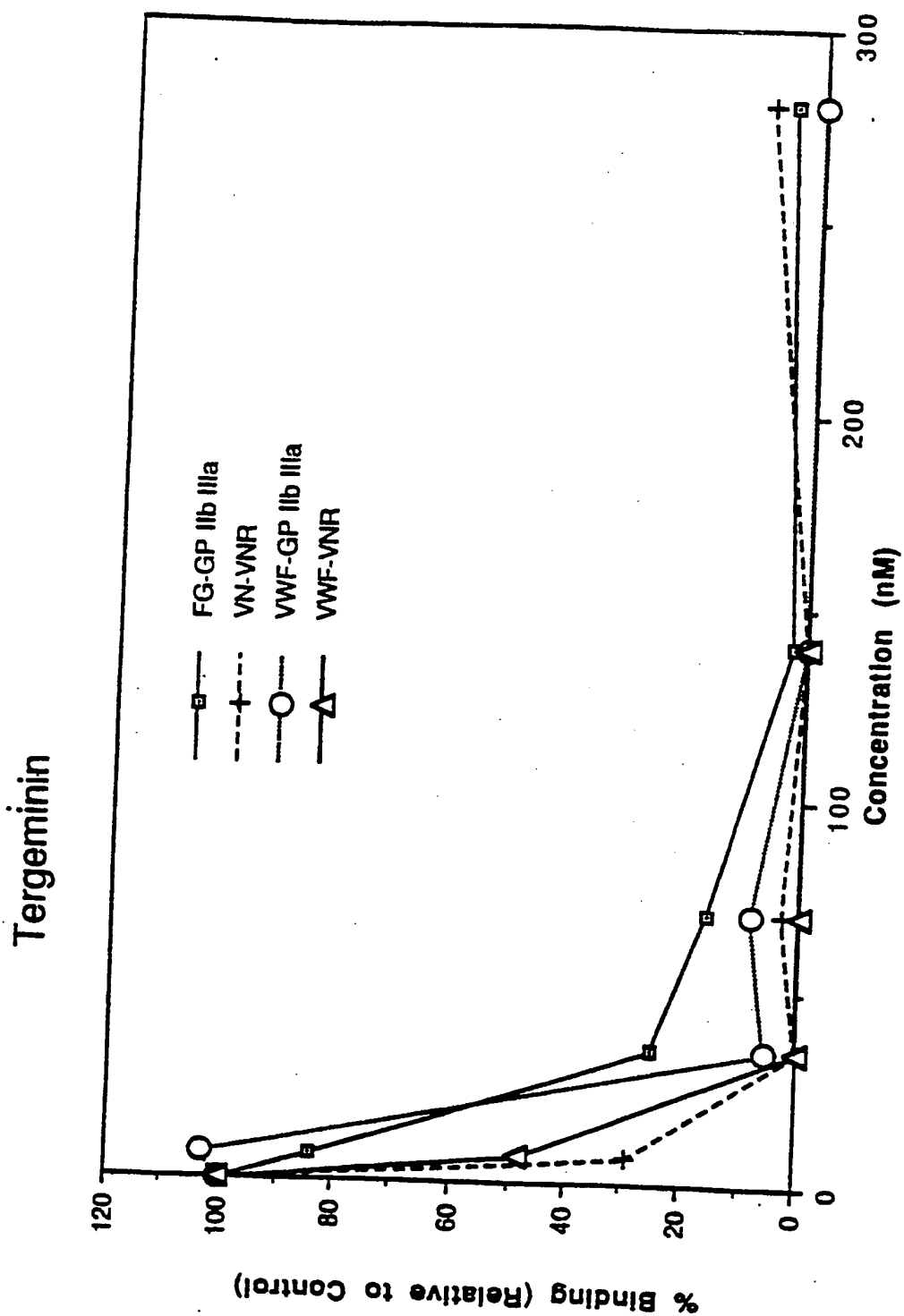


Figure 9

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HPLC Purification of Platelet Aggregation Inhibitor From *Sistrurus milaris barbouri*

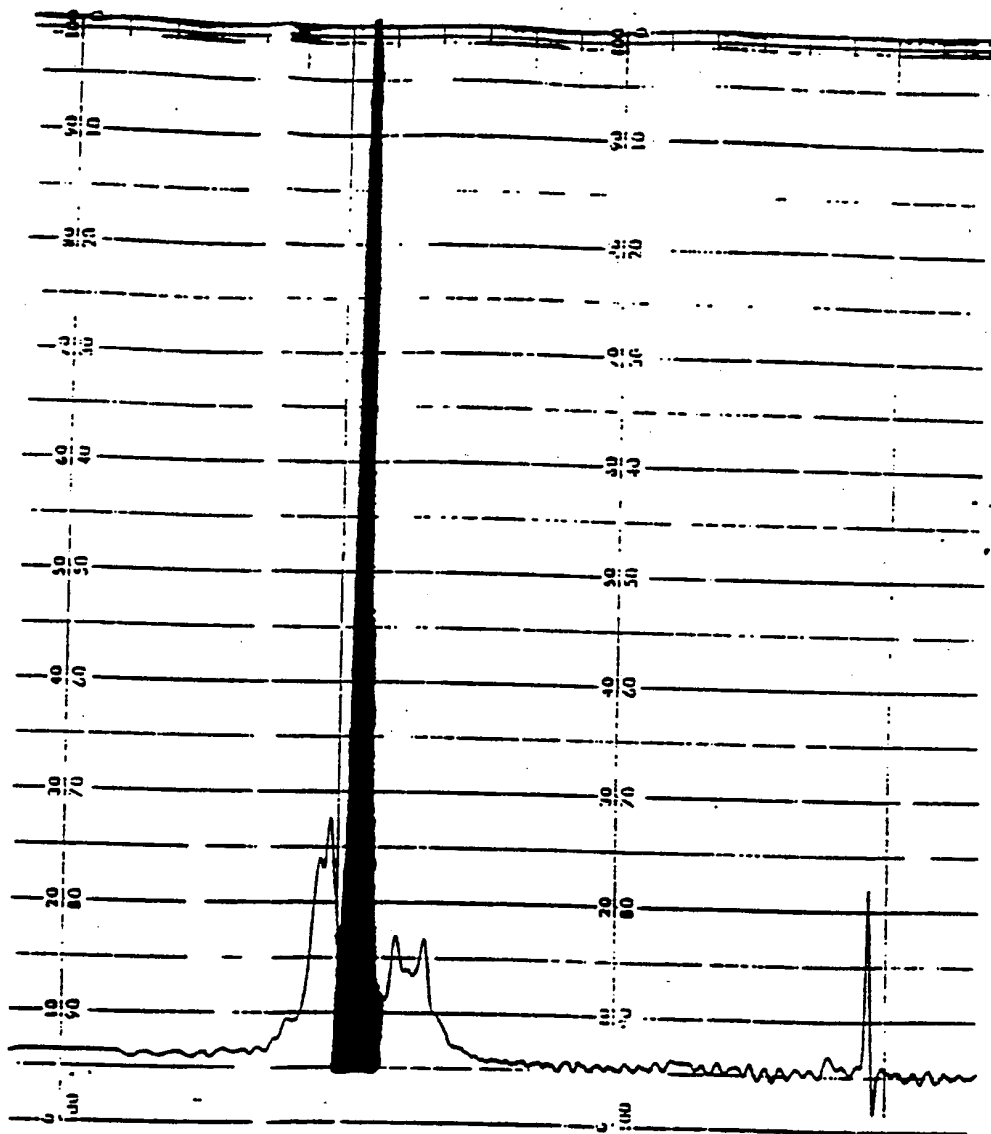


Figure 10

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**Purified Platelet Aggregation Inhibitor From
*Sistrurus mliarus barbouri***

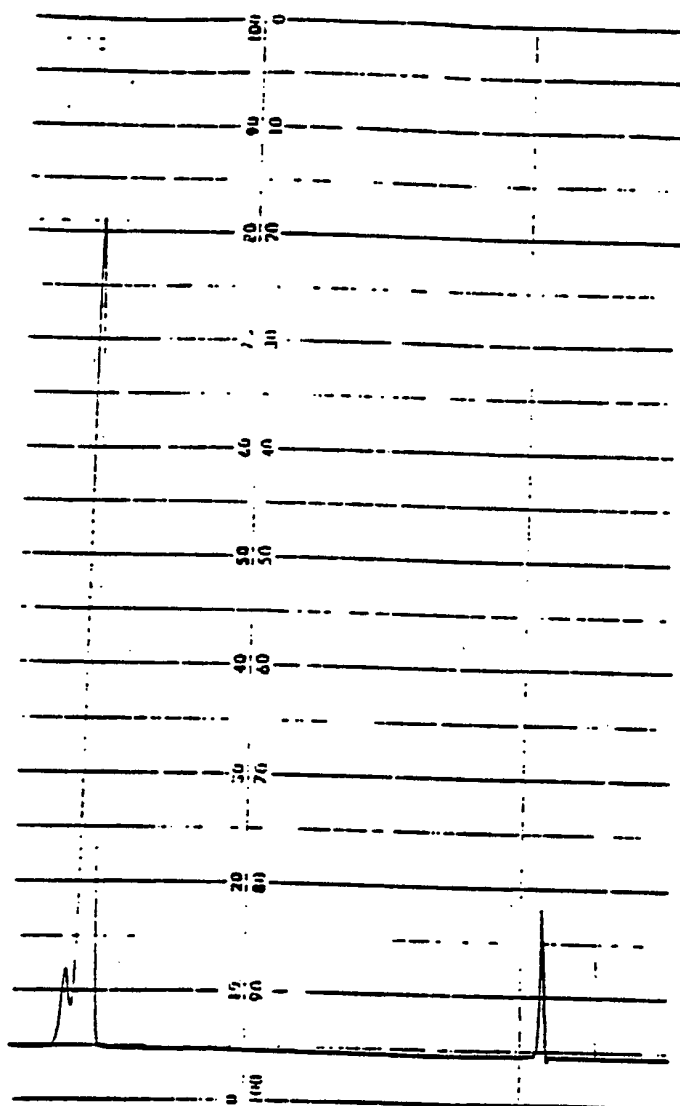


Figure 11

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Amino Acid Sequences of Snake Venom PAIs

	10	20	30	40	50	60	70	
	EAGEECDCGSPENPCCDAATCKLRPGAQCADGLCCDCRFMKGTVCRAKGDWNDTCTGQSADCPRNGLYG							Barbourin
	A				R			Tergeminin
	T				R			Cerastin
	A			I	R			Lutosin
-	A			I	P R			Crotatroxin
-	A			I	P R			Durissin
I	L			I	P R			Ruberin/Oreganin
I	L			I	KI R R NP R	RFHA		Mobassin
	A			I	KI R R NPN R	RFH-		Horridin
-	A			I	KI R R NP R	RFH-		Vindin
	A		E	I	KI R R NP R	RFH-		Cereberin
-	A		E	I	KI R R NP R	HFHA		Basilicin
	A A		E	I	KI R R NP R	Y		Lachesin
	T G		E	KGA KI	R R NP R	RFHA		Jararacin
	A		E	KGA KI	R R NP R	RF(H)-		Cotiarin
	A			K R I	R R NP R	-		Elegantin
D	A	I	GE	S	I R R DL Y N I G	P HA		Albolabrin
--	S			XX R I	I R FP R L N G	XNDL-		Flavovindin
D	A	I	GE	S IEE	I R DL Y N R G	PFH-		Trigramin
-- K	S		GE	K SRA KI	IPR MP R	YH---		Kistrin
			E	K E	- R R DVN N I G	PFH-		Applaggin

Figure 12-1

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10 20 30 40 50
 ZRQEPCATGPCCRCKFKRAGKVCRVARGDWNNDYCTGKSCDCPRNPWNG Eristicophin
 ECESGPCCRNCKFLKEGTICKRARGDDMDDYCNGKTCDCPRNPHKGPAT Echistatin
 10 20 30 40 50 60 70 80
 SPPVCGNEILEQGEDCDGSPANCDQCCNAATCKLTPGSQCNHGECCDQCKFKKARTVCRIARGDWNDDYCTGKSSDCPWNH Bitistatin
 K R Y R G Bitan-a

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HPLC Fractionation of Lachesis mutas Platelet Aggregation Inhibitor

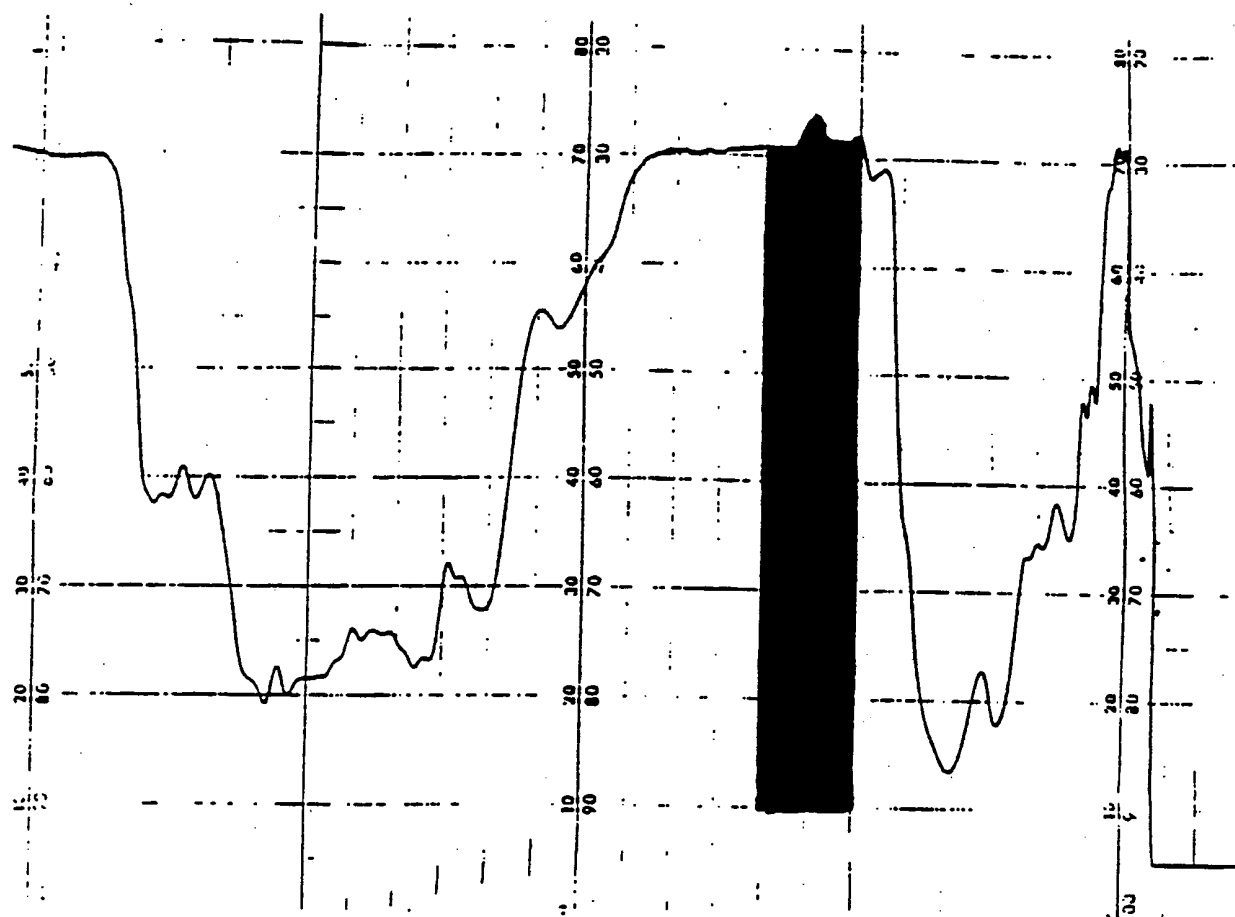


FIGURE 13

**Slow Gradient HPLC Purification of Platelet
Aggregation Inhibitor From Lachesis mutas**

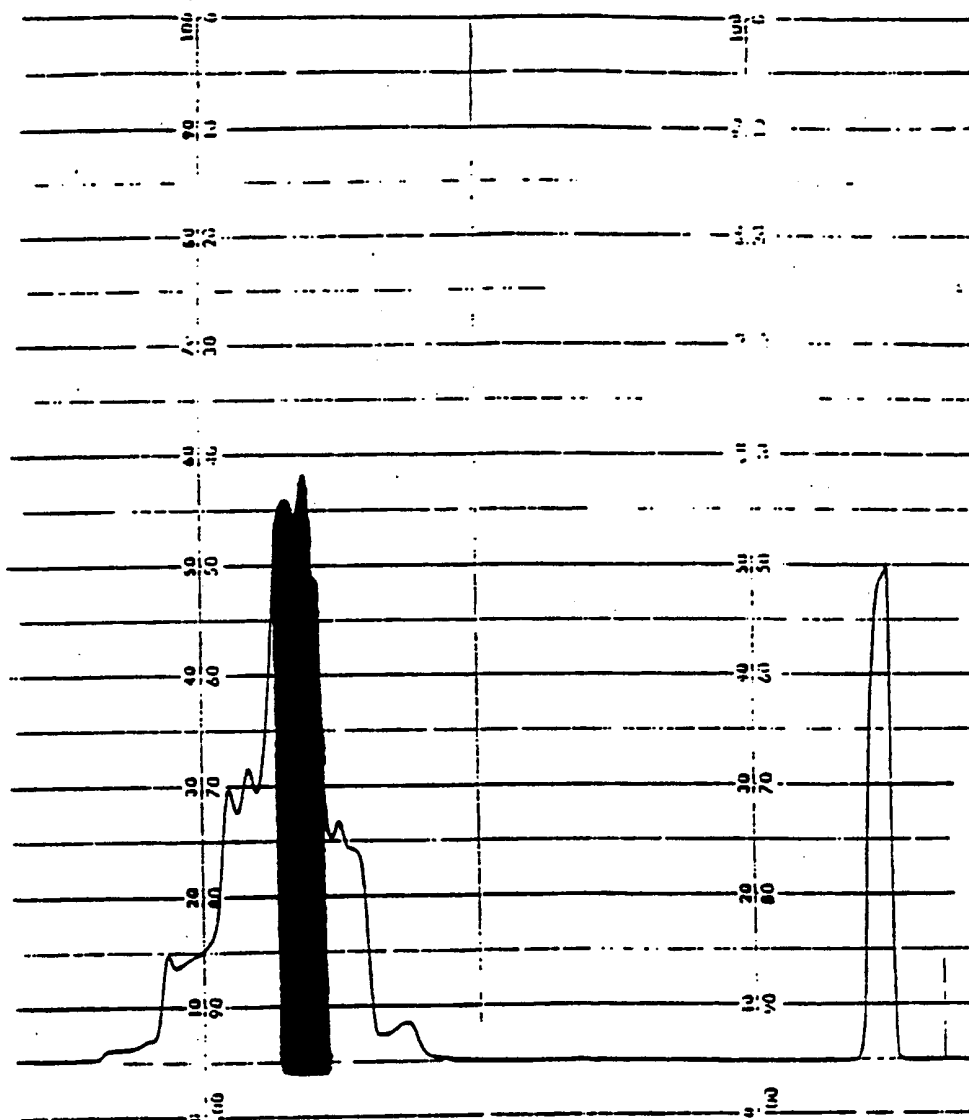


FIGURE 14

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**Purified Platelet Aggregation Inhibitor From
*Lachesis mutas***

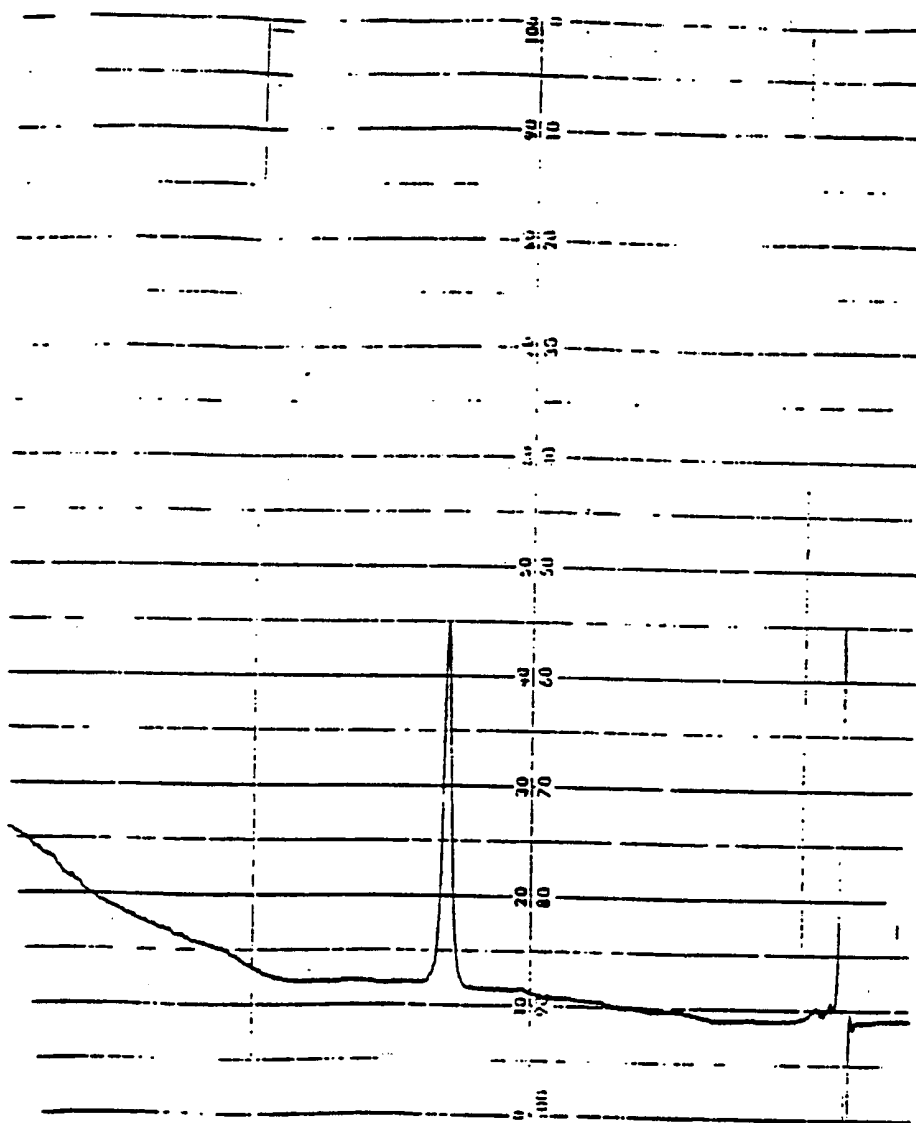


FIGURE 15

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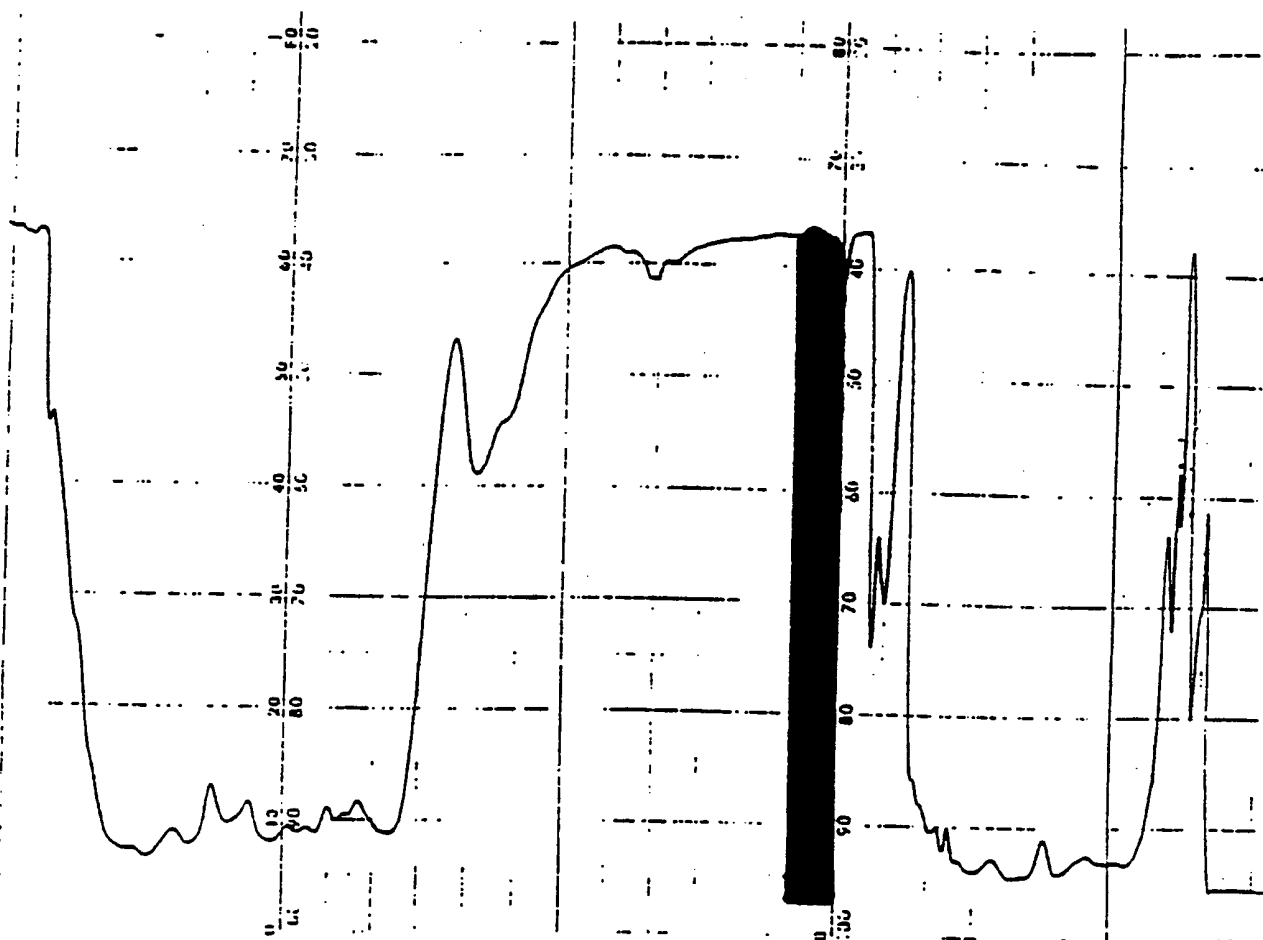
**HPLC Fractionation of Platelet Aggregation Inhibitor From
*Crotalus viridis viridis***

FIGURE 16

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Purified Platelet Aggregation Inhibitor From
Crotalus viridis viridis

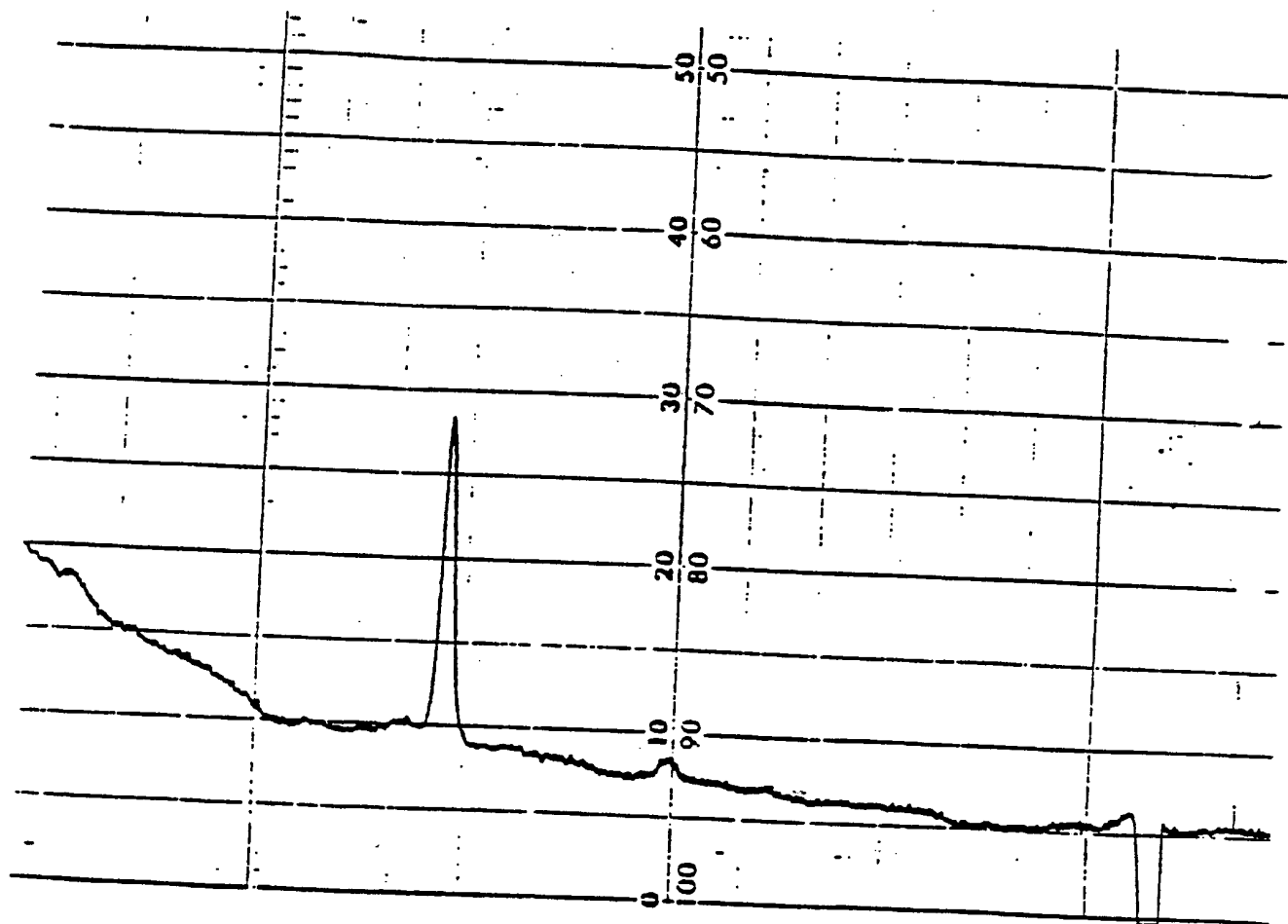
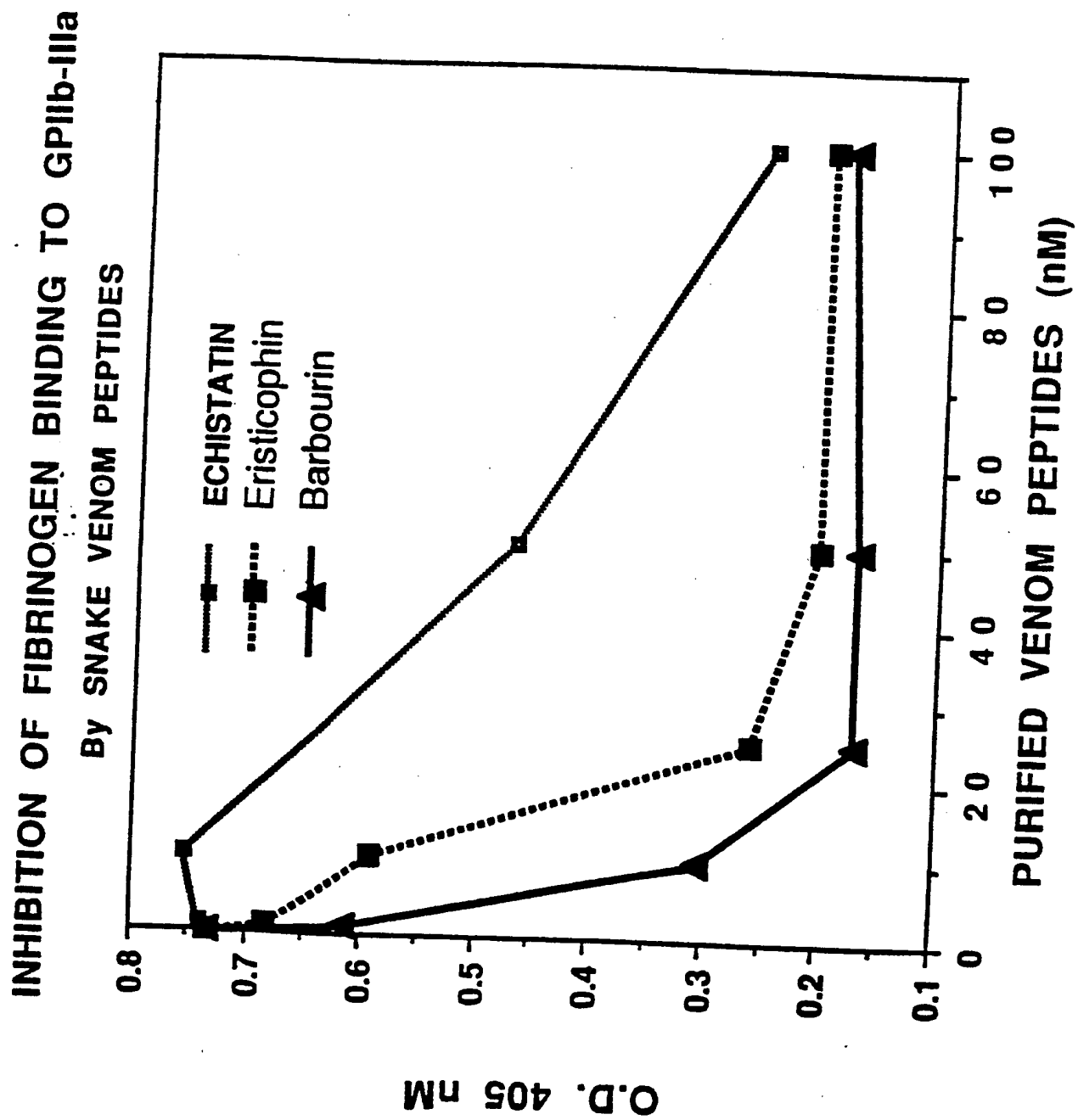


FIGURE 17

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FIGURE 18



Inhibition of ADP Induced Platelet Aggregation By Purified Snake Venom Peptides

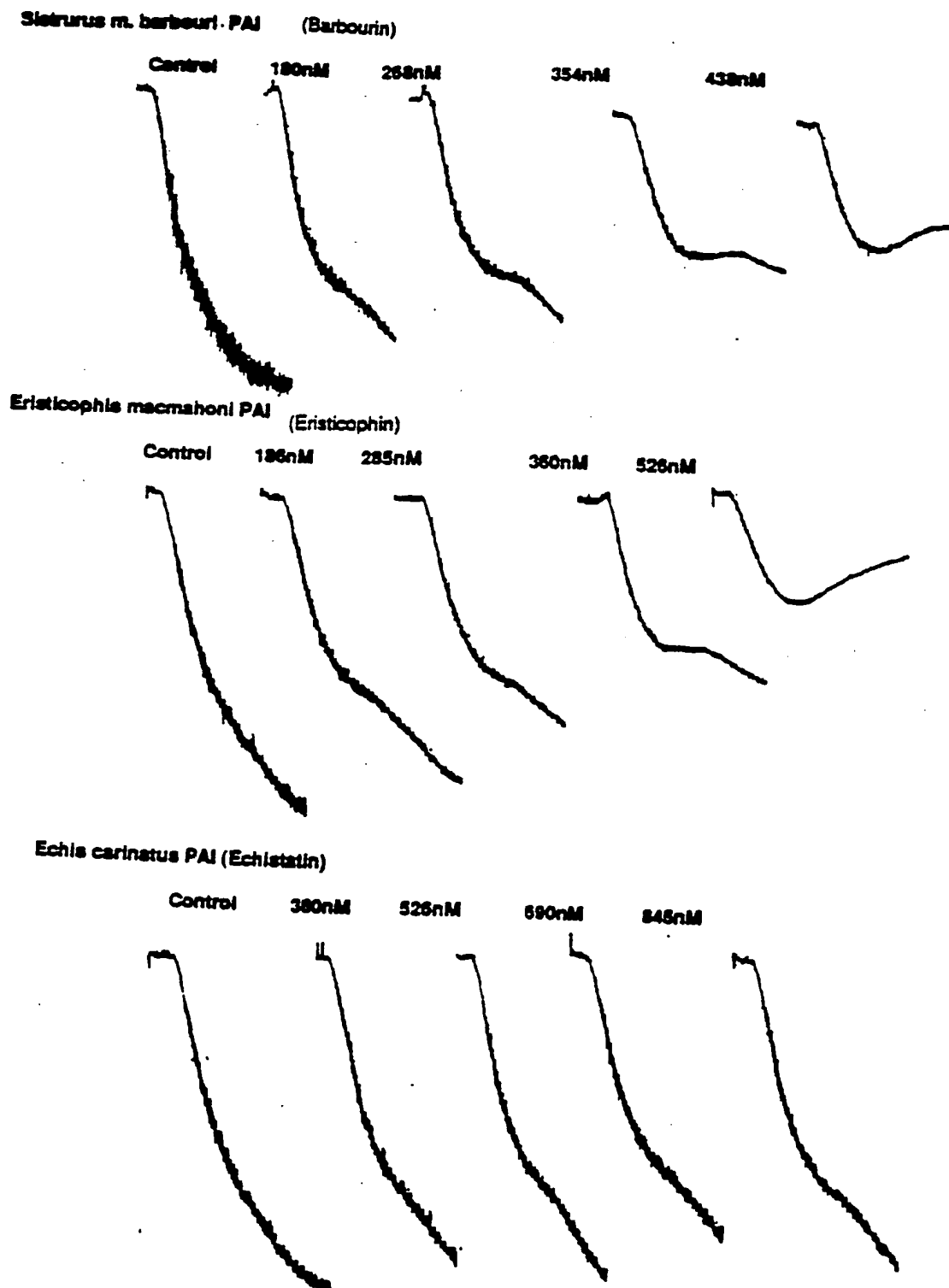


FIGURE 19

HPLC Purification of *Crotalus cerastes cerastes* Platelet Aggregation Inhibitor

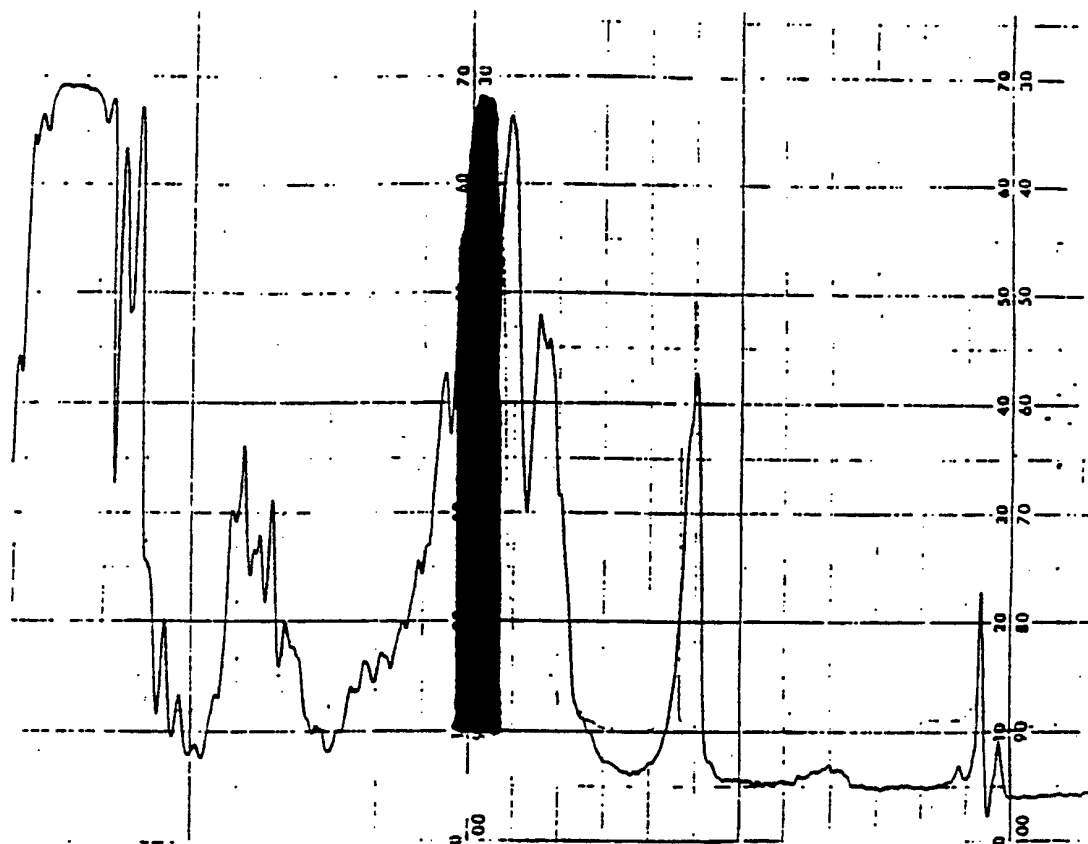


Figure 20

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**Purified Platelet Aggregation Inhibitor From
Crotalus cerastes cerastes**

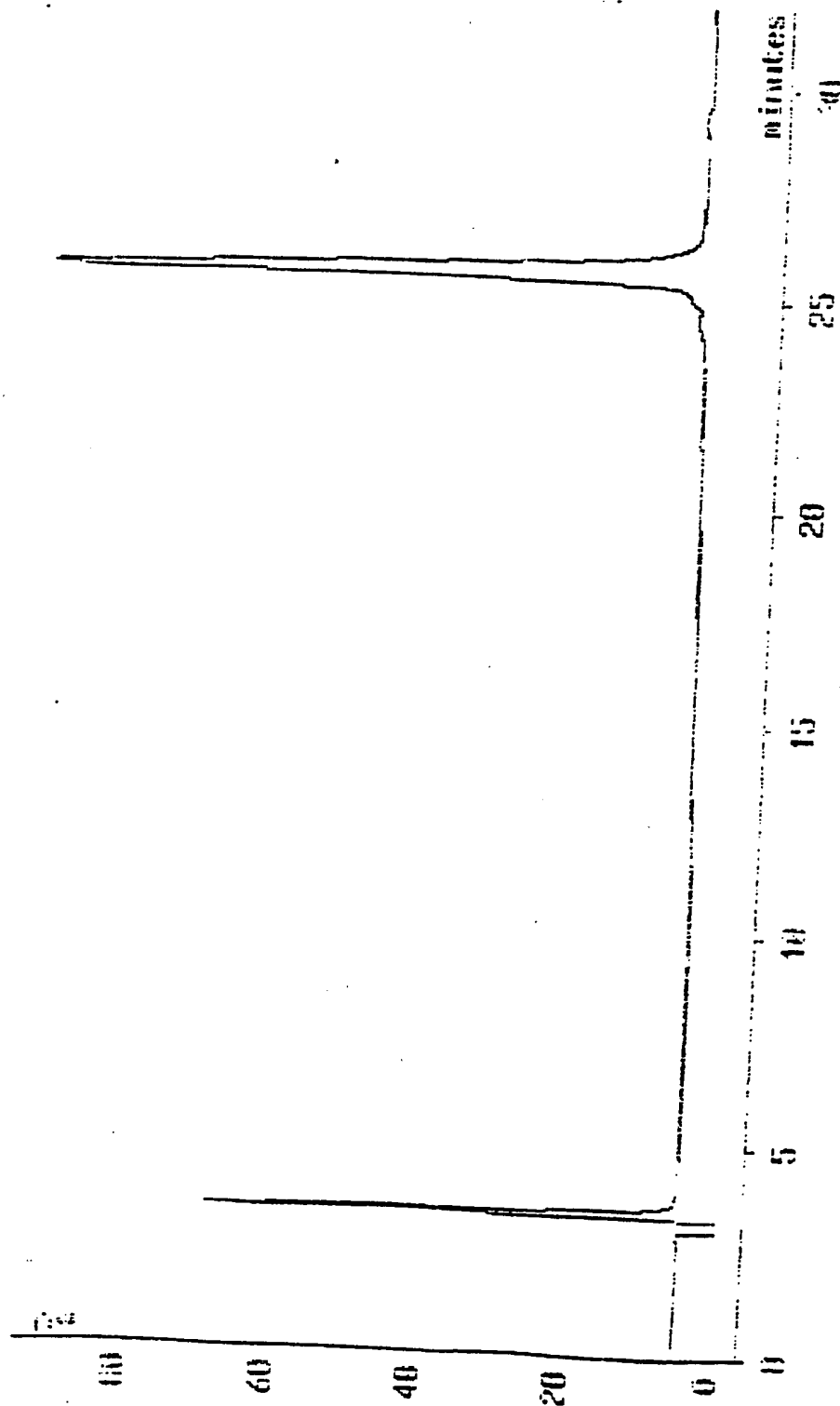


Figure 21

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HPLC Purification of *Crotalus ruber ruber* Platelet
Aggregation Inhibitor

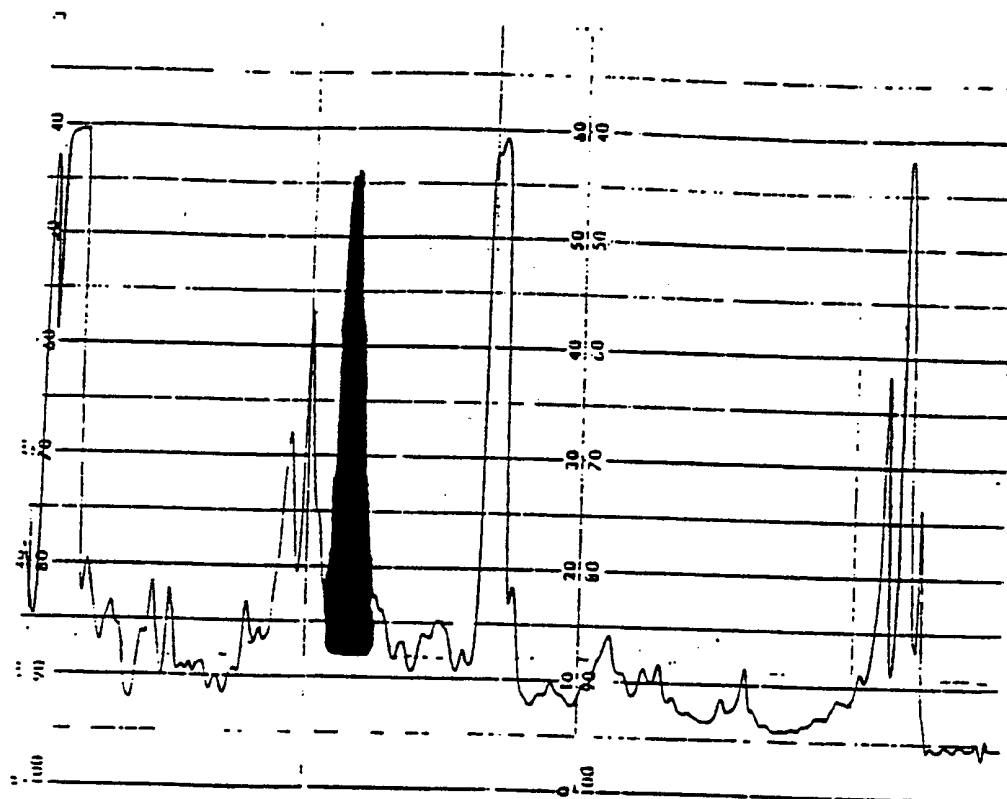


Figure 22

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Purified Platelet Aggregation Inhibitor From
Crotalus atrox

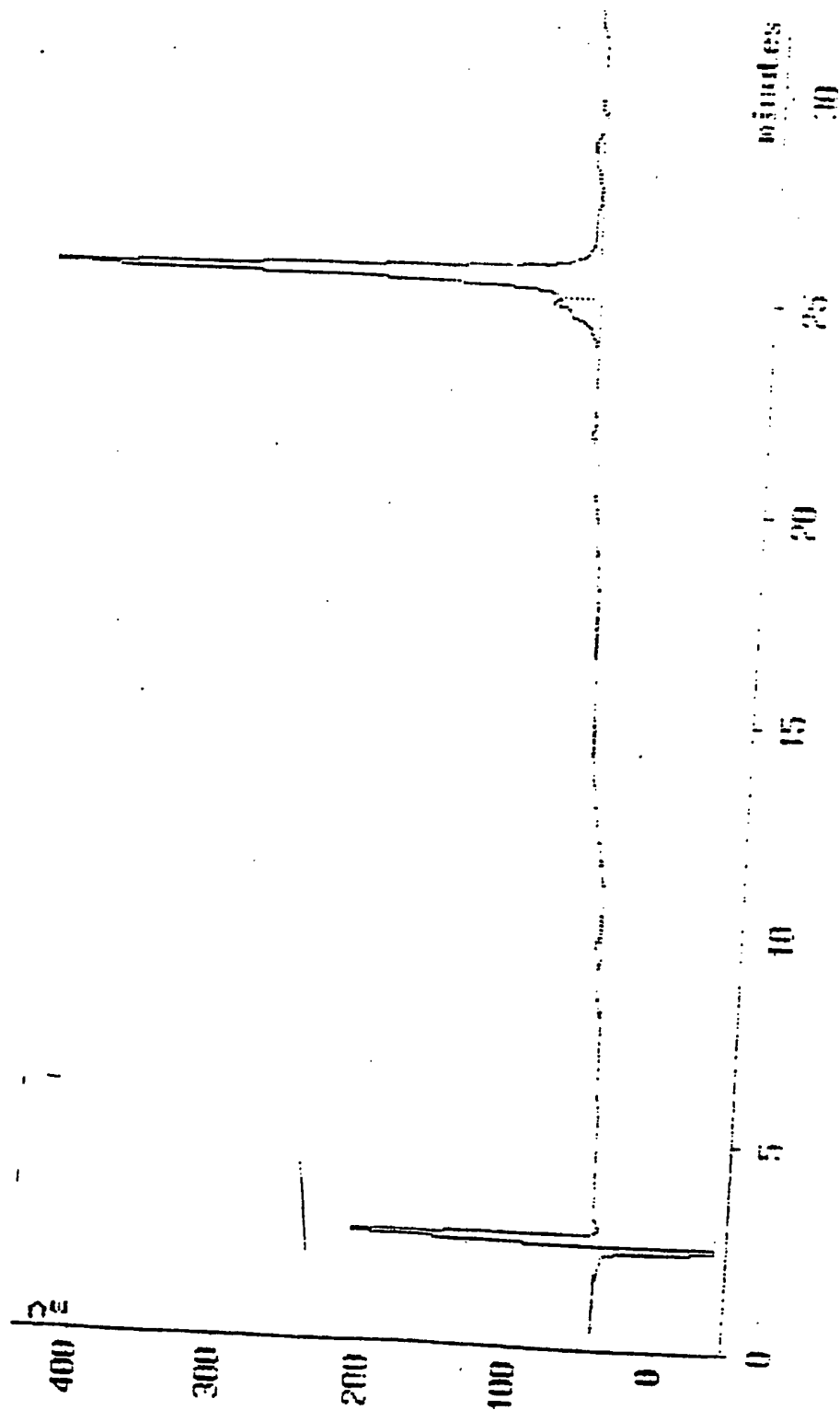


Figure 23

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Purified Platelet Aggregation Inhibitor From
Bothrops cotiara

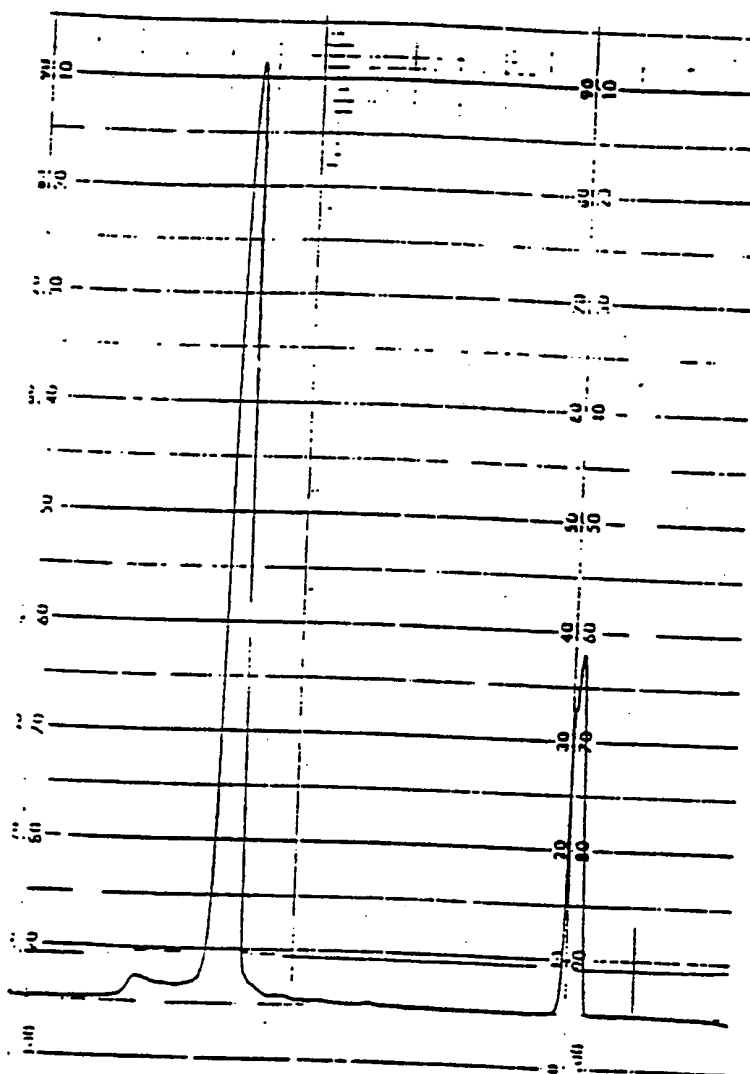


Figure 24

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Cotiarin

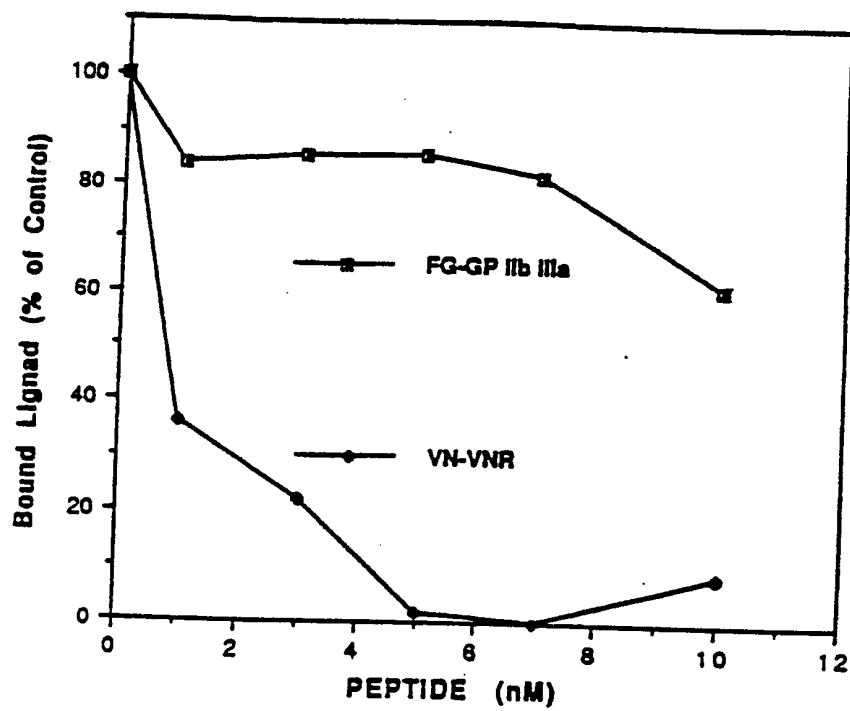


Figure 25

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Figure 26

EFFECTS OF PURIFIED SNAKE VENOM PEPTIDES ON LIGAND/RECEPTOR BINDING

	Fibrinogen/ GPIIb-IIIa 10nm*	Vitronectin/ VnR >100nm 16 >120
Barbourin		
Tergeminin	16	
Truncated Sist. m. barbouri. (Analog #1)	10	
Crotalus v. helleri	20	25
Cerastin	2	2
Ruberin	10	10
Crotalus atrox	5	5
Crotalus d. totonacus	1-2	
Crotalus v. lutosus	5	5
Bothrops jararacussu	25	5
Cotiarin	>10	1
Eristicophin	25	25
Synthetic "Eristicophin" w/ KGDW(Analog#2)	40	>150
Cyclic GCGKGDWPCA-NH2 (Analog #3)	900	>20,000

*Concentration of purified peptide which inhibits the binding of fibrinogen to GPIIb-IIIa or vitronectin to the vitronectin receptor by 50% (IC50). The symbol ">" indicates that the IC50 is greater than this value, which is the highest concentration examined to date. Note that substitution of the arginine in the "RGDW" sequence in Eristicophin with a lysine (KGDW) imparts specificity for inhibiting fibrinogen binding to GPIIb-IIIa versus vitronectin to the vitronectin receptor.

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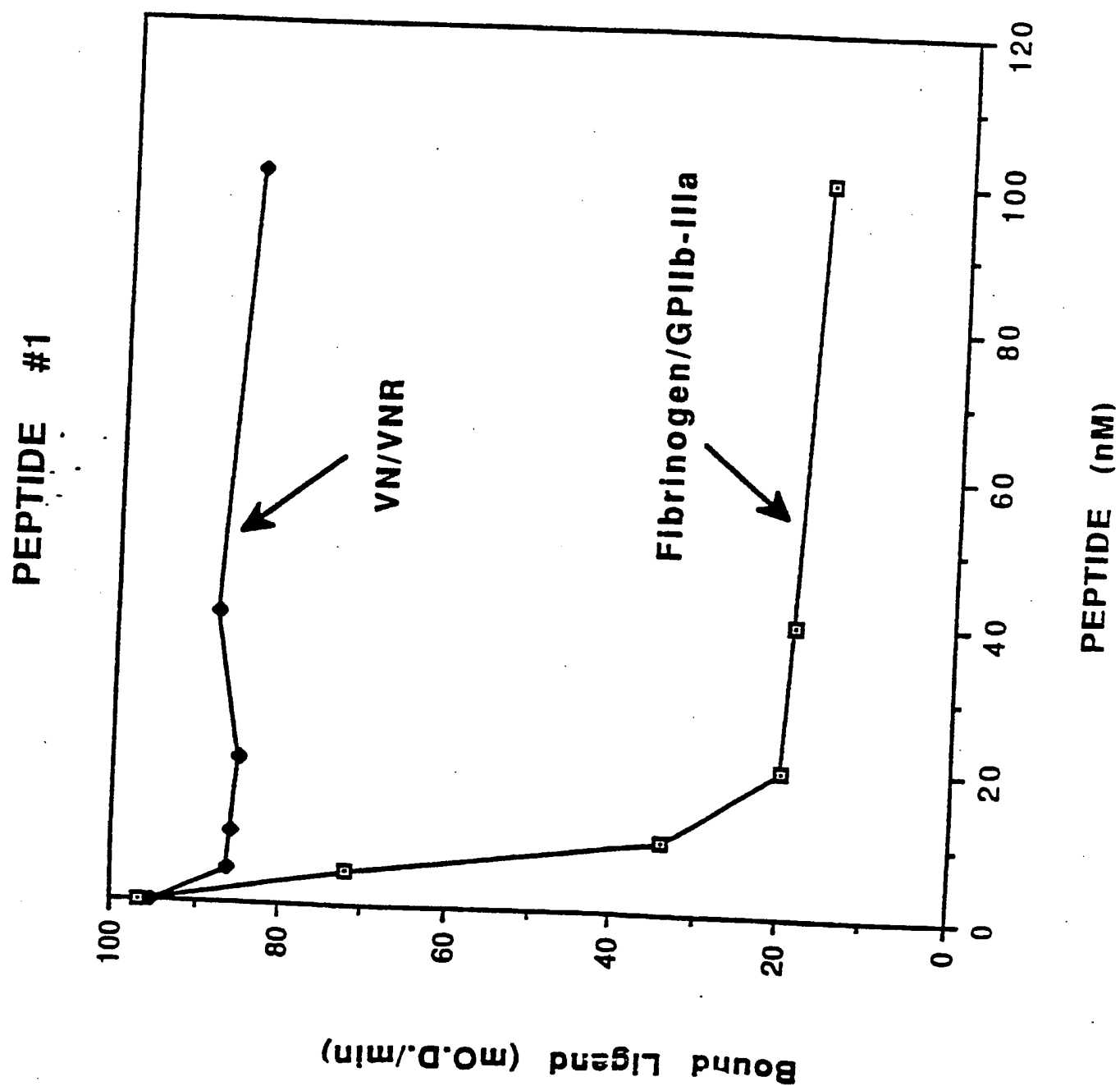


Figure 27

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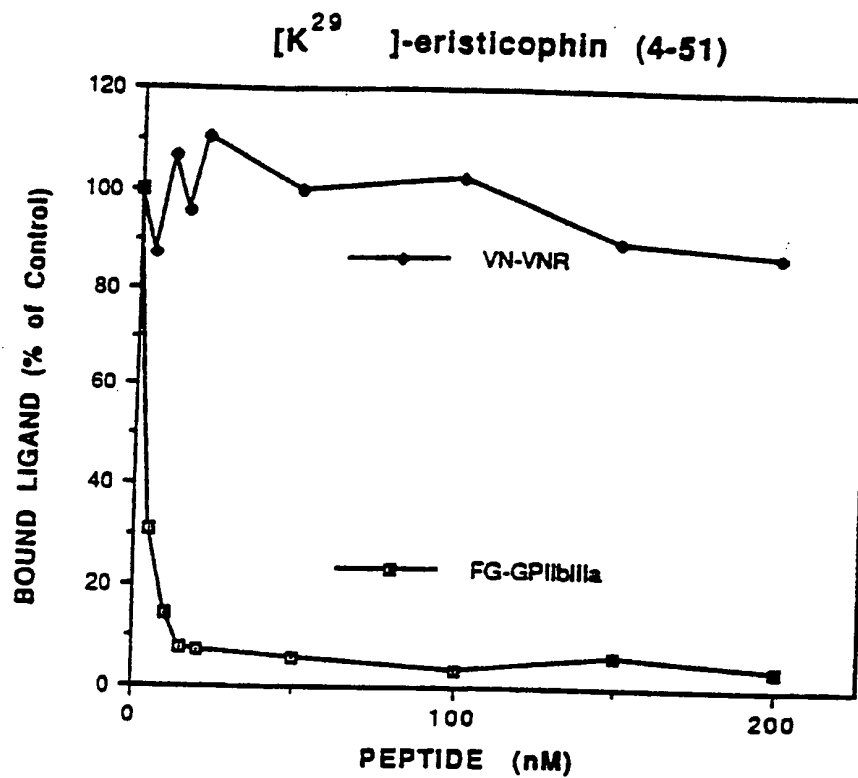


Figure 28

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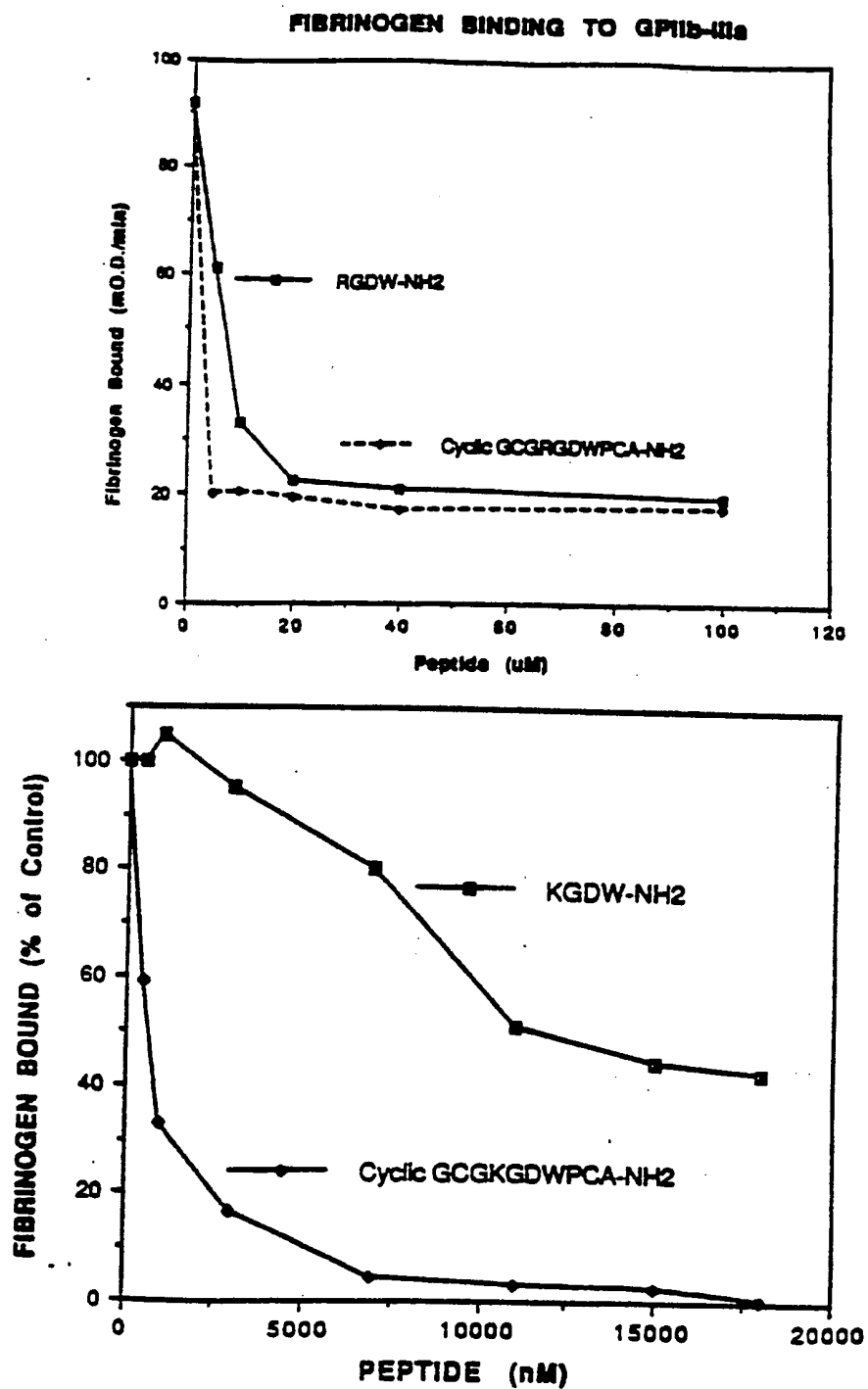
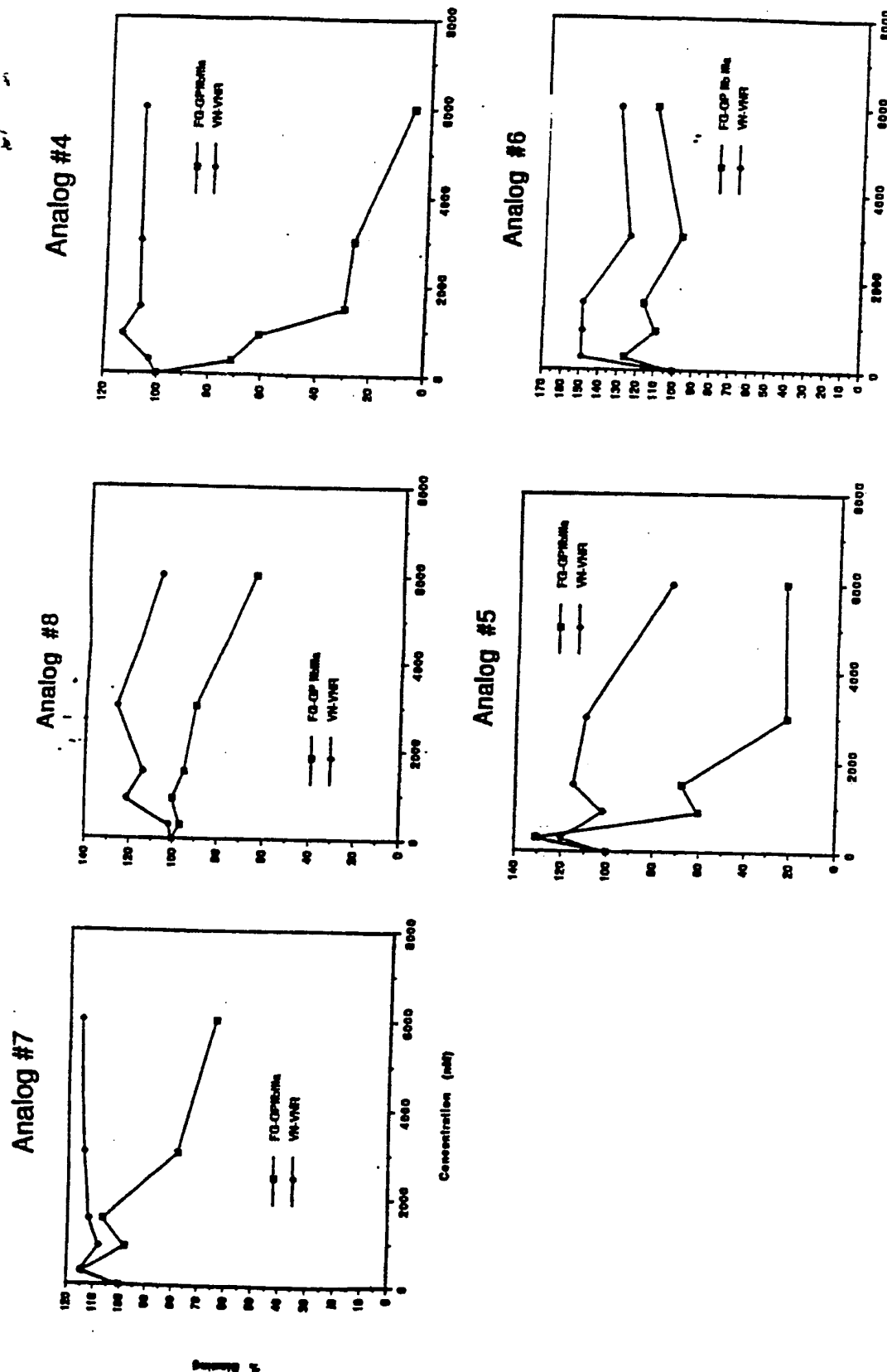


Figure 29

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Figure 30



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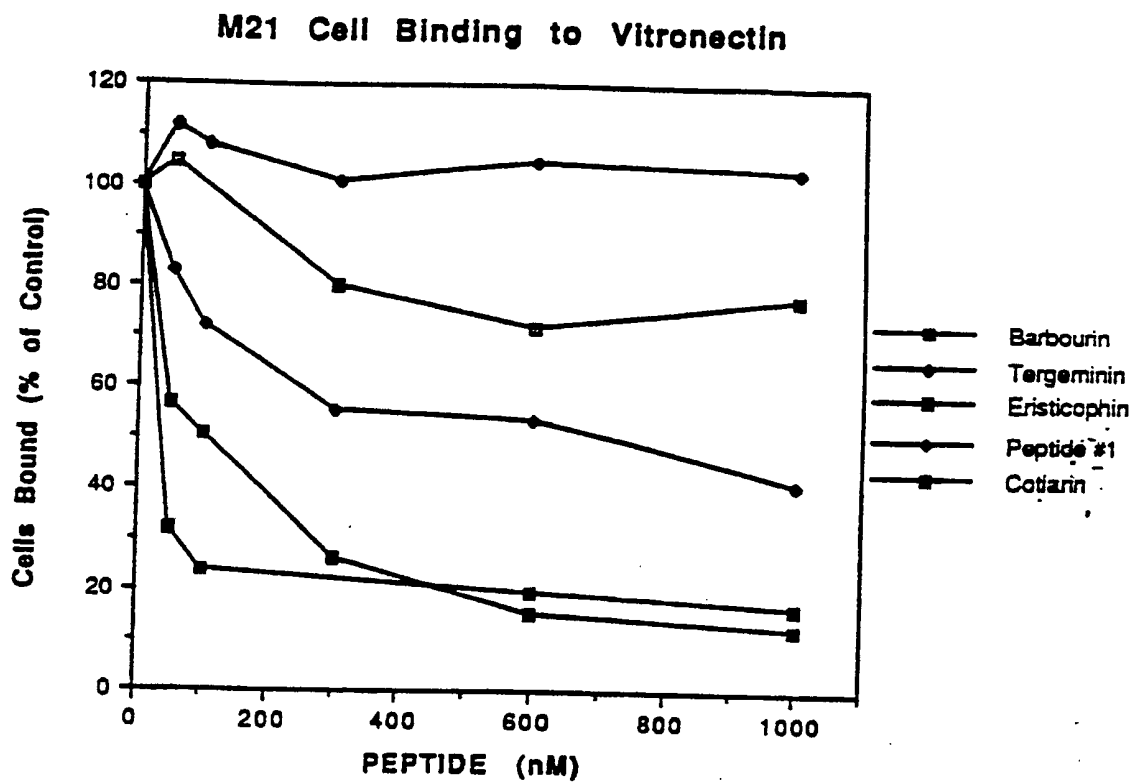


Figure 31

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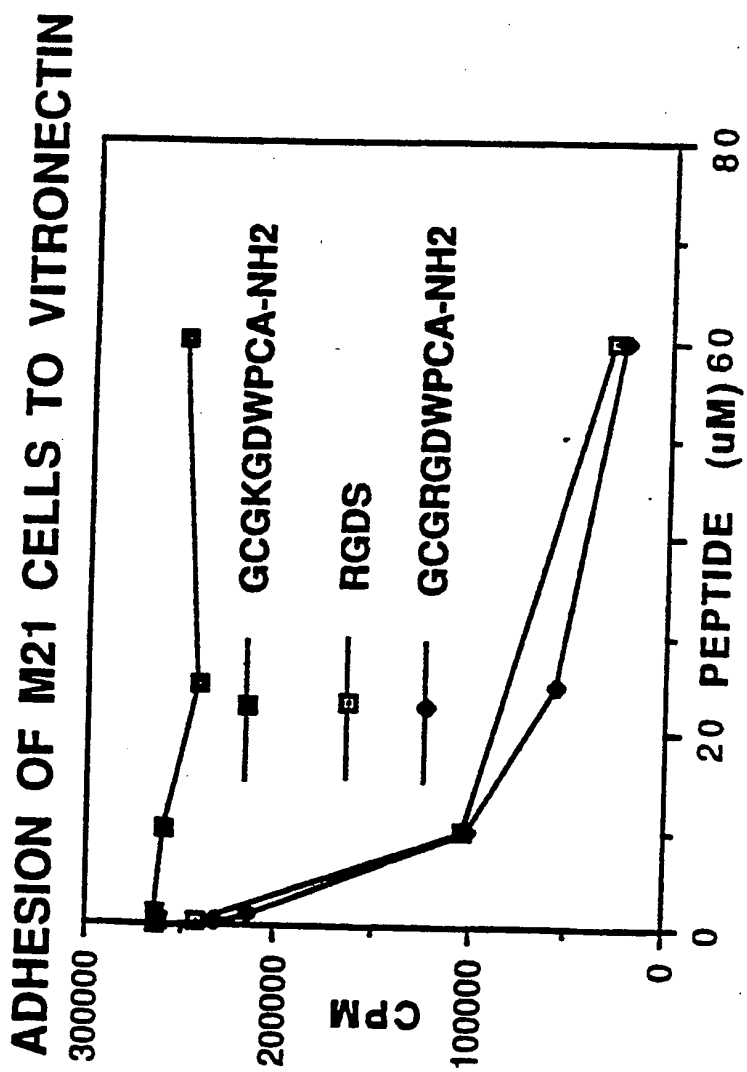


Figure 32

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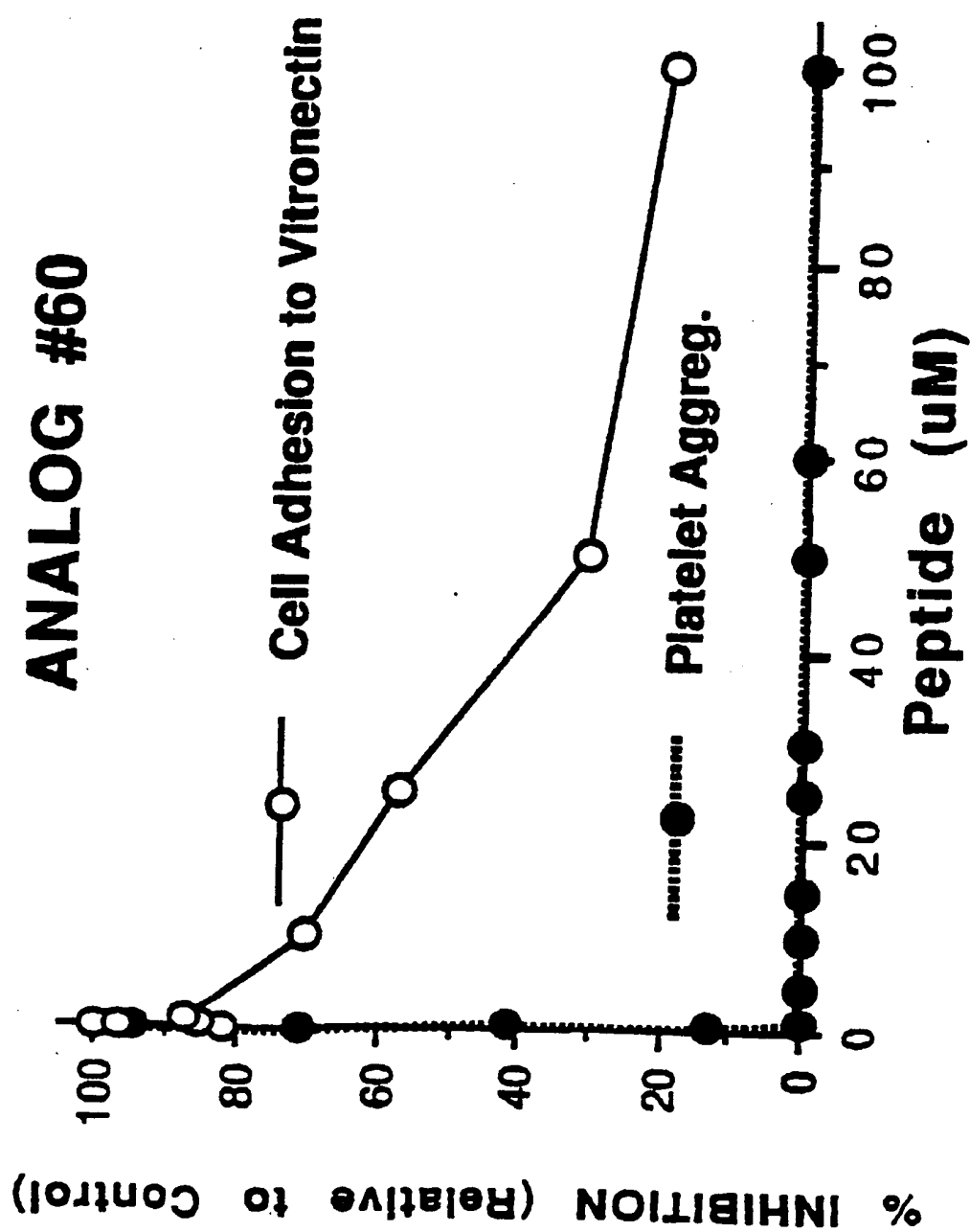
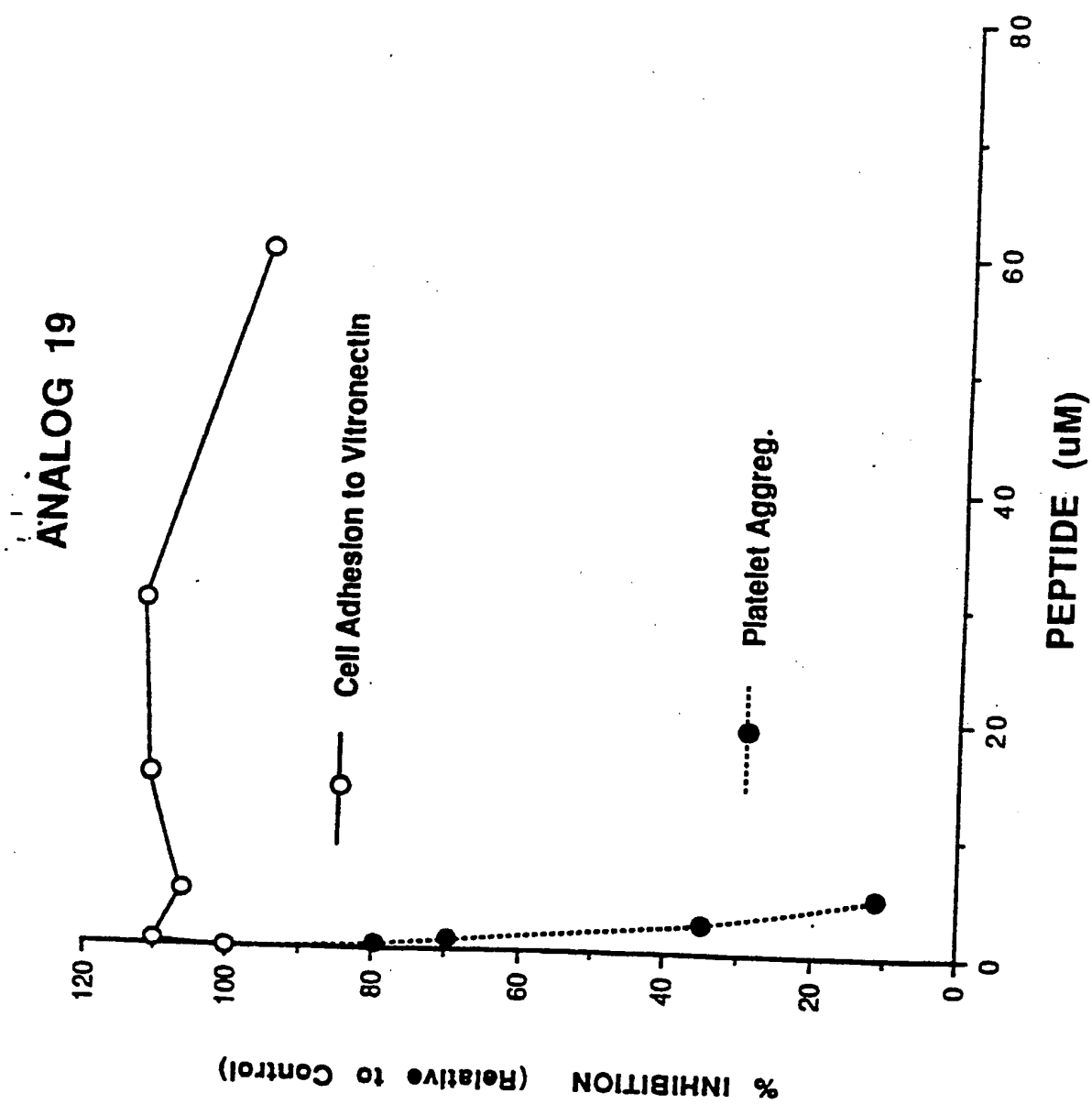


Figure 33

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FIGURE 34



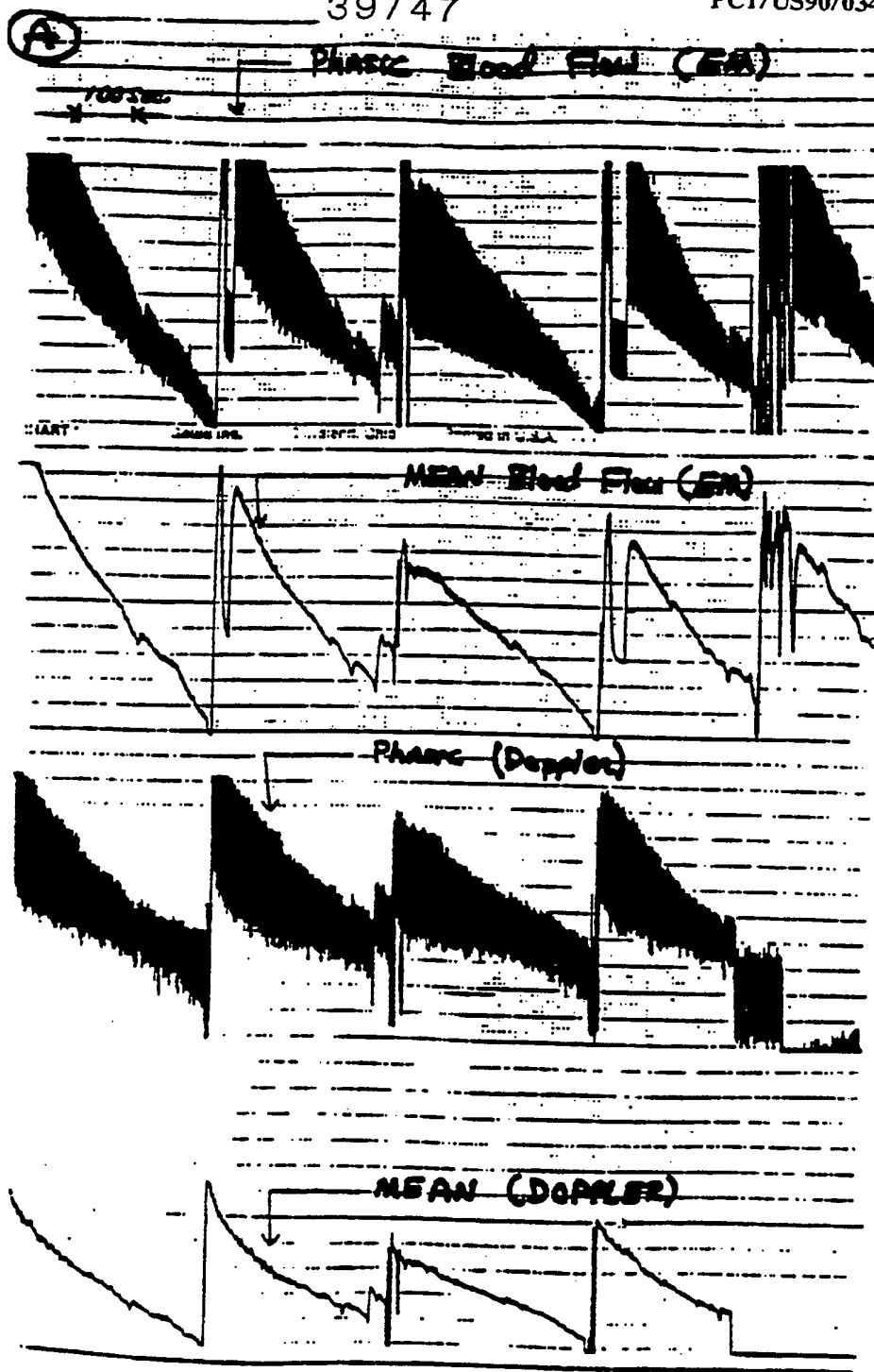


Figure 35

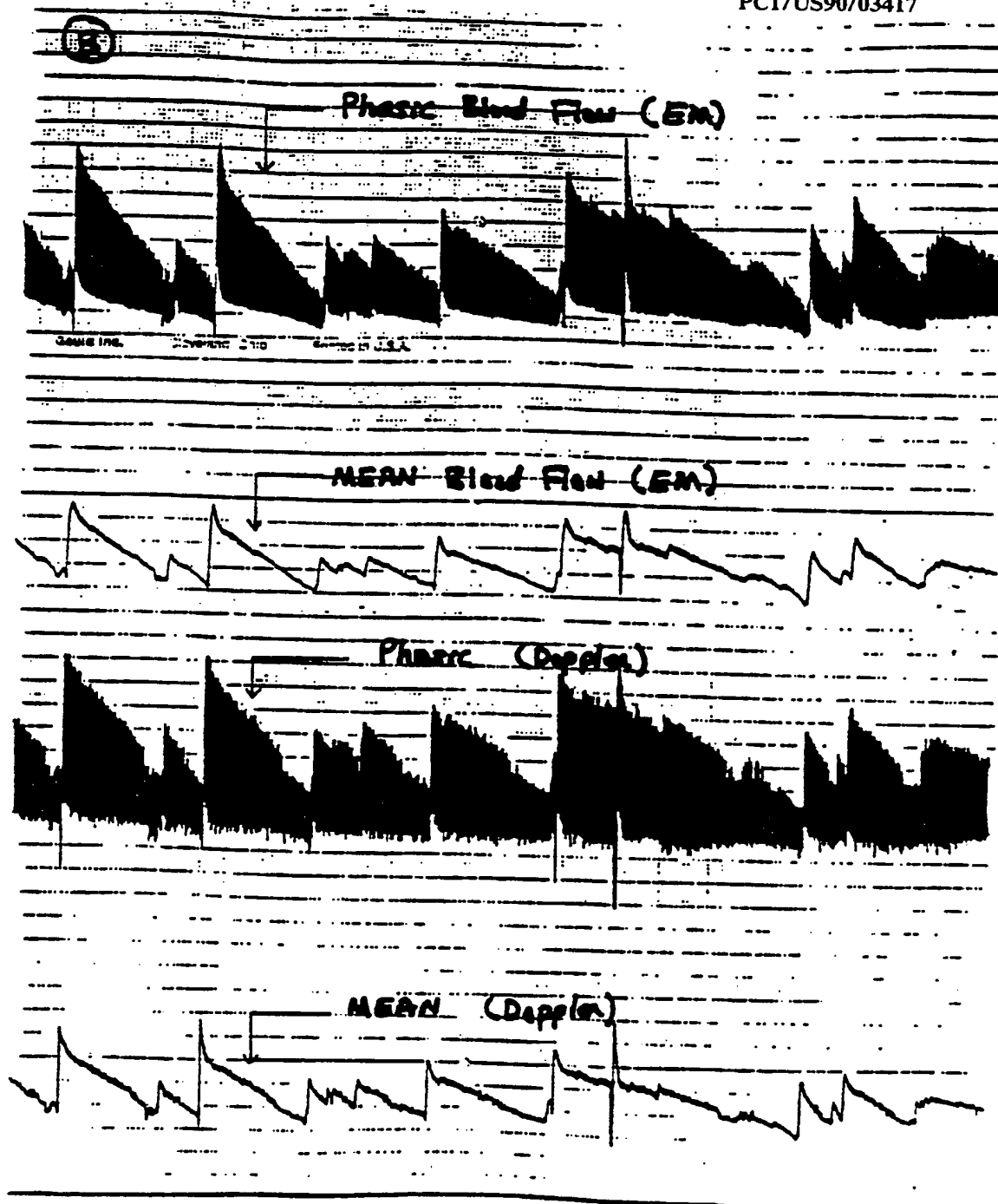


Figure 36

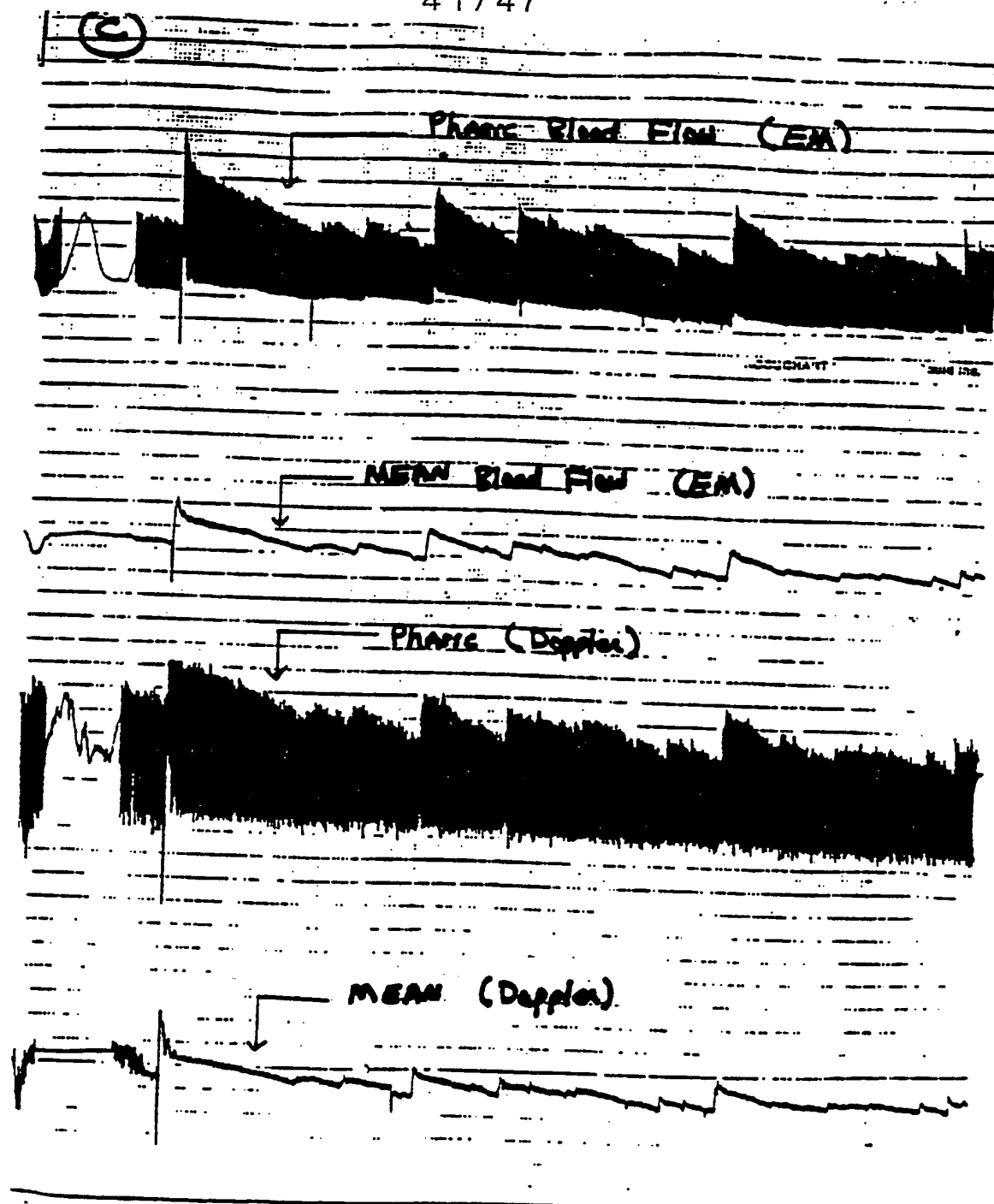


Figure 37

Barbourin(1-73) 42/47

M E A G E E C D C G S P E
AAT.TCC.ATG.GAA.GCT.GGT.GAA.GAA.TGC.GAC.TGC.GGT.TGT.CCG.GAA
^Eco R1 ^Nco 1

N P C C D A A T C K L R P G A
AAC.CCG.TGT.TGC.GAC.GCA.GCG.ACT.TGC.AAA.CTG.CGT.CCG.GGC.GCT

Q C A D G L C C D Q C R F L K
CAG.TGC.GCA.GAC.GGT.CTG.TGC.TGT.GAT.CAG.TGC.CGT.TTC.CTG.AAA

K G T V C R V A K G D W N D D
AAG.GGT.ACT.GTT.TGC.CGT.GTA.GCT.AAA.GGC.GAC.TGG.AAC.GAC.GAT

T C T G Q S A D C P R N G L Y
ACT.TGC.ACT.GGT.CAG.TCT.TGT.GAC.TGC.CCG.CGT.AAC.GGC.CTG.TAC

G
GGT.TGA.A....HindIII

Figure 38

PhoA/M-1, L⁴¹ Barbourin₍₁₋₇₃₎**Pho A:**

M K Q S T I A L A
R1.....AA.TTC.ATG.AAA.CAA.AGC.ACT.ATT.GCA.CTG.GCA

L L P L L F T P V T K A
CTC.TTA.CCG.TTA.CTG.TTC.ACC.CCT.GTG.ACC.AAA.GCC/ (splice site)

Barbourin:

M E A G E E C D C G S P E N P
ATG.GAA.GCT.GGT.GAA.GAA.TGC.GAC.TGC.GGT.TGT.CCG.GAA.AAC.CCG

C C D A A T C K L R P G A Q C
TGT.TGC.GAC.GCA.GCG.ACT.TGC.AAA.CTG.CGT.CCG.GGC.GCT.CAG.TGC

A D G L C C D Q C R F L K K G
GCA.GAC.GGT.CTG.TGC.TGT.GAT.CAG.TGC.CGT.TTC.CTG.AAA.AAG.GGT

T V C R V A K G D W N D D T C
ACT.GTT.TGC.CGT.GTA.GCT.AAA.GGC.GAC.TGG.AAC.GAC.GAT.ACT.TGC

T G Q S A D C P R N G L Y G
ACT.GGT.CAG.TCT.GCT.GAC.TGC.CCG.CGT.AAC.GGC.CTG.TAC.GGT

TGA.A....HindIII

Figure 39

PhoA/E²⁸, L⁴¹, C⁶⁴ Barbourin(28-73)

Pho A:

M K Q S T I A L A
Eco R1.....AA.TTC.ATG.AAA.CAA.AGC.ACT.ATT.GCA.CTG.GCA

L L P L L F T P V T K A
CTT.TTA.CCG.TTA.CTG.TTC.ACC.CCT.GTG.ACC.AAA.GCC/

truncated Barbourin:

E C A D G L C C D Q C R F
GAA.TGC.GCA.GAC.GGT.CTG.TGC.TGT.GAT.CAG.TGC.CGT.TTC

L K K G T V C R V A K G D W N
CTG.AAA.AAG.GGT.ACT.GTT.TGC.CGT.GTA.GCT.AAA.GGC.GAC.TGG.AAC

D D T C T G Q S C D C P R N G
GAC.GAT.ACT.TGC.ACT.GGT.CAG.TCT.TGT.GAC.TGC.CCG.CGT.AAC.GGC

L Y G
CTG.TAC.GGT.TGA.A....HindIII

Figure 40

5' oligonucleotide;

EcoRI BglII NcoI MstI

5' AAA GAA TTC CTA GAT CTC GAC GAA GCC ATG GAA TGC GCA GAC GGT CTG TGC 3'

D L D E A M E C A D G L C

D L D E A M E C A D G L C

3' oligonucleotide;

5' CGT ACC GGC CTG TAC GGT ATG GAT CCA TAA CGT TCC C 3'

R N G L Y G M D P Oe

BamHI HindIII

R N G L Y G M D P Oe

3' GCA TGG CCG GAC ATG CCA TAC CTA GGT ATT GCA AGG G 5'

Actual oligonucleotide used in the polymerase chain reaction.

Figure 41

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Polymer Junction Sequence

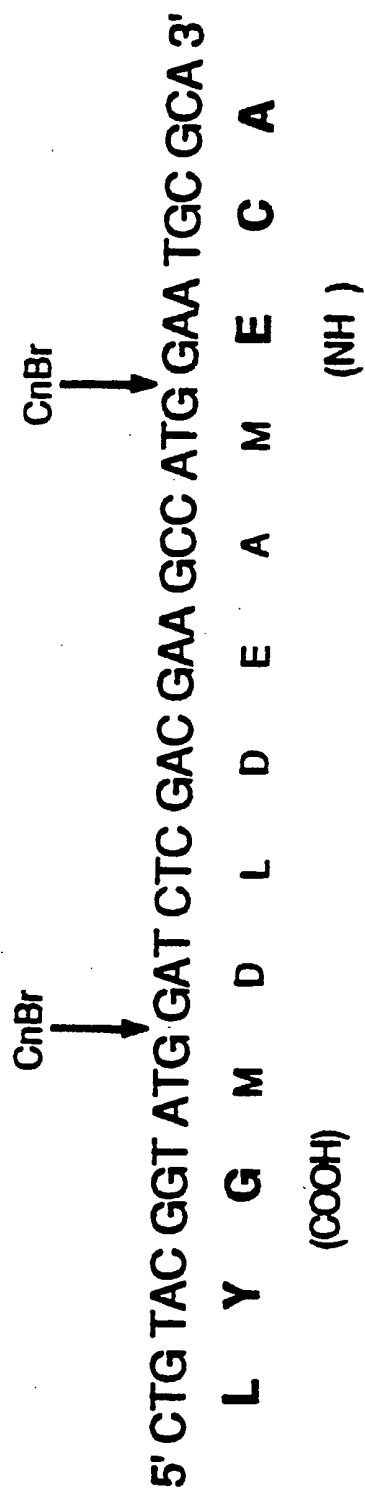
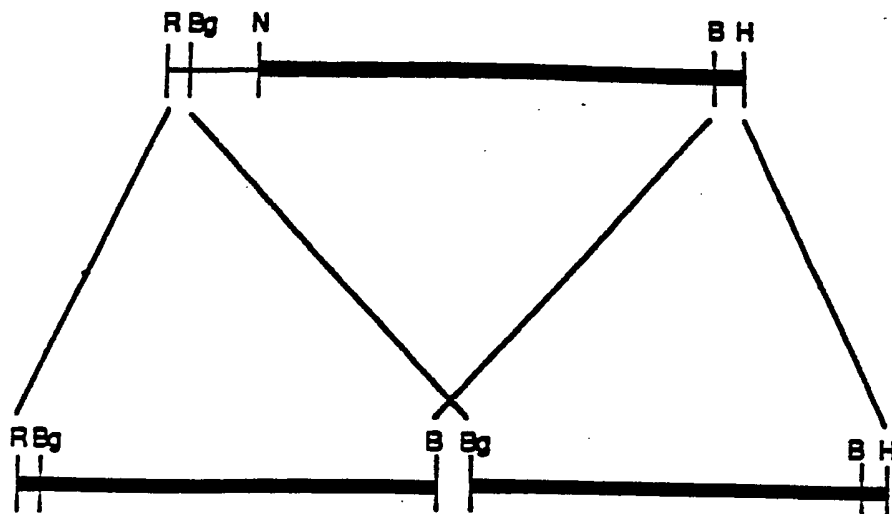


Figure 42

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A.



B.

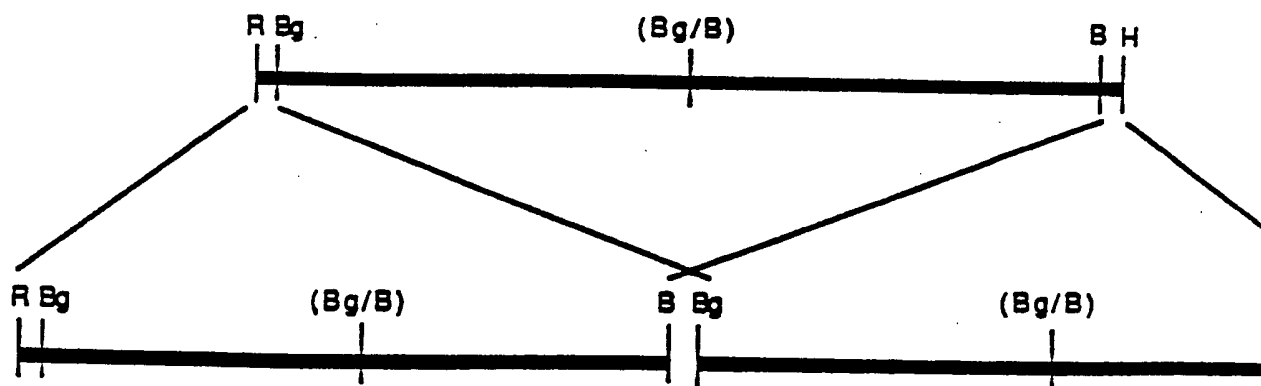


Figure 43

INTERNATIONAL SEARCH REPORT

International Application No PCT/US90/03417

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all)

According to International Patent Classification (IPC) or to both National Classification and IPC
 IPC(5): A61K 37/62; C07K 1/00, 7/00, 15/00; G01N 33/566
 U.S.CL.: 530/300, 317, 324, 345; 514/2, 11, 12, 21; 436/501

II. FIELDS SEARCHED

Minimum Documentation Searched *

Classification System

Classification Symbols

U.S. 530/300, 317, 324, 345, 856
 514/2, 11, 12, 21
 436/501

Documentation Searched other than Minimum Documentation
 to the Extent that such Documents are Included in the Fields Searched *

III. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No. *
Y	The Journal of Clinical Investigation, Volume 69, issued February 1982, R.L. Nachman et al., "Complex Formation of Platelet Membrane Glycoproteins IIb and IIIa with Fibrinogen," pages 263-269. see pages 264-265.	1-2
Y	Proceedings of the National Academy of Science USA, Volume 80, issued May 1983, J.S. Bennett et al, "Inhibition of Fibrinogen Binding to Stimulated Human Platelets by a monoclonal antibody," pages 2417-2421. see pages 2418-2419.	1-2
Y	Blood, volume 71, No. 4, issued April 1988, D.R. Phillips et al., "The Platelet Membrane Glycoprotein IIb-IIIa complex," pages 831-843. see page 842.	1-2
Y, P	EP, A, 0,341,915 (Ali et al) 15 November 1989. See the Abstract and pages 8-9.	3-12, 15, 16, 23-44

* Special categories of cited documents: *

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"A" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search *

26 September 1990

Date of Mailing of this International Search Report *

9 NOV 1990

International Searching Authority *

ISA/US

Signature of Authorized Officer **

R. KEITH BAKER, Ph.D.

III. DOCUMENTS CONSIDERED TO BE RELEVANT

(CONTINUED FROM THE SECOND SHEET)

Category *	Citation of Document, ^{1a} with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No ¹⁸
Y	Proceedings of the National Academy of Science USA, volume 87, issued April 1989, M.S. Dennis et al, "Platelet Glycoprotein IIb-IIIa Protein Antagonists from Snake Venoms: Evidence for a Family of Platelet Aggregation Inhibitors," pages 2471-2475. See the entire document.	3-12, 15, 16 & 23-44
Y, P	WO, A, 89/07609 (Lobl et al.) 24 August 1989, See page 8.	3-12, 15, 16, & 23-44
Y	The Journal of Biological Chemistry, volume 262, No. 36, issued 25 December 1987, M.D. Pierschbacher et al. "Influence of Stereochemistry of the Sequence Arg-Gly-Asp-Xaa on Binding Specificity in Cell Adhesion," pages 17, 294-17, 298. See the abstract.	3-12, 15, 16, & 23-44
A	Science, volume 238, issued 23 October 1987, E. Ruoslahti et al., "New Perspectives in Cell Adhesion: RGD and Integrins," pages 491-497. See page 492.	3-12, 15, 16, 23-44

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

V. ☐ OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE¹

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. ☐ Claim numbers because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claim numbers because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out¹, specifically:

3. ☐ Claim numbers because they are dependent claims not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI. ☐ OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING²

This International Searching Authority found multiple inventions in this international application as follows:

(See attached sheet)

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.
2. ☒ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims: 1-12, 15, 16 & 23-44 (Telephone Practice)
3. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:
4. ☐ As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remark on Protest

- ☐ The additional search fees were accompanied by applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

Attachment to PCT/ISA/210, part VI.

- Group I, claims 1-12 and 15-16, drawn to platelet aggregation inhibitors, a purification method, a method of inhibiting thrombus formation, and a method of detection, classified in classes 530, 514 and 436, subclasses 300, 2 and 501, respectively.
- Group II, claims 13 and 17, drawn to a method of treating platelet associated ischemic syndrome, classified in class 514, subclass 2.
- Group III, claims 14 and 18, drawn to a method of preventing platelet loss during extra corporeal circulation, classified in class 514, subclass 2.
- Group IV, claims 19-22, drawn to the DNA coding for the platelet aggregation inhibitor, classified in class 536, subclass 27, for example.
- Group V, claims 23-24, drawn to platelet aggregation inhibitors having Lys-Arg-Asp as the inhibitory sequence, classified in class 530, subclass 324, for example.
- Group VI, claims 25-31, 43 and 44, drawn to modified, naturally occurring platelet aggregation inhibitors, classified in class 530, subclasses 300 and 345, for example.
- Group VII, claims 32-42, drawn to modified, cyclic platelet aggregation inhibitors, classified in class 530, subclass 300 and 345, for example.
- Group VII, Claim 45, drawn to a method of treating platelet-associated ischemic syndrome using the inhibitor of Group VI, classified in class 514, subclass 2.
- Group IX, claim 46, drawn to a method of preventing platelet loss during extra corporeal circulation using the inhibitor of Group VI, classified in class 514, subclass 2.
- Group X, claim 47, drawn to a method of inhibiting metastases with cotianin, classified in class 514, subclass 12.